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Title

**COMPOSITIONS AND METHODS FOR THE THERAPY
AND DIAGNOSIS OF BREAST CANCER**

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EL615487974US**APPLICATION ELEMENTS**

See MPEP chapter 600 concerning utility patent application contents.

1. General Authorization Form & Fee Transmittal
(Submit an original and a duplicate for fee processing)2. Specification [Total Pages] **116**
(preferred arrangement set forth below)

- Descriptive Title of the Invention
- Cross References to Related Applications
- Statement Regarding Fed sponsored R & D
- Reference to Microfiche Appendix
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description
- Claim(s)
- Abstract of the Disclosure

3. Drawing(s) (35 USC 113) [Total Sheets] **25**4. Oath or Declaration [Total Pages]

- a. Newly executed (original or copy)
- b. Copy from a prior application (37 CFR 1.63(d))
(for continuation/divisional with Box 17 completed)
 - i. **DELETION OF INVENTOR(S)**
Signed statement attached deleting inventor(s) named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b)

5. Incorporation By Reference (useable if box 4b is checked) The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered to be part of the disclosure of the accompanying application and is hereby incorporated by reference therein.

17. If a CONTINUING APPLICATION, check appropriate box and supply the requisite information below and in a preliminary amendment

 Continuation Divisional Continuation-In-Part (CIP) of prior Application No.: **09/590,583**

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COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF
BREAST CANCER

CROSS REFERENCE TO RELATED APPLICATIONS

5 This application is a continuation-in-part of U.S. Patent Application No. 09/590,583, filed June 8, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/577,505, filed May 24, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/534,825, filed March 22, 2000, which is a continuation-in-part of U.S. Patent Application No. 09/429,755, filed October 28, 1999, which is a continuation-in-part
10 of U.S. Patent Application No. 09/289,198, filed April 9, 1999, which is a continuation-in-part of U.S. Patent Application No. 09/062,451, filed April 17, 1998, which is a continuation in part of U.S. Patent Application No. 08/991,789, filed December 11, 1997, which is a continuation-in-part of U.S. Patent Application No. 08/838,762, filed April 9, 1997, now abandoned, which claims priority from International Patent Application
15 No. PCT/US97/00485, filed January 10, 1997, and is a continuation-in-part of U.S. Patent Application No. 08/700,014, filed August 20, 1996, which is a continuation-in-part of U.S. Patent Application No. 08/585,392, filed January 11, 1996, now abandoned.

TECHNICAL FIELD OF THE INVENTION

The present invention relates generally to therapy and diagnosis of cancer,
20 such as breast cancer. The invention is more specifically related to polypeptides, comprising at least a portion of a breast tumor protein, and to polynucleotides encoding such polypeptides. Such polypeptides and polynucleotides are useful in pharmaceutical compositions, *e.g.*, vaccines, and other compositions for the diagnosis and treatment of breast cancer.

25 BACKGROUND OF THE INVENTION

Breast cancer is a significant health problem for women in the United States and throughout the world. Although advances have been made in detection and treatment

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of the disease, breast cancer remains the second leading cause of cancer-related deaths in women, affecting more than 180,000 women in the United States each year. For women in North America, the life-time odds of getting breast cancer are now one in eight.

No vaccine or other universally successful method for the prevention or
5 treatment of breast cancer is currently available. Management of the disease currently
relies on a combination of early diagnosis (through routine breast screening procedures)
and aggressive treatment, which may include one or more of a variety of treatments such as
surgery, radiotherapy, chemotherapy and hormone therapy. The course of treatment for a
particular breast cancer is often selected based on a variety of prognostic parameters,
10 including an analysis of specific tumor markers. *See, e.g.,* Porter-Jordan and Lippman,
Breast Cancer 8:73-100 (1994). However, the use of established markers often leads to a
result that is difficult to interpret, and the high mortality observed in breast cancer patients
indicates that improvements are needed in the treatment, diagnosis and prevention of the
disease.

15 Accordingly, there is a need in the art for improved methods for therapy and
diagnosis of breast cancer. The present invention fulfills these needs and further provides
other related advantages.

SUMMARY OF THE INVENTION

In one aspect, the present invention provides polynucleotide compositions
20 comprising a sequence selected from the group consisting of:

- (a) sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307,
313, 314, 316, 317 and 325;
- (b) complements of the sequences provided in SEQ ID NO: 1, 3-86,
142-298, 301-303, 307, 313, 314, 316, 317 and 325;
- 25 (c) sequences consisting of at least 20 contiguous residues of a sequence
provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;

- (d) sequences that hybridize to a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, under moderately stringent conditions;
- 5 (e) sequences having at least 75% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
- (f) sequences having at least 90% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325; and
- (g) degenerate variants of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

10

In one preferred embodiment, the polynucleotide compositions of the invention are expressed in at least about 20%, more preferably in at least about 30%, and most preferably in at least about 50% of breast tumors samples tested, at a level that is at least about 2-fold, preferably at least about 5-fold, and most preferably at least about 10-fold higher than that for normal tissues.

15 The present invention, in another aspect, provides polypeptide compositions comprising an amino acid sequence that is encoded by a polynucleotide sequence described above.

The present invention further provides polypeptide compositions comprising 20 an amino acid sequence selected from the group consisting of sequences recited in SEQ ID NO: 299, 300, 304-306, 308-312, 314 and 326.

In certain preferred embodiments, the polypeptides and/or polynucleotides 25 of the present invention are immunogenic, *i.e.*, they are capable of eliciting an immune response, particularly a humoral and/or cellular immune response, as further described herein.

The present invention further provides fragments, variants and/or derivatives of the disclosed polypeptide and/or polynucleotide sequences, wherein the fragments, variants and/or derivatives preferably have a level of immunogenic activity of at least about 50%, preferably at least about 70% and more preferably at least about 90% of the level of

immunogenic activity of a polypeptide sequence set forth in SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326 or a polypeptide sequence encoded by a polynucleotide sequence set forth in SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

5 The present invention further provides polynucleotides that encode a polypeptide described above, expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

Within other aspects, the present invention provides pharmaceutical compositions comprising a polypeptide or polynucleotide as described above and a 10 physiologically acceptable carrier.

Within a related aspect of the present invention, the pharmaceutical compositions, *e.g.*, vaccine compositions, are provided for prophylactic or therapeutic applications. Such compositions generally comprise an immunogenic polypeptide or polynucleotide of the invention and an immunostimulant, such as an adjuvant.

15 The present invention further provides pharmaceutical compositions that comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to a polypeptide of the present invention, or a fragment thereof; and (b) a physiologically acceptable carrier.

Within further aspects, the present invention provides pharmaceutical 20 compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) a pharmaceutically acceptable carrier or excipient. Illustrative antigen presenting cells include dendritic cells, macrophages, monocytes, fibroblasts and B cells.

Within related aspects, pharmaceutical compositions are provided that 25 comprise: (a) an antigen presenting cell that expresses a polypeptide as described above and (b) an immunostimulant.

The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins, typically in the form of pharmaceutical compositions, *e.g.*, vaccine

compositions, comprising a physiologically acceptable carrier and/or an immunostimulant. The fusions proteins may comprise multiple immunogenic polypeptides or portions/variants thereof, as described herein, and may further comprise one or more polypeptide segments for facilitating the expression, purification and/or immunogenicity of
5 the polypeptide(s).

Within further aspects, the present invention provides methods for stimulating an immune response in a patient, preferably a T cell response in a human patient, comprising administering a pharmaceutical composition described herein. The patient may be afflicted with breast cancer, in which case the methods provide treatment for
10 the disease, or patient considered at risk for such a disease may be treated prophylactically.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition as recited above. The patient may be afflicted with breast cancer, in which case the methods provide treatment for the disease, or patient considered
15 at risk for such a disease may be treated prophylactically.

The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a polypeptide of the present invention, wherein the step of contacting is performed under conditions and for a time sufficient to permit the
20 removal of cells expressing the protein from the sample.

Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating and/or expanding T cells specific for a polypeptide of the present invention, comprising contacting T cells with one or more of: (i) a polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or
25

expansion of T cells. Isolated T cell populations comprising T cells prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the development of a cancer in a patient, comprising the steps of: (a) incubating CD4⁺ and/or CD8⁺ T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of polypeptide disclosed herein; (ii) a polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expressed such a polypeptide; and (b) administering to the patient an effective amount of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

Within further aspects, the present invention provides methods for determining the presence or absence of a cancer, preferably a breast cancer, in a patient comprising: (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent is an antibody, more preferably a monoclonal antibody.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) 5 detecting in the sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one oligonucleotide 10 primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

15 In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b) using a 20 biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

25 Within further aspects, the present invention provides antibodies, such as monoclonal antibodies, that bind to a polypeptide as described above, as well as diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references

disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the differential display PCR products, separated by gel electrophoresis, obtained from cDNA prepared from normal breast tissue (lanes 1 and 2) and from cDNA prepared from breast tumor tissue from the same patient (lanes 3 and 4). The arrow indicates the band corresponding to B18Ag1.

Figure 2 is a northern blot comparing the level of B18Ag1 mRNA in breast tumor tissue (lane 1) with the level in normal breast tissue.

Figure 3 shows the level of B18Ag1 mRNA in breast tumor tissue compared to that in various normal and non-breast tumor tissues as determined by RNase protection assays.

Figure 4 is a genomic clone map showing the location of additional retroviral sequences obtained from ends of XbaI restriction digests (provided in SEQ ID NO:3 - SEQ ID NO:10) relative to B18Ag1.

Figures 5A and 5B show the sequencing strategy, genomic organization and predicted open reading frame for the retroviral element containing B18Ag1.

Figure 6 shows the nucleotide sequence of the representative breast tumor-specific cDNA B18Ag1.

Figure 7 shows the nucleotide sequence of the representative breast tumor-specific cDNA B17Ag1.

Figure 8 shows the nucleotide sequence of the representative breast tumor-specific cDNA B17Ag2.

Figure 9 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag2a.

Figure 10 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag1b.

Figure 11 shows the nucleotide sequence of the representative breast tumor-specific cDNA B13Ag1a.

Figure 12 shows the nucleotide sequence of the representative breast tumor-specific cDNA B11Ag1.

5 Figure 13 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA3c.

Figure 14 shows the nucleotide sequence of the representative breast tumor-specific cDNA B9CG1.

10 Figure 15 shows the nucleotide sequence of the representative breast tumor-specific cDNA B9CG3.

Figure 16 shows the nucleotide sequence of the representative breast tumor-specific cDNA B2CA2.

Figure 17 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA1.

15 Figure 18 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA2.

Figure 19 shows the nucleotide sequence of the representative breast tumor-specific cDNA B3CA3.

20 Figure 20 shows the nucleotide sequence of the representative breast tumor-specific cDNA B4CA1.

Figure 21A depicts RT-PCR analysis of breast tumor genes in breast tumor tissues (lanes 1-8) and normal breast tissues (lanes 9-13) and H₂O (lane 14).

25 Figure 21B depicts RT-PCR analysis of breast tumor genes in prostate tumors (lane 1, 2), colon tumors (lane 3), lung tumor (lane 4), normal prostate (lane 5), normal colon (lane 6), normal kidney (lane 7), normal liver (lane 8), normal lung (lane 9), normal ovary (lanes 10, 18), normal pancreases (lanes 11, 12), normal skeletal muscle (lane 13), normal skin (lane 14), normal stomach (lane 15), normal testes (lane 16), normal small intestine (lane 17), HBL-100 (lane 19), MCF-12A (lane 20), breast tumors (lanes 21-23), H₂O (lane 24), and colon tumor (lane 25).

Figure 22 shows the recognition of a B11Ag1 peptide (referred to as B11-8) by an anti-B11-8 CTL line.

Figure 23 shows the recognition of a cell line transduced with the antigen B11Ag1 by the B11-8 specific clone A1.

5 Figure 24 shows recognition of a lung adenocarcinoma line (LT-140-22) and a breast adenocarcinoma line (CAMA-1) by the B11-8 specific clone A1.

DETAILED DESCRIPTION OF THE INVENTION

The present invention is directed generally to compositions and their use in the therapy and diagnosis of cancer, particularly breast cancer. As described further below, 10 illustrative compositions of the present invention include, but are not restricted to, polypeptides, particularly immunogenic polypeptides, polynucleotides encoding such polypeptides, antibodies and other binding agents, antigen presenting cells (APCs) and immune system cells (*e.g.*, T cells).

The practice of the present invention will employ, unless indicated 15 specifically to the contrary, conventional methods of virology, immunology, microbiology, molecular biology and recombinant DNA techniques within the skill of the art, many of which are described below for the purpose of illustration. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, et al. *Molecular Cloning: A Laboratory Manual* (2nd Edition, 1989); Maniatis et al. *Molecular Cloning: A Laboratory Manual* (1982); 20 *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., 1984); *Nucleic Acid Hybridization* (B. Hames & S. Higgins, eds., 1985); *Transcription and Translation* (B. Hames & S. Higgins, eds., 1984); *Animal Cell Culture* (R. Freshney, ed., 1986); Perbal, *A Practical Guide to Molecular Cloning* (1984).

All publications, patents and patent applications cited herein, whether supra 25 or infra, are hereby incorporated by reference in their entirety.

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates otherwise.

Polypeptide Compositions

As used herein, the term "polypeptide" is used in its conventional meaning, *i.e.*, as a sequence of amino acids. The polypeptides are not limited to a specific length of the product; thus, peptides, oligopeptides, and proteins are included within the definition of polypeptide, and such terms may be used interchangeably herein unless specifically indicated otherwise. This term also does not refer to or exclude post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like, as well as other modifications known in the art, both naturally occurring and non-naturally occurring. A polypeptide may be an entire protein, or a subsequence thereof. Particular polypeptides of interest in the context of this invention are amino acid subsequences comprising epitopes, *i.e.*, antigenic determinants substantially responsible for the immunogenic properties of a polypeptide and being capable of evoking an immune response.

Particularly illustrative polypeptides of the present invention comprise those encoded by a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, or a sequence that hybridizes under moderately stringent conditions, or, alternatively, under highly stringent conditions, to a polynucleotide sequence set forth in any one of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325. Certain other illustrative polypeptides of the invention comprise amino acid sequences as set forth in any one of SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326.

The polypeptides of the present invention are sometimes herein referred to as breast tumor proteins or breast tumor polypeptides, as an indication that their identification has been based at least in part upon their increased levels of expression in breast tumor samples. Thus, a "breast tumor polypeptide" or "breast tumor protein," refers generally to a polypeptide sequence of the present invention, or a polynucleotide sequence encoding such a polypeptide, that is expressed in a substantial proportion of breast tumor samples, for example preferably greater than about 20%, more preferably greater than about 30%, and most preferably greater than about 50% or more of breast

tumor samples tested, at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in normal tissues, as determined using a representative assay provided herein. A breast tumor polypeptide sequence of the invention, based upon its increased level of expression in tumor cells, has particular utility both as a diagnostic 5 marker as well as a therapeutic target, as further described below.

In certain preferred embodiments, the polypeptides of the invention are immunogenic, *i.e.*, they react detectably within an immunoassay (such as an ELISA or T-cell stimulation assay) with antisera and/or T-cells from a patient with breast cancer. Screening for immunogenic activity can be performed using techniques well known to the 10 skilled artisan. For example, such screens can be performed using methods such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In one illustrative example, a polypeptide may be immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies 15 detected using, for example, ¹²⁵I-labeled Protein A.

As would be recognized by the skilled artisan, immunogenic portions of the polypeptides disclosed herein are also encompassed by the present invention. An “immunogenic portion,” as used herein, is a fragment of an immunogenic polypeptide of the invention that itself is immunologically reactive (*i.e.*, specifically binds) with the B-cells and/or T-cell surface antigen receptors that recognize the polypeptide. Immunogenic 20 portions may generally be identified using well known techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used 25 herein, antisera and antibodies are “antigen-specific” if they specifically bind to an antigen (*i.e.*, they react with the protein in an ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well-known techniques.

In one preferred embodiment, an immunogenic portion of a polypeptide of the present invention is a portion that reacts with antisera and/or T-cells at a level that is not substantially less than the reactivity of the full-length polypeptide (*e.g.*, in an ELISA and/or T-cell reactivity assay). Preferably, the level of immunogenic activity of the immunogenic portion is at least about 50%, preferably at least about 70% and most preferably greater than about 90% of the immunogenicity for the full-length polypeptide. In some instances, preferred immunogenic portions will be identified that have a level of immunogenic activity greater than that of the corresponding full-length polypeptide, *e.g.*, having greater than about 100% or 150% or more immunogenic activity.

10 In certain other embodiments, illustrative immunogenic portions may include peptides in which an N-terminal leader sequence and/or transmembrane domain have been deleted. Other illustrative immunogenic portions will contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

15 In another embodiment, a polypeptide composition of the invention may also comprise one or more polypeptides that are immunologically reactive with T cells and/or antibodies generated against a polypeptide of the invention, particularly a polypeptide having an amino acid sequence disclosed herein, or to an immunogenic fragment or variant thereof.

20 In another embodiment of the invention, polypeptides are provided that comprise one or more polypeptides that are capable of eliciting T cells and/or antibodies that are immunologically reactive with one or more polypeptides described herein, or one or more polypeptides encoded by contiguous nucleic acid sequences contained in the polynucleotide sequences disclosed herein, or immunogenic fragments or variants thereof, 25 or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency.

The present invention, in another aspect, provides polypeptide fragments comprising at least about 5, 10, 15, 20, 25, 50, or 100 contiguous amino acids, or more, including all intermediate lengths, of a polypeptide compositions set forth herein, such as

those set forth in SEQ ID NOs: 299, 300, 304-306, 308-312, 314 and 326, or those encoded by a polynucleotide sequence set forth in a sequence of SEQ ID NOs: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.

In another aspect, the present invention provides variants of the polypeptide compositions described herein. Polypeptide variants generally encompassed by the present invention will typically exhibit at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% or more identity (determined as described below), along its length, to a polypeptide sequences set forth herein.

In one preferred embodiment, the polypeptide fragments and variants provide by the present invention are immunologically reactive with an antibody and/or T-cell that reacts with a full-length polypeptide specifically set for the herein.

In another preferred embodiment, the polypeptide fragments and variants provided by the present invention exhibit a level of immunogenic activity of at least about 50%, preferably at least about 70%, and most preferably at least about 90% or more of that exhibited by a full-length polypeptide sequence specifically set forth herein.

A polypeptide "variant," as the term is used herein, is a polypeptide that typically differs from a polypeptide specifically disclosed herein in one or more substitutions, deletions, additions and/or insertions. Such variants may be naturally occurring or may be synthetically generated, for example, by modifying one or more of the above polypeptide sequences of the invention and evaluating their immunogenic activity as described herein and/or using any of a number of techniques well known in the art.

For example, certain illustrative variants of the polypeptides of the invention include those in which one or more portions, such as an N-terminal leader sequence or transmembrane domain, have been removed. Other illustrative variants include variants in which a small portion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

In many instances, a variant will contain conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would

expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. As described above, modifications may be made in the structure of the polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics,
5 *e.g.*, with immunogenic characteristics. When it is desired to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, immunogenic variant or portion of a polypeptide of the invention, one skilled in the art will typically change one or more of the codons of the encoding DNA sequence according to Table 1.

For example, certain amino acids may be substituted for other amino acids
10 in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and
15 nevertheless obtain a protein with like properties. It is thus contemplated that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

TABLE 1

Amino Acids		Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU		
Cysteine	Cys	C	UGC	UGU				
Aspartic acid	Asp	D	GAC	GAU				
Glutamic acid	Glu	E	GAA	GAG				
Phenylalanine	Phe	F	UUC	UUU				
Glycine	Gly	G	GGA	GGC	GGG	GGU		
Histidine	His	H	CAC	CAU				
Isoleucine	Ile	I	AUA	AUC	AUU			
Lysine	Lys	K	AAA	AAG				
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU
Methionine	Met	M	AUG					
Asparagine	Asn	N	AAC	AAU				
Proline	Pro	P	CCA	CCC	CCG	CCU		
Glutamine	Gln	Q	CAA	CAG				
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU
Threonine	Thr	T	ACA	ACC	ACG	ACU		
Valine	Val	V	GU	GUC	GUG	GUU		
Tryptophan	Trp	W	UGG					
Tyrosine	Tyr	Y	UAC	UAU				

In making such changes, the hydropathic index of amino acids may be
5 considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes,

substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8);
5 glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5); glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with
10 similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101
15 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate ($+3.0 \pm 1$); glutamate ($+3.0 \pm 1$); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (-0.4); proline (-0.5 ± 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0); methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3); phenylalanine (-2.5); tryptophan (-3.4). It is understood that an amino acid can be substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within ± 2 is preferred, those within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than 10 phosphodiester linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine and wybutoxine, as well as acetyl-, methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

Amino acid substitutions may further be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of 15 the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may represent conservative 20 changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes. In a preferred embodiment, variant 25 polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer. Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure and hydropathic nature of the polypeptide.

As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other

sequence for ease of synthesis, purification or identification of the polypeptide (*e.g.*, poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

When comparing polypeptide sequences, two sequences are said to be
 5 “identical” if the sequence of amino acids in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A “comparison window” as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to
 10 about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment
 15 schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) *Atlas of Protein Sequence and Structure*, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183,
 20 Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J.
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Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl.*

Acad. Sci. USA 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides and polypeptides of the invention. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment.

In one preferred approach, the “percentage of sequence identity” is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Within other illustrative embodiments, a polypeptide may be a fusion polypeptide that comprises multiple polypeptides as described herein, or that comprises at least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native recombinant protein. Certain preferred fusion partners are both immunological and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the polypeptide or to enable the polypeptide to be targeted to desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the polypeptide.

Fusion polypeptides may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion polypeptide is expressed as a recombinant polypeptide, allowing the production of increased levels, relative to a non-fused polypeptide, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion polypeptide that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion polypeptide using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes.

Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea et al., *Gene* 40:39-46, 1985; Murphy et al., *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 5 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

The ligated DNA sequences are operably linked to suitable transcriptional or 10 translational regulatory elements. The regulatory elements responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the second polypeptide.

The fusion polypeptide can comprise a polypeptide as described herein 15 together with an unrelated immunogenic protein, such as an immunogenic protein capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (see, for example, Stoute et al. *New Engl. J. Med.*, 336:86-91, 1997).

In one preferred embodiment, the immunological fusion partner is derived 20 from a *Mycobacterium* sp., such as a *Mycobacterium tuberculosis*-derived Ra12 fragment. Ra12 compositions and methods for their use in enhancing the expression and/or 25 immunogenicity of heterologous polynucleotide/polypeptide sequences is described in U.S. Patent Application 60/158,585, the disclosure of which is incorporated herein by reference in its entirety. Briefly, Ra12 refers to a polynucleotide region that is a subsequence of a *Mycobacterium tuberculosis* MTB32A nucleic acid. MTB32A is a serine protease of 32 KD molecular weight encoded by a gene in virulent and avirulent strains of *M. tuberculosis*. The nucleotide sequence and amino acid sequence of MTB32A have been described (for example, U.S. Patent Application 60/158,585; see also, Skeiky et al., *Infection and Immun.* (1999) 67:3998-4007, incorporated herein by reference). C-terminal fragments of the MTB32A coding sequence express at high levels and remain as a soluble

polypeptides throughout the purification process. Moreover, Ra12 may enhance the immunogenicity of heterologous immunogenic polypeptides with which it is fused. One preferred Ra12 fusion polypeptide comprises a 14 KD C-terminal fragment corresponding to amino acid residues 192 to 323 of MTB32A. Other preferred Ra12 polynucleotides 5 generally comprise at least about 15 consecutive nucleotides, at least about 30 nucleotides, at least about 60 nucleotides, at least about 100 nucleotides, at least about 200 nucleotides, or at least about 300 nucleotides that encode a portion of a Ra12 polypeptide. Ra12 polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a Ra12 polypeptide or a portion thereof) or may comprise a variant of such a 10 sequence. Ra12 polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions such that the biological activity of the encoded fusion polypeptide is not substantially diminished, relative to a fusion polypeptide comprising a native Ra12 polypeptide. Variants preferably exhibit at least about 70% identity, more 15 preferably at least about 80% identity and most preferably at least about 90% identity to a polynucleotide sequence that encodes a native Ra12 polypeptide or a portion thereof.

Within other preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenzae* B (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (*e.g.*, the first N-terminal 100-110 amino acids), and a protein D 20 derivative may be lipidated. Within certain preferred embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells. Other fusion partners include the non-structural 25 protein from influenzae virus, NS1 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known

as amidase LYTA (encoded by the LytA gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This property has been exploited for the 5 development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (*see Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of LYTA may be incorporated into a fusion polypeptide. A repeat portion is found in the C-terminal region starting at residue 178. A particularly 10 preferred repeat portion incorporates residues 188-305.

Yet another illustrative embodiment involves fusion polypeptides, and the polynucleotides encoding them, wherein the fusion partner comprises a targeting signal capable of directing a polypeptide to the endosomal/lysosomal compartment, as described in U.S. Patent No. 5,633,234. An immunogenic polypeptide of the invention, when fused 15 with this targeting signal, will associate more efficiently with MHC class II molecules and thereby provide enhanced in vivo stimulation of CD4⁺ T-cells specific for the polypeptide.

Polypeptides of the invention are prepared using any of a variety of well known synthetic and/or recombinant techniques, the latter of which are further described below. Polypeptides, portions and other variants generally less than about 150 amino acids 20 can be generated by synthetic means, using techniques well known to those of ordinary skill in the art. In one illustrative example, such polypeptides are synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. *See Merrifield, J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated 25 synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and may be operated according to the manufacturer's instructions.

In general, polypeptide compositions (including fusion polypeptides) of the invention are isolated. An "isolated" polypeptide is one that is removed from its original

environment. For example, a naturally-occurring protein or polypeptide is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are also purified, *e.g.*, are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure.

5 Polynucleotide Compositions

The present invention, in other aspects, provides polynucleotide compositions. The terms "DNA" and "polynucleotide" are used essentially interchangeably herein to refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. "Isolated," as used herein, means that a 10 polynucleotide is substantially away from other coding sequences, and that the DNA molecule does not contain large portions of unrelated coding DNA, such as large chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA molecule as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

15 As will be understood by those skilled in the art, the polynucleotide compositions of this invention can include genomic sequences, extra-genomic and plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

20 As will be also recognized by the skilled artisan, polynucleotides of the invention may be single-stranded (coding or antisense) or double-stranded, and may be DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules may include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or 25 non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules and/or support materials.

Polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a polypeptide/protein of the invention or a portion thereof) or may comprise a sequence that encodes a variant or derivative, preferably and immunogenic variant or derivative, of such a sequence.

5 Therefore, according to another aspect of the present invention, polynucleotide compositions are provided that comprise some or all of a polynucleotide sequence set forth in any one of SEQ ID NOS: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, complements of a polynucleotide sequence set forth in any one of SEQ ID NOS: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, and degenerate 10 variants of a polynucleotide sequence set forth in any one of SEQ ID NOS: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325. In certain preferred embodiments, the polynucleotide sequences set forth herein encode immunogenic polypeptides, as described above.

In other related embodiments, the present invention provides polynucleotide 15 variants having substantial identity to the sequences disclosed herein in SEQ ID NOS: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, for example those comprising at least 70% sequence identity, preferably at least 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence identity compared to a polynucleotide sequence of this 20 invention using the methods described herein, (*e.g.*, BLAST analysis using standard parameters, as described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like.

Typically, polynucleotide variants will contain one or more substitutions, 25 additions, deletions and/or insertions, preferably such that the immunogenicity of the polypeptide encoded by the variant polynucleotide is not substantially diminished relative to a polypeptide encoded by a polynucleotide sequence specifically set forth herein). The term “variants” should also be understood to encompass homologous genes of xenogenic origin.

In additional embodiments, the present invention provides polynucleotide fragments comprising various lengths of contiguous stretches of sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at least about 10, 15, 20, 30, 5 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate lengths", in this context, means any length between the quoted values, such as 16, 17, 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers 10 through 200-500; 500-1,000, and the like.

In another embodiment of the invention, polynucleotide compositions are provided that are capable of hybridizing under moderate to high stringency conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. 15 For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50°C-60°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS. One skilled in the art will 20 understand that the stringency of hybridization can be readily manipulated, such as by altering the salt content of the hybridization solution and/or the temperature at which the hybridization is performed. For example, in another embodiment, suitable highly stringent hybridization conditions include those described above, with the exception that the temperature of hybridization is increased, *e.g.*, to 60-65°C or 65-70°C.

25 In certain preferred embodiments, the polynucleotides described above, *e.g.*, polynucleotide variants, fragments and hybridizing sequences, encode polypeptides that are immunologically cross-reactive with a polypeptide sequence specifically set forth herein. In other preferred embodiments, such polynucleotides encode polypeptides that have a level of immunogenic activity of at least about 50%, preferably at least about 70%, and

more preferably at least about 90% of that for a polypeptide sequence specifically set forth herein.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, illustrative 5 polynucleotide segments with total lengths of about 10,000, about 5000, about 3000, about 10 2,000, about 1,000, about 500, about 200, about 100, about 50 base pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many 15 implementations of this invention.

When comparing polynucleotide sequences, two sequences are said to be 15 “identical” if the sequence of nucleotides in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A “comparison window” as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to 20 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment 25 schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183,

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Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 10 48:443, by the search for similarity methods of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides of the invention. Software for performing 20 BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the 25 quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62

scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments, (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

Preferably, the “percentage of sequence identity” is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid bases occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

It will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention. Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

Therefore, in another embodiment of the invention, a mutagenesis approach, such as site-specific mutagenesis, is employed for the preparation of immunogenic variants and/or derivatives of the polypeptides described herein. By this approach, specific modifications in a polypeptide sequence can be made through mutagenesis of the underlying polynucleotides that encode them. These techniques provides a straightforward

approach to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the polynucleotide.

Site-specific mutagenesis allows the production of mutants through the use
5 of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Mutations may be employed in a selected polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the
10 properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or more properties of the encoded polypeptide, such as the immunogenicity of a polypeptide vaccine. The
15 techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example, site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so in length is employed, with about 5 to about 10 residues on both sides of the junction of the
20 sequence being altered.

As will be appreciated by those of skill in the art, site-specific mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially-available and their use is
25 generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double-stranded

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vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to 5 complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

10 The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic 15 agents, such as hydroxylamine, to obtain sequence variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

As used herein, the term “oligonucleotide directed mutagenesis procedure”
20 refers to template-dependent processes and vector-mediated propagation which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable signal, such as amplification. As used herein, the term “oligonucleotide directed mutagenesis procedure” is intended to refer to a process that involves the template-dependent extension of a primer
25 molecule. The term template dependent process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known rules of complementary base pairing (see, for example, Watson, 1987). Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the

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recovery of the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

In another approach for the production of polypeptide variants of the present invention, recursive sequence recombination, as described in U.S. Patent No. 5,837,458, may be employed. In this approach, iterative cycles of recombination and screening or selection are performed to "evolve" individual polynucleotide variants of the invention having, for example, enhanced immunogenic activity.

In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic acid hybridization. As such, it is contemplated that nucleic acid segments that comprise a sequence region of at least about 15 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000 (including all intermediate lengths) and even up to full length sequences will also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned, such as the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately

depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the
5 length complementary sequences one wishes to detect.

The use of a hybridization probe of about 15-25 nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 15 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and
10 thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 25 contiguous nucleotides, or even longer where desired.

Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequences set forth herein,
15 or to any continuous portion of the sequences, from about 15-25 nucleotides in length up to and including the full length sequence, that one wishes to utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

Small polynucleotide segments or fragments may be readily prepared by, for
20 example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCRTM technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA
25 techniques generally known to those of skill in the art of molecular biology.

The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of

PCT/US2012/050000

probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, e.g., one will select relatively low salt and/or high temperature conditions, such as provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about 5 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating related sequences.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template, less 10 stringent (reduced stringency) hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any 15 case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

According to another embodiment of the present invention, polynucleotide 20 compositions comprising antisense oligonucleotides are provided. Antisense oligonucleotides have been demonstrated to be effective and targeted inhibitors of protein synthesis, and, consequently, provide a therapeutic approach by which a disease can be treated by inhibiting the synthesis of proteins that contribute to the disease. The efficacy of 25 antisense oligonucleotides for inhibiting protein synthesis is well established. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829). Further, examples of antisense inhibition have been demonstrated with the nuclear protein cyclin, the multiple drug resistance gene

(MDG1), ICAM-1, E-selectin, STK-1, striatal GABA_A receptor and human EGF (Jaskulska *et al.*, Science. 1988 Jun 10;240(4858):1544-6; Vasantha Kumar and Ahmed, Cancer Commun. 1989;1(4):225-32; Peris *et al.*, Brain Res Mol Brain Res. 1998 Jun 15;57(2):310-20; U. S. Patent 5,801,154; U.S. Patent 5,789,573; U. S. Patent 5,718,709 and U.S. Patent 5,610,288). Antisense constructs have also been described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683).

Therefore, in certain embodiments, the present invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary, and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein. Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence and determination of secondary structure, T_m, binding energy, and relative stability. Antisense compositions may be selected based upon their relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell. Highly preferred target regions of the mRNA, are those which are at or near the AUG translation initiation codon, and those sequences which are substantially complementary to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations can be performed, for example, using v.4 of the OLIGO primer analysis software and/or the BLASTN 2.0.5 algorithm software (Altschul *et al.*, Nucleic Acids Res. 1997, 25(17):3389-402).

The use of an antisense delivery method employing a short peptide vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, Nucleic Acids Res. 1997 Jul 15;25(14):2730-6). It has been demonstrated that several molecules of the MPG peptide coat the antisense oligonucleotides and can be delivered into cultured mammalian cells in less than 1 hour with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane.

According to another embodiment of the invention, the polynucleotide compositions described herein are used in the design and preparation of ribozyme molecules for inhibiting expression of the tumor polypeptides and proteins of the present invention in tumor cells. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, Proc Natl Acad Sci U S A. 1987 Dec;84(24):8788-92; Forster and Symons, Cell. 1987 Apr 24;49(2):211-20). For example, a large number of ribozymes accelerate phosphoester transfer reactions with a high degree of specificity, often cleaving only one of several phosphoesters in an oligonucleotide substrate (Cech *et al.*, Cell. 1981 Dec;27(3 Pt 2):487-96; Michel and Westhof, J Mol Biol. 1990 Dec 5;216(3):585-610; Reinhold-Hurek and Shub, Nature. 1992 May 14;357(6374):173-6). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide sequence ("IGS") of the ribozyme prior to chemical reaction.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general, enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic

nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it 5 is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its translation) since the concentration of ribozyme necessary 10 to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding to the target RNA, but also on the mechanism of target RNA 15 cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woolf *et al.*, Proc Natl Acad Sci U S A. 1992 Aug 15;89(16):7305-9). Thus, the specificity of action of a ribozyme is greater than that of an antisense oligonucleotide binding the same RNA site.

20 The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis δ virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* Nucleic Acids Res. 1992 Sep 11;20(17):4559-65. Examples of hairpin motifs are described by Hampel *et al.* (Eur. Pat. Appl. Publ. No. EP 0360257), 25 Hampel and Tritz, Biochemistry 1989 Jun 13;28(12):4929-33; Hampel *et al.*, Nucleic Acids Res. 1990 Jan 25;18(2):299-304 and U. S. Patent 5,631,359. An example of the hepatitis δ virus motif is described by Perrotta and Been, Biochemistry. 1992 Dec 1;31(47):11843-52; an example of the RNaseP motif is described by Guerrier-Takada *et al.*, Cell. 1983 Dec;35(3 Pt 2):849-57; Neurospora VS RNA ribozyme motif is described by

Collins (Saville and Collins, Cell. 1990 May 18;61(4):685-96; Saville and Collins, Proc Natl Acad Sci U S A. 1991 Oct 1;88(19):8826-30; Collins and Olive, Biochemistry. 1993 Mar 23;32(11):2795-9); and an example of the Group I intron is described in (U. S. Patent 4,987,071). All that is important in an enzymatic nucleic acid molecule of this invention is
 5 that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO 10 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

15 Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see e.g., Int. Pat. Appl. Publ. No. WO 92/07065; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688,
 20 which describe various chemical modifications that can be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the
 25 general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be directly delivered *ex*

vivo to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, subcutaneous or joint injection, aerosol inhalation, 5 oral (tablet or pill form), topical, systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

Another means of accumulating high concentrations of a ribozyme(s) within 10 cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the nature of the gene 15 regulatory sequences (enhancers, silencers, etc.) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells. Ribozymes expressed from such promoters have been shown to function in mammalian cells. Such transcription units can be incorporated into a variety of vectors for introduction into mammalian cells, including but 20 not restricted to, plasmid DNA vectors, viral DNA vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

In another embodiment of the invention, peptide nucleic acids (PNAs) 25 compositions are provided. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, Antisense Nucleic Acid Drug Dev. 1997 7(4) 431-37). PNA is able to be utilized in a number methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided

by Corey (*Trends Biotechnol* 1997 Jun;15(6):224-9). As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-specific mRNA, and thereby alter the level of ACE activity 5 in a host cell to which such PNA compositions have been administered.

PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, *Science* 1991 Dec 6;254(5037):1497-500; Hanvey *et al.*, *Science*. 1992 Nov 27;258(5087):1481-5; Hyrup and Nielsen, *Bioorg Med Chem.* 1996 Jan;4(1):5-23). This chemistry has three important consequences: 10 firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral molecules; secondly, PNAs are achiral, which avoids the need to develop a stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc or Fmoc protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used.

15 PNA monomers or ready-made oligomers are commercially available from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, *Bioorg Med Chem.* 1995 Apr;3(4):437-45). The manual protocol lends itself to the production of chemically modified PNAs or the simultaneous synthesis of families of closely related 20 PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this difficulty, it is 25 suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography, providing yields and purity of product similar to those observed during the synthesis of peptides.

Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The ease with which PNAs can be modified facilitates optimization for better solubility or for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (for example, Norton *et al.*, *Bioorg Med Chem.* 1995 Apr;3(4):437-45; Petersen *et al.*, *J Pept Sci.* 1995 May-Jun;1(3):175-83; Orum *et al.*, *Biotechniques.* 1995 Sep;19(3):472-80; Footer *et al.*, *Biochemistry.* 1996 Aug 20;35(33):10673-9; Griffith *et al.*, *Nucleic Acids Res.* 1995 Aug 11;23(15):3003-8; Pardridge *et al.*, *Proc Natl Acad Sci U S A.* 1995 Jun 6;92(12):5592-6; Boffa *et al.*, *Proc Natl Acad Sci U S A.* 1995 Mar 14;92(6):1901-5; Gambacorti-Passerini *et al.*, *Blood.* 1996 Aug 15;88(4):1411-7; Armitage *et al.*, *Proc Natl Acad Sci U S A.* 1997 Nov 11;94(23):12320-5; Seeger *et al.*, *Biotechniques.* 1997 Sep;23(3):512-7). U.S. Patent No. 5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to therapeutics.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (*Anal Chem.* 1993 Dec 15;65(24):3545-9) and Jensen *et al.* (*Biochemistry.* 1997 Apr 22;36(16):5072-7). Rose uses capillary gel electrophoresis to determine binding of PNAs to their complementary oligonucleotide, measuring the relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BIACore™ technology.

Other applications of PNAs that have been described and will be apparent to the skilled artisan include use in DNA strand invasion, antisense inhibition, mutational analysis, enhancers of transcription, nucleic acid purification, isolation of transcriptionally active genes, blocking of transcription factor binding, genome cleavage, biosensors, *in situ* hybridization, and the like.

Polynucleotide Identification, Characterization and Expression

Polynucleotides compositions of the present invention may be identified, prepared and/or manipulated using any of a variety of well established techniques (see generally, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989, and other like references). For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for tumor-associated expression (*i.e.*, expression that is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using the microarray technology of Affymetrix, Inc. (Santa Clara, CA) according to the manufacturer's instructions (and essentially as described by Schena et al., *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller et al., *Proc. Natl. Acad. Sci. USA* 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as tumor cells.

Many template dependent processes are available to amplify a target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCR™) which is described in detail in U.S. Patent Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCR™, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present in a sample, the primers will bind to the target and the polymerase will cause the primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCR™ amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Any of a number of other template dependent processes, many of which are variations of the PCR™ amplification technique, are readily known and available in the art. Illustratively, some such methods include the ligase chain reaction (referred to as LCR), described, for example, in Eur. Pat. Appl. Publ. No. 320,308 and U.S. Patent No. 5 4,883,750; Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880; Strand Displacement Amplification (SDA) and Repair Chain Reaction (RCR). Still other amplification methods are described in Great Britain Pat. Appl. No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025. Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (PCT Intl. Pat. Appl. Publ. No. 10 WO 88/10315), including nucleic acid sequence based amplification (NASBA) and 3SR. Eur. Pat. Appl. Publ. No. 329,822 describes a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA). PCT Intl. Pat. Appl. Publ. No. WO 89/06700 describes a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence 15 to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. Other amplification methods such as "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) are also well-known to those of skill in the art.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (e.g., a tumor cDNA library) using 20 well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (e.g., by 25 nick-translation or end-labeling with ³²P) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (see Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor

Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences 5 may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

10 Alternatively, amplification techniques, such as those described above, can be useful for obtaining a full length coding sequence from a partial cDNA sequence. One such amplification technique is inverse PCR (*see* Triglia et al., *Nucl. Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template 15 for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known region. A variation on this procedure, 20 which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Additional techniques include 25 capture PCR (Lagerstrom et al., *PCR Methods Applic.* 1:111-19, 1991) and walking PCR (Parker et al., *Nucl. Acids. Res.* 19:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that

available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (*e.g.*, NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences may also be obtained by analysis of genomic fragments.

5 In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences that encode substantially the same or a functionally 10 equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or 15 eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding 20 sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction sites, alter glycosylation 25 patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it

may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved and purified away from the heterologous moiety.

5 Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, 10 peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. et al. (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (e.g., Creighton, T. (1983) *Proteins, Structures 15 and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from 20 other proteins, or any part thereof, to produce a variant polypeptide.

In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into appropriate expression vector, *i.e.*, a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well known to those skilled 25 in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Such techniques are described, for example, in Sambrook, J. et al. (1989) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Plainview,

N.Y., and Ausubel, F. M. et al. (1989) Current Protocols in Molecular Biology, John Wiley & Sons, New York. N.Y.

A variety of expression vector/host systems may be utilized to contain and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (*e.g.*, baculovirus); plant cell systems transformed with virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmids); or animal cell systems.

The "control elements" or "regulatory sequences" present in an expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or PSSPORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV may be advantageously used with an appropriate selectable marker.

In bacterial systems, any of a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example, when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used. Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence encoding the polypeptide of

interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of .beta.-galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel et al. (supra) and Grant et al. (1987) *Methods Enzymol.* 153:516-544.

In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) *EMBO J.* 3:1671-1680; Broglie, R. et al. (1984) *Science* 224:838-843; and Winter, J. et al. (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, for example, Hobbs, S. or Murry, L. E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in

Trichoplusia larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein.

- 5 The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or Trichoplusia larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. et al. (1994) *Proc. Natl. Acad. Sci.* 91 :3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which are appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, COS, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery and characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be transformed using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. et al. (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase (Lowy, I. et al. (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or aprt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. et al. (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. et al (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*). Additional selectable genes have been

described, for example, *trpB*, which allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). The use of visible markers has gained popularity with such markers as anthocyanins, beta-glucuronidase and its substrate GUS, 5 and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. et al. (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that the 10 gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter. Expression of the marker gene in 15 response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells that contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations and protein bioassay or immunoassay techniques which include, for example, membrane, 20 solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies specific for the product are known in the art. Examples include enzyme-linked immunosorbent 25 assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed. These and other assays are described, among other places, in Hampton, R. et al. (1990; Serological

Methods, a Laboratory Manual, APS Press, St Paul. Minn.) and Maddox, D. E. et al. (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for
5 producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by
10 addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

15 Host cells transformed with a polynucleotide sequence of interest may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of the invention may
20 be designed to contain signal sequences which direct secretion of the encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are not limited to, metal chelating peptides
25 such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen. San Diego, Calif.) between the purification domain and the

encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography) as described in Porath, J. et al. (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase cleavage site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. et al. (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length molecule.

Antibody Compositions, Fragments Thereof and Other Binding Agents

According to another aspect, the present invention further provides binding agents, such as antibodies and antigen-binding fragments thereof, that exhibit immunological binding to a tumor polypeptide disclosed herein, or to a portion, variant or derivative thereof. An antibody, or antigen-binding fragment thereof, is said to "specifically bind," "immunologically bind," and/or is "immunologically reactive" to a polypeptide of the invention if it reacts at a detectable level (within, for example, an ELISA assay) with the polypeptide, and does not react detectably with unrelated polypeptides under similar conditions.

Immunological binding, as used in this context, generally refers to the non-covalent interactions of the type which occur between an immunoglobulin molecule and an antigen for which the immunoglobulin is specific. The strength, or affinity of immunological binding interactions can be expressed in terms of the dissociation constant

(K_d) of the interaction, wherein a smaller K_d represents a greater affinity. Immunological binding properties of selected polypeptides can be quantified using methods well known in the art. One such method entails measuring the rates of antigen-binding site/antigen complex formation and dissociation, wherein those rates depend on the concentrations of 5 the complex partners, the affinity of the interaction, and on geometric parameters that equally influence the rate in both directions. Thus, both the "on rate constant" (K_{on}) and the "off rate constant" (K_{off}) can be determined by calculation of the concentrations and the actual rates of association and dissociation. The ratio of K_{off}/K_{on} enables cancellation of all 10 parameters not related to affinity, and is thus equal to the dissociation constant K_d . See, generally, Davies et al. (1990) Annual Rev. Biochem. 59:439-473.

An "antigen-binding site," or "binding portion" of an antibody refers to the part of the immunoglobulin molecule that participates in antigen binding. The antigen binding site is formed by amino acid residues of the N-terminal variable ("V") regions of the heavy ("H") and light ("L") chains. Three highly divergent stretches within the V 15 regions of the heavy and light chains are referred to as "hypervariable regions" which are interposed between more conserved flanking stretches known as "framework regions," or "FRs". Thus the term "FR" refers to amino acid sequences which are naturally found between and adjacent to hypervariable regions in immunoglobulins. In an antibody molecule, the three hypervariable regions of a light chain and the three hypervariable 20 regions of a heavy chain are disposed relative to each other in three dimensional space to form an antigen-binding surface. The antigen-binding surface is complementary to the three-dimensional surface of a bound antigen, and the three hypervariable regions of each of the heavy and light chains are referred to as "complementarity-determining regions," or "CDRs."

25 Binding agents may be further capable of differentiating between patients with and without a cancer, such as breast cancer, using the representative assays provided herein. For example, antibodies or other binding agents that bind to a tumor protein will preferably generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, more preferably at least about 30% of patients. Alternatively, or

in addition, the antibody will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the cancer. To determine whether a binding agent satisfies this requirement, biological samples (*e.g.*, blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using 5 standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. Preferably, a statistically significant number of samples with and without the disease will be assayed. Each binding agent should satisfy the above criteria; however, those of ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

10 Any agent that satisfies the above requirements may be a binding agent. For example, a binding agent may be a ribosome, with or without a peptide component, an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. *See, e.g.*, Harlow and Lane, 15 *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, antibodies can be produced by cell culture techniques, including the generation of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is 20 initially injected into any of a wide variety of mammals (*e.g.*, mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the 25 animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically. Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J. Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the desired specificity (*i.e.*, reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

A number of therapeutically useful molecules are known in the art which comprise antigen-binding sites that are capable of exhibiting immunological binding properties of an antibody molecule. The proteolytic enzyme papain preferentially cleaves IgG molecules to yield several fragments, two of which (the "F(ab)" fragments) each comprise a covalent heterodimer that includes an intact antigen-binding site. The enzyme pepsin is able to cleave IgG molecules to provide several fragments, including the "F(ab')₂"

fragment which comprises both antigen-binding sites. An "Fv" fragment can be produced by preferential proteolytic cleavage of an IgM, and on rare occasions IgG or IgA immunoglobulin molecule. Fv fragments are, however, more commonly derived using recombinant techniques known in the art. The Fv fragment includes a non-covalent $V_H::V_L$ heterodimer including an antigen-binding site which retains much of the antigen recognition and binding capabilities of the native antibody molecule. Inbar et al. (1972) Proc. Nat. Acad. Sci. USA 69:2659-2662; Hochman et al. (1976) Biochem 15:2706-2710; and Ehrlich et al. (1980) Biochem 19:4091-4096.

A single chain Fv ("sFv") polypeptide is a covalently linked $V_H::V_L$ heterodimer which is expressed from a gene fusion including V_H - and V_L -encoding genes linked by a peptide-encoding linker. Huston et al. (1988) Proc. Nat. Acad. Sci. USA 85(16):5879-5883. A number of methods have been described to discern chemical structures for converting the naturally aggregated--but chemically separated--light and heavy polypeptide chains from an antibody V region into an sFv molecule which will fold into a three dimensional structure substantially similar to the structure of an antigen-binding site. See, e.g., U.S. Pat. Nos. 5,091,513 and 5,132,405, to Huston et al.; and U.S. Pat. No. 4,946,778, to Ladner et al.

Each of the above-described molecules includes a heavy chain and a light chain CDR set, respectively interposed between a heavy chain and a light chain FR set which provide support to the CDRS and define the spatial relationship of the CDRs relative to each other. As used herein, the term "CDR set" refers to the three hypervariable regions of a heavy or light chain V region. Proceeding from the N-terminus of a heavy or light chain, these regions are denoted as "CDR1," "CDR2," and "CDR3" respectively. An antigen-binding site, therefore, includes six CDRs, comprising the CDR set from each of a heavy and a light chain V region. A polypeptide comprising a single CDR, (e.g., a CDR1, CDR2 or CDR3) is referred to herein as a "molecular recognition unit." Crystallographic analysis of a number of antigen-antibody complexes has demonstrated that the amino acid residues of CDRs form extensive contact with bound antigen, wherein the most extensive

antigen contact is with the heavy chain CDR3. Thus, the molecular recognition units are primarily responsible for the specificity of an antigen-binding site.

As used herein, the term "FR set" refers to the four flanking amino acid sequences which frame the CDRs of a CDR set of a heavy or light chain V region. Some 5 FR residues may contact bound antigen; however, FRs are primarily responsible for folding the V region into the antigen-binding site, particularly the FR residues directly adjacent to the CDRS. Within FRs, certain amino residues and certain structural features are very highly conserved. In this regard, all V region sequences contain an internal disulfide loop of around 90 amino acid residues. When the V regions fold into a binding-site, the CDRs 10 are displayed as projecting loop motifs which form an antigen-binding surface. It is generally recognized that there are conserved structural regions of FRs which influence the folded shape of the CDR loops into certain "canonical" structures--regardless of the precise CDR amino acid sequence. Further, certain FR residues are known to participate in non-covalent interdomain contacts which stabilize the interaction of the antibody heavy and 15 light chains.

A number of "humanized" antibody molecules comprising an antigen-binding site derived from a non-human immunoglobulin have been described, including chimeric antibodies having rodent V regions and their associated CDRs fused to human constant domains (Winter et al. (1991) Nature 349:293-299; Lobuglio et al. (1989) Proc. 20 Nat. Acad. Sci. USA 86:4220-4224; Shaw et al. (1987) J Immunol. 138:4534-4538; and Brown et al. (1987) Cancer Res. 47:3577-3583), rodent CDRs grafted into a human supporting FR prior to fusion with an appropriate human antibody constant domain (Riechmann et al. (1988) Nature 332:323-327; Verhoeyen et al. (1988) Science 239:1534- 1536; and Jones et al. (1986) Nature 321:522-525), and rodent CDRs supported by 25 recombinantly veneered rodent FRs (European Patent Publication No. 519,596, published Dec. 23, 1992). These "humanized" molecules are designed to minimize unwanted immunological response toward rodent antihuman antibody molecules which limits the duration and effectiveness of therapeutic applications of those moieties in human recipients.

As used herein, the terms "veneered FRs" and "recombinantly veneered FRs" refer to the selective replacement of FR residues from, *e.g.*, a rodent heavy or light chain V region, with human FR residues in order to provide a xenogeneic molecule comprising an antigen-binding site which retains substantially all of the native FR polypeptide folding structure. Veneering techniques are based on the understanding that the ligand binding characteristics of an antigen-binding site are determined primarily by the structure and relative disposition of the heavy and light chain CDR sets within the antigen-binding surface. Davies et al. (1990) Ann. Rev. Biochem. 59:439-473. Thus, antigen binding specificity can be preserved in a humanized antibody only wherein the CDR structures, their interaction with each other, and their interaction with the rest of the V region domains are carefully maintained. By using veneering techniques, exterior (*e.g.*, solvent-accessible) FR residues which are readily encountered by the immune system are selectively replaced with human residues to provide a hybrid molecule that comprises either a weakly immunogenic, or substantially non-immunogenic veneered surface.

The process of veneering makes use of the available sequence data for human antibody variable domains compiled by Kabat et al., in Sequences of Proteins of Immunological Interest, 4th ed., (U.S. Dept. of Health and Human Services, U.S. Government Printing Office, 1987), updates to the Kabat database, and other accessible U.S. and foreign databases (both nucleic acid and protein). Solvent accessibilities of V region amino acids can be deduced from the known three-dimensional structure for human and murine antibody fragments. There are two general steps in veneering a murine antigen-binding site. Initially, the FRs of the variable domains of an antibody molecule of interest are compared with corresponding FR sequences of human variable domains obtained from the above-identified sources. The most homologous human V regions are then compared residue by residue to corresponding murine amino acids. The residues in the murine FR which differ from the human counterpart are replaced by the residues present in the human moiety using recombinant techniques well known in the art. Residue switching is only carried out with moieties which are at least partially exposed (solvent accessible), and care is exercised in the replacement of amino acid residues which may have a significant effect

on the tertiary structure of V region domains, such as proline, glycine and charged amino acids.

In this manner, the resultant "veneered" murine antigen-binding sites are thus designed to retain the murine CDR residues, the residues substantially adjacent to the 5 CDRs, the residues identified as buried or mostly buried (solvent inaccessible), the residues believed to participate in non-covalent (*e.g.*, electrostatic and hydrophobic) contacts between heavy and light chain domains, and the residues from conserved structural regions of the FRs which are believed to influence the "canonical" tertiary structures of the CDR loops. These design criteria are then used to prepare recombinant nucleotide sequences 10 which combine the CDRs of both the heavy and light chain of a murine antigen-binding site into human-appearing FRs that can be used to transfect mammalian cells for the expression of recombinant human antibodies which exhibit the antigen specificity of the murine antibody molecule.

In another embodiment of the invention, monoclonal antibodies of the 15 present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include ⁹⁰Y, ¹²³I, ¹²⁵I, ¹³¹I, ¹⁸⁶Re, ¹⁸⁸Re, ²¹¹At, and ²¹²Bi. Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include 20 ricin, abrin, diphtheria toxin, cholera toxin, gelonin, Pseudomonas exotoxin, Shigella toxin, and pokeweed antiviral protein.

A therapeutic agent may be coupled (*e.g.*, covalently bonded) to a suitable monoclonal antibody either directly or indirectly (*e.g.*, via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent 25 capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulphydryl group, on one may be capable of reacting with a carbonyl-containing group, such as an anhydride or an acid halide, or with an alkyl group containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an agent or an antibody, 5 and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described in the 10 catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulphydryl groups or oxidized carbohydrate residues. There are numerous references describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.

Where a therapeutic agent is more potent when free from the antibody 15 portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitzer), by irradiation of a photolabile 20 bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter et al.), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn et al.), by serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler et al.).

It may be desirable to couple more than one agent to an antibody. In one 25 embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent may be prepared in a variety of ways. For example, more than one agent may be coupled

directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as 5 albumins (e.g., U.S. Patent No. 4,507,234, to Kato et al.), peptides and polysaccharides such as aminodextran (e.g., U.S. Patent No. 4,699,784, to Shih et al.). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (e.g., U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide 10 agents include radiohalogenated small molecules and chelating compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison et al. discloses representative chelating compounds and their synthesis.

15 T Cell Compositions

The present invention, in another aspect, provides T cells specific for a tumor polypeptide disclosed herein, or for a variant or derivative thereof. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example, T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or 20 peripheral blood of a patient, using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or unrelated humans, non-human mammals, cell lines or cultures.

25 T cells may be stimulated with a polypeptide, polynucleotide encoding a polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide of interest. Preferably, a tumor

polypeptide or polynucleotide of the invention is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

T cells are considered to be specific for a polypeptide of the present invention if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell specificity may be evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen et al., 10 *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with a tumor polypeptide (100 ng/ml - 100 µg/ml, 15 preferably 200 ng/ml - 25 µg/ml) for 3 - 7 days will typically result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN- γ) is indicative of T cell activation (see Coligan et al., *Current Protocols in Immunology*, vol. 1, Wiley Interscience 20 (Greene 1998)). T cells that have been activated in response to a tumor polypeptide, polynucleotide or polypeptide-expressing APC may be CD4 $^{+}$ and/or CD8 $^{+}$. Tumor polypeptide-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

25 For therapeutic purposes, CD4 $^{+}$ or CD8 $^{+}$ T cells that proliferate in response to a tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a tumor polypeptide, or a short peptide corresponding to an immunogenic portion of such a polypeptide, with or without the

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addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize a tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of the tumor polypeptide can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

5 Pharmaceutical Compositions

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable carriers for administration to a cell or an animal, either alone, or in combination with one or more other modalities of therapy.

10 It will be understood that, if desired, a composition as disclosed herein may be administered in combination with other agents as well, such as, *e.g.*, other proteins or polypeptides or various pharmaceutically-active agents. In fact, there is virtually no limit to other components that may also be included, given that the additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The 15 compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from host cells or other biological sources, or alternatively may be chemically synthesized as described herein. Likewise, such compositions may further comprise substituted or derivatized RNA or DNA compositions.

20 Therefore, in another aspect of the present invention, pharmaceutical compositions are provided comprising one or more of the polynucleotide, polypeptide, antibody, and/or T-cell compositions described herein in combination with a physiologically acceptable carrier. In certain preferred embodiments, the pharmaceutical compositions of the invention comprise immunogenic polynucleotide and/or polypeptide 25 compositions of the invention for use in prophylactic and therapeutic vaccine applications. Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant approach)," Plenum Press (NY, 1995).

Generally, such compositions will comprise one or more polynucleotide and/or polypeptide compositions of the present invention in combination with one or more immunostimulants.

It will be apparent that any of the pharmaceutical compositions described herein can contain pharmaceutically acceptable salts of the polynucleotides and 5 polypeptides of the invention. Such salts can be prepared, for example, from pharmaceutically acceptable non-toxic bases, including organic bases (e.g., salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (e.g., sodium, potassium, lithium, ammonium, calcium and magnesium salts).

In another embodiment, illustrative immunogenic compositions, e.g., 10 vaccine compositions, of the present invention comprise DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the polynucleotide may be administered within any of a variety of delivery systems known to those of ordinary skill in the art. Indeed, numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate polynucleotide expression systems will, of course, contain the necessary regulatory DNA regulatory 15 sequences for expression in a patient (such as a suitable promoter and terminating signal). Alternatively, bacterial delivery systems may involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the 20 polypeptide on its cell surface or secretes such an epitope.

Therefore, in certain embodiments, polynucleotides encoding immunogenic polypeptides described herein are introduced into suitable mammalian host cells for expression using any of a number of known viral-based systems. In one illustrative embodiment, retroviruses provide a convenient and effective platform for gene delivery 25 systems. A selected nucleotide sequence encoding a polypeptide of the present invention can be inserted into a vector and packaged in retroviral particles using techniques known in the art. The recombinant virus can then be isolated and delivered to a subject. A number of illustrative retroviral systems have been described (e.g., U.S. Pat. No. 5,219,740; Miller and Rosman (1989) BioTechniques 7:980-990; Miller, A. D. (1990) Human Gene Therapy

1:5-14; Scarpa et al. (1991) *Virology* 180:849-852; Burns et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:8033-8037; and Boris-Lawrie and Temin (1993) *Cur. Opin. Genet. Develop.* 3:102-109.

In addition, a number of illustrative adenovirus-based systems have also
 5 been described. Unlike retroviruses which integrate into the host genome, adenoviruses persist extrachromosomally thus minimizing the risks associated with insertional mutagenesis (Haj-Ahmad and Graham (1986) *J. Virol.* 57:267-274; Bett et al. (1993) *J. Virol.* 67:5911-5921; Mittereder et al. (1994) *Human Gene Therapy* 5:717-729; Seth et al.
 10 (1994) *J. Virol.* 68:933-940; Barr et al. (1994) *Gene Therapy* 1:51-58; Berkner, K. L. (1988) *BioTechniques* 6:616-629; and Rich et al. (1993) *Human Gene Therapy* 4:461-476).

Various adeno-associated virus (AAV) vector systems have also been developed for polynucleotide delivery. AAV vectors can be readily constructed using techniques well known in the art. See, e.g., U.S. Pat. Nos. 5,173,414 and 5,139,941; International Publication Nos. WO 92/01070 and WO 93/03769; Lebkowski et al. (1988)
 15 *Molec. Cell. Biol.* 8:3988-3996; Vincent et al. (1990) *Vaccines* 90 (Cold Spring Harbor Laboratory Press); Carter, B. J. (1992) *Current Opinion in Biotechnology* 3:533-539; Muzychka, N. (1992) *Current Topics in Microbiol. and Immunol.* 158:97-129; Kotin, R. M. (1994) *Human Gene Therapy* 5:793-801; Shelling and Smith (1994) *Gene Therapy* 1:165-169; and Zhou et al. (1994) *J. Exp. Med.* 179:1867-1875.

Additional viral vectors useful for delivering the polynucleotides encoding
 20 polypeptides of the present invention by gene transfer include those derived from the pox family of viruses, such as vaccinia virus and avian poxvirus. By way of example, vaccinia virus recombinants expressing the novel molecules can be constructed as follows. The DNA encoding a polypeptide is first inserted into an appropriate vector so that it is adjacent to a vaccinia promoter and flanking vaccinia DNA sequences, such as the sequence
 25 encoding thymidine kinase (TK). This vector is then used to transfect cells which are simultaneously infected with vaccinia. Homologous recombination serves to insert the vaccinia promoter plus the gene encoding the polypeptide of interest into the viral genome.

The resulting TK.sup.(-) recombinant can be selected by culturing the cells in the presence of 5-bromodeoxyuridine and picking viral plaques resistant thereto.

A vaccinia-based infection/transfection system can be conveniently used to provide for inducible, transient expression or coexpression of one or more polypeptides described herein in host cells of an organism. In this particular system, cells are first infected in vitro with a vaccinia virus recombinant that encodes the bacteriophage T7 RNA polymerase. This polymerase displays exquisite specificity in that it only transcribes templates bearing T7 promoters. Following infection, cells are transfected with the polynucleotide or polynucleotides of interest, driven by a T7 promoter. The polymerase expressed in the cytoplasm from the vaccinia virus recombinant transcribes the transfected DNA into RNA which is then translated into polypeptide by the host translational machinery. The method provides for high level, transient, cytoplasmic production of large quantities of RNA and its translation products. See, e.g., Elroy-Stein and Moss, Proc. Natl. Acad. Sci. USA (1990) 87:6743-6747; Fuerst et al. Proc. Natl. Acad. Sci. USA (1986) 83:8122-8126.

Alternatively, avipoxviruses, such as the fowlpox and canarypox viruses, can also be used to deliver the coding sequences of interest. Recombinant avipox viruses, expressing immunogens from mammalian pathogens, are known to confer protective immunity when administered to non-avian species. The use of an Avipox vector is particularly desirable in human and other mammalian species since members of the Avipox genus can only productively replicate in susceptible avian species and therefore are not infective in mammalian cells. Methods for producing recombinant Avipoxviruses are known in the art and employ genetic recombination, as described above with respect to the production of vaccinia viruses. See, e.g., WO 91/12882; WO 89/03429; and WO 92/03545.

Any of a number of alphavirus vectors can also be used for delivery of polynucleotide compositions of the present invention, such as those vectors described in U.S. Patent Nos. 5,843,723; 6,015,686; 6,008,035 and 6,015,694. Certain vectors based on Venezuelan Equine Encephalitis (VEE) can also be used, illustrative examples of which can be found in U.S. Patent Nos. 5,505,947 and 5,643,576.

Moreover, molecular conjugate vectors, such as the adenovirus chimeric vectors described in Michael et al. *J. Biol. Chem.* (1993) 268:6866-6869 and Wagner et al. *Proc. Natl. Acad. Sci. USA* (1992) 89:6099-6103, can also be used for gene delivery under the invention.

5 Additional illustrative information on these and other known viral-based delivery systems can be found, for example, in Fisher-Hoch et al., *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner et al., *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner et al., *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; 10 Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld et al., *Science* 252:431-434, 1991; Kolls et al., *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler et al., *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman et al., *Circulation* 88:2838-2848, 1993; and Guzman et al., *Cir. Res.* 73:1202-1207, 1993.

15 In certain embodiments, a polynucleotide may be integrated into the genome of a target cell. This integration may be in the specific location and orientation *via* homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the polynucleotide may be stably maintained in the cell as a separate, episomal segment of DNA. Such polynucleotide segments or "episomes" encode sequences sufficient to permit maintenance 20 and replication independent of or in synchronization with the host cell cycle. The manner in which the expression construct is delivered to a cell and where in the cell the polynucleotide remains is dependent on the type of expression construct employed.

In another embodiment of the invention, a polynucleotide is administered/delivered as "naked" DNA, for example as described in Ulmer et al., *Science* 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake 25 of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

In still another embodiment, a composition of the present invention can be delivered via a particle bombardment approach, many of which have been described. In

PCT/US94/03636 2000-03-03 2000-03-03

one illustrative example, gas-driven particle acceleration can be achieved with devices such as those manufactured by Powderject Pharmaceuticals PLC (Oxford, UK) and Powderject Vaccines Inc. (Madison, WI), some examples of which are described in U.S. Patent Nos. 5,846,796; 6,010,478; 5,865,796; 5,584,807; and EP Patent No. 0500 799. This approach 5 offers a needle-free delivery approach wherein a dry powder formulation of microscopic particles, such as polynucleotide or polypeptide particles, are accelerated to high speed within a helium gas jet generated by a hand held device, propelling the particles into a target tissue of interest.

In a related embodiment, other devices and methods that may be useful for 10 gas-driven needle-less injection of compositions of the present invention include those provided by Bioject, Inc. (Portland, OR), some examples of which are described in U.S. Patent Nos. 4,790,824; 5,064,413; 5,312,335; 5,383,851; 5,399,163; 5,520,639 and 5,993,412.

According to another embodiment, the pharmaceutical compositions 15 described herein will comprise one or more immunostimulants in addition to the immunogenic polynucleotide, polypeptide, antibody, T-cell and/or APC compositions of this invention. An immunostimulant refers to essentially any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. One preferred type of immunostimulant comprises an adjuvant. Many adjuvants contain a 20 substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins. Certain adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, 25 Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA); aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres;

monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF, interleukin-2, -7, -12, and other like growth factors, may also be used as adjuvants.

- Within certain embodiments of the invention, the adjuvant composition is preferably one that induces an immune response predominantly of the Th1 type. High levels of Th1-type cytokines (e.g., IFN- γ , TNF α , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast, high levels of Th2-type cytokines (e.g., IL-4, IL-5, IL-6 and IL-10) tend to favor the induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-type responses.
- 10 Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.
- 15 Certain preferred adjuvants for eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-de-O-acetylated monophosphoryl lipid A, together with an aluminum salt. MPL® adjuvants are available from Corixa Corporation (Seattle, WA; *see*, for example, US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato et al., *Science* 273:352, 1996. Another preferred adjuvant comprises a saponin, such as Quil A, or derivatives thereof, including QS21 and QS7 (Aquila Biopharmaceuticals Inc., Framingham, MA); Escin; Digitonin; or *Gypsophila* or *Chenopodium quinoa* saponins. Other preferred formulations include more than one saponin in the adjuvant combinations of the present invention, for example combinations of at least two of the following group comprising QS21, QS7, Quil A, β -escin, or digitonin.

Alternatively the saponin formulations may be combined with vaccine vehicles composed of chitosan or other polycationic polymers, polylactide and polylactide-co-glycolide particles, poly-N-acetyl glucosamine-based polymer matrix, particles composed of polysaccharides or chemically modified polysaccharides, liposomes and lipid-based particles, particles composed of glycerol monoesters, etc. The saponins may also be formulated in the presence of cholesterol to form particulate structures such as liposomes or ISCOMs. Furthermore, the saponins may be formulated together with a polyoxyethylene ether or ester, in either a non-particulate solution or suspension, or in a particulate structure such as a paucilamellar liposome or ISCOM. The saponins may also be formulated with excipients such as Carbopol^R to increase viscosity, or may be formulated in a dry powder form with a powder excipient such as lactose.

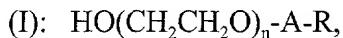
In one preferred embodiment, the adjuvant system includes the combination of a monophosphoryl lipid A and a saponin derivative, such as the combination of QS21 and 3D-MPL[®] adjuvant, as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO 96/33739. Other preferred formulations comprise an oil-in-water emulsion and tocopherol. Another particularly preferred adjuvant formulation employing QS21, 3D-MPL[®] adjuvant and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

Another enhanced adjuvant system involves the combination of a CpG-containing oligonucleotide and a saponin derivative particularly the combination of CpG and QS21 is disclosed in WO 00/09159. Preferably the formulation additionally comprises an oil in water emulsion and tocopherol.

Additional illustrative adjuvants for use in the pharmaceutical compositions of the invention include Montanide ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (*e.g.*, SBAS-2 or SBAS-4, available from SmithKline Beecham, Rixensart, Belgium), Detox (Enhanzym[®]) (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures of which are

incorporated herein by reference in their entireties, and polyoxyethylene ether adjuvants such as those described in WO 99/52549A1.

Other preferred adjuvants include adjuvant molecules of the general formula



- 5 wherein, n is 1-50, A is a bond or $-\text{C}(\text{O})-$, R is C_{1-50} alkyl or Phenyl C_{1-50} alkyl.

One embodiment of the present invention consists of a vaccine formulation comprising a polyoxyethylene ether of general formula (I), wherein n is between 1 and 50, preferably 4-24, most preferably 9; the R component is C_{1-50} , preferably $\text{C}_4\text{-C}_{20}$ alkyl and most preferably C_{12} alkyl, and A is a bond. The concentration of the polyoxyethylene ethers 10 should be in the range 0.1-20%, preferably from 0.1-10%, and most preferably in the range 0.1-1%. Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether, polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether. Polyoxyethylene ethers such as polyoxyethylene lauryl 15 ether are described in the Merck index (12th edition: entry 7717). These adjuvant molecules are described in WO 99/52549.

The polyoxyethylene ether according to the general formula (I) above may, if desired, be combined with another adjuvant. For example, a preferred adjuvant combination is preferably with CpG as described in the pending UK patent application GB 20 9820956.2.

According to another embodiment of this invention, an immunogenic composition described herein is delivered to a host via antigen presenting cells (APCs), such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to 25 increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

Certain preferred embodiments of the present invention use dendritic cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (see Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In general, dendritic cells may be identified based on their typical shape (stellate *in situ*, with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (see Zitvogel et al., *Nature Med.* 4:594-600, 1998).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF α to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF α , CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fc γ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface

molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide of the invention
5 (or portion or other variant thereof) such that the encoded polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a pharmaceutical composition comprising such transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient,
10 resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi et al., *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the tumor polypeptide, DNA
15 (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the
20 polypeptide.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier will typically vary depending on the mode of administration. Compositions of the present invention may be formulated for any appropriate manner of administration, including for
25 example, topical, oral, nasal, mucosal, intravenous, intracranial, intraperitoneal, subcutaneous and intramuscular administration.

Carriers for use within such pharmaceutical compositions are biocompatible, and may also be biodegradable. In certain embodiments, the formulation preferably provides a relatively constant level of active component release. In other embodiments,

however, a more rapid rate of release immediately upon administration may be desired. The formulation of such compositions is well within the level of ordinary skill in the art using known techniques. Illustrative carriers useful in this regard include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like.

5 Other illustrative delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 10 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

In another illustrative embodiment, biodegradable microspheres (*e.g.*, polylactate polyglycolate) are employed as carriers for the compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344, 15 5,407,609 and 5,942,252. Modified hepatitis B core protein carrier systems, such as described in WO/99 40934, and references cited therein, will also be useful for many applications. Another illustrative carrier/delivery system employs a carrier comprising particulate-protein complexes, such as those described in U.S. Patent No. 5,928,647, which 20 are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

The pharmaceutical compositions of the invention will often further comprise one or more buffers (*e.g.*, neutral buffered saline or phosphate buffered saline), carbohydrates (*e.g.*, glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents 25 such as EDTA or glutathione, adjuvants (*e.g.*, aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate.

The pharmaceutical compositions described herein may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are typically sealed in such a way to preserve the sterility and stability of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily 5 or aqueous vehicles. Alternatively, a pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

The development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including *e.g.*, 10 oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation, is well known in the art, some of which are briefly discussed below for general purposes of illustration.

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be 15 formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, 20 syrups, wafers, and the like (see, for example, Mathiowitz *et al.*, Nature 1997 Mar 27;386(6623):410-4; Hwang *et al.*, Crit Rev Ther Drug Carrier Syst 1998;15(3):243-84; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451). Tablets, troches, pills, capsules and the like may also contain any of a variety of additional components, for example, a binder, such as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such 25 as dicalcium phosphate; a disintegrating agent, such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be

present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may
5 be incorporated into sustained-release preparation and formulations.

Typically, these formulations will contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active
10 compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and
15 treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. Alternatively, the active ingredient may be incorporated into an oral solution such as one
20 containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

25 In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even intraperitoneally. Such approaches are well known to the skilled artisan, some of which are further described, for example, in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363. In certain embodiments, solutions of the active compounds as free base

or pharmacologically acceptable salts may be prepared in water suitably mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations generally will contain a preservative to prevent the
5 growth of microorganisms.

Illustrative pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (for example, see U. S. Patent 5,466,468). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists.
10 It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms, such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (e.g., glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for
15 example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and/or by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or
20 sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

In one embodiment, for parenteral administration in an aqueous solution, the solution should be suitably buffered if necessary and the liquid diluent first rendered
25 isotonic with sufficient saline or glucose. These particular aqueous solutions are especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or

injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. Moreover, for human administration, preparations will of course preferably meet sterility, pyrogenicity,
5 and the general safety and purity standards as required by FDA Office of Biologics standards.

In another embodiment of the invention, the compositions disclosed herein may be formulated in a neutral or salt form. Illustrative pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the protein) and which
10 are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation,
15 solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective.

The carriers can further comprise any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of
20 such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions. The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not
25 produce an allergic or similar untoward reaction when administered to a human.

In certain embodiments, the pharmaceutical compositions may be delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the lungs *via* nasal aerosol sprays has been described, *e.g.*, in U. S. Patent 5,756,353 and U. S. Patent

5,804,212. Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, J Controlled Release 1998 Mar 2;52(1-2):81-7) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871) are also well-known in the pharmaceutical arts. Likewise, illustrative transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045.

10 In certain embodiments, liposomes, nanocapsules, microparticles, lipid particles, vesicles, and the like, are used for the introduction of the compositions of the present invention into suitable host cells/organisms. In particular, the compositions of the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like. Alternatively, compositions of the present invention can be bound, either covalently or non-covalently, to the surface of such carrier vehicles.

15 The formation and use of liposome and liposome-like preparations as potential drug carriers is generally known to those of skill in the art (see for example, Lasic, Trends Biotechnol 1998 Jul;16(7):307-21; Takakura, Nippon Rinsho 1998 Mar;56(3):691-5; Chandran *et al.*, Indian J Exp Biol. 1997 Aug;35(8):801-9; Margalit, Crit Rev Ther Drug Carrier Syst. 1995;12(2-3):233-61; U.S. Patent 5,567,434; U.S. Patent 5,552,157; U.S. Patent 5,565,213; U.S. Patent 5,738,868 and U.S. Patent 5,795,587, each specifically incorporated herein by reference in its entirety).

20 Liposomes have been used successfully with a number of cell types that are normally difficult to transfet by other procedures, including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, J Biol Chem. 1990 Sep 25;265(27):16337-42; Muller *et al.*, DNA Cell Biol. 1990 Apr;9(3):221-9). In addition, liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, various drugs, radiotherapeutic agents, enzymes, viruses, transcription factors, allosteric effectors and the like, into a variety of cultured cell lines and animals. Furthermore, the use of liposomes does not appear to be associated with autoimmune responses or unacceptable toxicity after systemic delivery.

In certain embodiments, liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs)).

Alternatively, in other embodiments, the invention provides for pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (see, for example, Quintanar-Guerrero *et al.*, Drug Dev Ind Pharm. 1998 Dec;24(12):1113-28). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1 μm) may be designed using polymers able to be degraded *in vivo*. Such particles can be made as described, for example, by Couvreur *et al.*, Crit Rev Ther Drug Carrier Syst. 1988;5(1):1-20; zur Muhlen *et al.*, Eur J Pharm Biopharm. 1998 Mar;45(2):149-55; Zambaux *et al.* J Controlled Release. 1998 Jan 2;50(1-3):31-40; and U. S. Patent 5,145,684.

Cancer Therapeutic Methods

In further aspects of the present invention, the pharmaceutical compositions described herein may be used for the treatment of cancer, particularly for the immunotherapy of breast cancer. Within such methods, the pharmaceutical compositions described herein are administered to a patient, typically a warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer. Accordingly, the above pharmaceutical compositions may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. As discussed above, administration of the pharmaceutical compositions may be by any suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous host

immune system to react against tumors with the administration of immune response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established tumor-
5 immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8⁺ cytotoxic T lymphocytes and CD4⁺ T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-activated killer
10 cells), B cells and antigen-presenting cells (such as dendritic cells and macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic antibodies (as described above
15 and in U.S. Patent No. 4,918,164) for passive immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture conditions typically use
20 intermittent stimulation with antigen, often in the presence of cytokines (such as IL-2) and non-dividing feeder cells. As noted above, immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage, monocyte, fibroblast and/or B cells, may be pulsed with
25 immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a recombinant virus or other expression system. Cultured effector cells for use in therapy must be able to grow and distribute widely, and to survive long term *in vivo*.

Studies have shown that cultured effector cells can be induced to grow in vivo and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (see, for example, Cheever et al., *Immunological Reviews* 157:177, 1997).

5 Alternatively, a vector expressing a polypeptide recited herein may be introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitory, intraperitoneal or intratumor administration.

10 Routes and frequency of administration of the therapeutic compositions described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably, 15 between 1 and 10 doses may be administered over a 52 week period. Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the 20 basal (*i.e.*, untreated) level. Such response can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor cells *in vitro*. Such vaccines should also be capable of causing an immune response that leads to an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial or longer disease-free survival) in vaccinated 25 patients as compared to non-vaccinated patients. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 25 µg to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit. Such a response can be monitored by establishing an improved clinical outcome (e.g., more frequent remissions, complete or partial, or longer disease-free survival) in treated patients
5 as compared to non-treated patients. Increases in preexisting immune responses to a tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using standard proliferation, cytotoxicity or cytokine assays, which may be performed using samples obtained from a patient before and after treatment.

Cancer Detection and Diagnostic Compositions, Methods and Kits

10 In general, a cancer may be detected in a patient based on the presence of one or more breast tumor proteins and/or polynucleotides encoding such proteins in a biological sample (for example, blood, sera, sputum urine and/or tumor biopsies) obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as breast cancer. In addition, such proteins may be
15 useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the biological sample. Polynucleotide primers and probes may be used to detect the level of mRNA encoding a tumor protein, which is also indicative of the presence or absence of a cancer. In general, a breast tumor sequence should be present at a level that is at least three fold higher in tumor
20 tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in the art for using a binding agent to detect polypeptide markers in a sample. See, e.g., Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, the presence or absence of a cancer in a patient may be determined by (a)
25 contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c) comparing the level of polypeptide with a predetermined cut-off value.

- In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding agent/polypeptide complex.
- 5 Such detection reagents may comprise, for example, a binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized binding agent after incubation of the binding agent
10 with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length breast tumor proteins and polypeptide portions thereof to which the binding agent binds, as described above.
- 15 The solid support may be any material known to those of ordinary skill in the art to which the tumor protein may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic
20 particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681. The binding agent may be immobilized on the solid support using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which
25 may be a direct linkage between the agent and functional groups on the support or may be a linkage by way of a cross-linking agent). Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1

hour and about 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10 µg, and preferably about 100 ng to about 1 µg, is sufficient to immobilize an adequate amount of binding agent.

5 Covalent attachment of binding agent to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group
10 on the support with an amine and an active hydrogen on the binding partner (*see, e.g.,* Pierce Immunotechnology Catalog and Handbook, 1991, at A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay. This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the sample, such that
15 polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a method appropriate for the
20 specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20TM (Sigma Chemical Co., St. Louis, MO). The immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with breast cancer. Preferably, the contact time is
25

sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve equilibrium may be readily determined by assaying the level of binding that occurs over a period of time. At room temperature, an 5 incubation time of about 30 minutes is generally sufficient.

Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20TM. The second antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

10 The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide. An appropriate amount of time may generally be determined by assaying the level of binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting 15 the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may 20 generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

To determine the presence or absence of a cancer, such as breast cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred 25 embodiment, the cut-off value for the detection of a cancer is the average mean signal obtained when the immobilized antibody is incubated with samples from patients without the cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve,

according to the method of Sackett et al., *Clinical Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985, p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%-specificity) that correspond to each possible cut-off value for 5 the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false 10 negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the 15 immobilized binding agent as the sample passes through the membrane. A second, labeled binding agent then binds to the binding agent-polypeptide complex as a solution containing the second binding agent flows through the membrane. The detection of bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the 20 sample. The sample migrates along the membrane through a region containing second binding agent and to the area of immobilized binding agent. Concentration of second binding agent at the area of immobilized antibody indicates the presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. 25 In general, the amount of binding agent immobilized on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a positive signal in the two-antibody sandwich assay, in the format discussed above. Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of

antibody immobilized on the membrane ranges from about 25 ng to about 1 μ g, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological sample.

Of course, numerous other assay protocols exist that are suitable for use
5 with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such tumor protein specific antibodies may correlate with the presence of a cancer.

10 A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with a tumor protein in a biological sample. Within certain methods, a biological sample comprising CD4 $^{+}$ and/or CD8 $^{+}$ T cells isolated from a patient is incubated with a tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the
15 presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For example, T cells may be isolated from a patient by routine techniques (such as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (*e.g.*, 5 - 25 μ g/ml). It may be desirable to
20 incubate another aliquot of a T cell sample in the absence of tumor polypeptide to serve as a control. For CD4 $^{+}$ T cells, activation is preferably detected by evaluating proliferation of the T cells. For CD8 $^{+}$ T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a
25 cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on the level of mRNA encoding a tumor protein in a biological sample. For example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of a tumor cDNA derived from a biological sample, wherein at

least one of the oligonucleotide primers is specific for (*i.e.*, hybridizes to) a polynucleotide encoding the tumor protein. The amplified cDNA is then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a polynucleotide encoding a tumor protein may be
5 used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers and probes should comprise an oligonucleotide sequence that has at least about 60%, preferably at least about 75% and more preferably at least about 90%, identity to a portion of a
10 polynucleotide encoding a tumor protein of the invention that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a polypeptide described herein under moderately stringent conditions, as defined above. Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are
15 at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule having a sequence as disclosed herein. Techniques for both PCR based assays and hybridization assays are well known in the art (*see*, for example, Mullis et al., *Cold Spring Harbor Symp. Quant. Biol.*, 51:263, 1987; Erlich ed.,
20 *PCR Technology*, Stockton Press, NY, 1989).

One preferred assay employs RT-PCR, in which PCR is applied in conjunction with reverse transcription. Typically, RNA is extracted from a biological sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be
25 separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and from an individual who is not afflicted with a cancer. The amplification reaction may be performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in

expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered positive.

In another embodiment, the compositions described herein may be used as markers for the progression of cancer. In this embodiment, assays as described above for 5 the diagnosis of a cancer may be performed over time, and the change in the level of reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter performed as needed. In general, a cancer is progressing in those patients in whom the level of polypeptide or polynucleotide detected increases over time. In contrast, the cancer is not 10 progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

Certain *in vivo* diagnostic assays may be performed directly on a tumor. One such assay involves contacting tumor cells with a binding agent. The bound binding agent may then be detected directly or indirectly via a reporter group. Such binding agents 15 may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

As noted above, to improve sensitivity, multiple tumor protein markers may be assayed within a given sample. It will be apparent that binding agents specific for different proteins provided herein may be combined within a single assay. Further, 20 multiple primers or probes may be used concurrently. The selection of tumor protein markers may be based on routine experiments to determine combinations that results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

The present invention further provides kits for use within any of the above 25 diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment. For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to a tumor protein. Such antibodies or fragments may be provided attached to a support material, as described above. One or

more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

5 Alternatively, a kit may be designed to detect the level of mRNA encoding a tumor protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridizes to a polynucleotide encoding a tumor protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that may be present within such kits
10 include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a tumor protein.

The following Examples are offered by way of illustration and not by way of limitation.

15

EXAMPLE 1

PREPARATION OF BREAST TUMOR-SPECIFIC cDNAs USING DIFFERENTIAL DISPLAY RT-PCR

This Example illustrates the preparation of cDNA molecules encoding
20 breast tumor-specific polypeptides using a differential display screen.

A. Preparation of B18Ag1 cDNA and Characterization of mRNA Expression

Tissue samples were prepared from breast tumor and normal tissue of a patient with breast cancer that was confirmed by pathology after removal from the patient. Normal RNA and tumor RNA was extracted from the samples and mRNA was isolated and
25 converted into cDNA using a (dT)₁₂AG (SEQ ID NO:130) anchored 3' primer. Differential display PCR was then executed using a randomly chosen primer (CTTCAACCTC) (SEQ ID NO:103). Amplification conditions were standard buffer containing 1.5 mM MgCl₂, 20 pmol of primer, 500 pmol dNTP, and 1 unit of *Taq* DNA polymerase (Perkin-Elmer,

Branchburg, NJ). Forty cycles of amplification were performed using 94°C denaturation for 30 seconds, 42°C annealing for 1 minute, and 72°C extension for 30 seconds. An RNA fingerprint containing 76 amplified products was obtained. Although the RNA fingerprint of breast tumor tissue was over 98% identical to that of the normal breast tissue, a band was
5 repeatedly observed to be specific to the RNA fingerprint pattern of the tumor. This band was cut out of a silver stained gel, subcloned into the T-vector (Novagen, Madison, WI) and sequenced.

The sequence of the cDNA, referred to as B18Ag1, is provided in SEQ ID NO:1. A database search of GENBANK and EMBL revealed that the B18Ag1 fragment
10 initially cloned is 77% identical to the endogenous human retroviral element S71, which is a truncated retroviral element homologous to the Simian Sarcoma Virus (SSV). S71 contains an incomplete *gag* gene, a portion of the *pol* gene and an LTR-like structure at the 3' terminus (*see* Werner et al., *Virology* 174:225-238 (1990)). B18Ag1 is also 64% identical to SSV in the region corresponding to the P30 (*gag*) locus. B18Ag1 contains
15 three separate and incomplete reading frames covering a region which shares considerable homology to a wide variety of *gag* proteins of retroviruses which infect mammals. In addition, the homology to S71 is not just within the *gag* gene, but spans several kb of sequence including an LTR.

B18Ag1-specific PCR primers were synthesized using computer analysis
20 guidelines. RT-PCR amplification (94°C, 30 seconds; 60°C → 42°C, 30 seconds; 72°C, 30 seconds for 40 cycles) confirmed that B18Ag1 represents an actual mRNA sequence present at relatively high levels in the patient's breast tumor tissue. The primers used in amplification were B18Ag1-1 (CTG CCT GAG CCA CAA ATG) (SEQ ID NO:128) and B18Ag1-4 (CCG GAG GAG GAA GCT AGA GGA ATA) (SEQ ID NO:129) at a 3.5 mM
25 magnesium concentration and a pH of 8.5, and B18Ag1-2 (ATG GCT ATT TTC GGG GCC TGA CA) (SEQ ID NO:126) and B18Ag1-3 (CCG GTA TCT CCT CGT GGG TAT T) (SEQ ID NO:127) at 2 mM magnesium at pH 9.5. The same experiments showed exceedingly low to nonexistent levels of expression in this patient's normal breast tissue (*see* Figure 1). RT-PCR experiments were then used to show that B18Ag1 mRNA is

present in nine other breast tumor samples (from Brazilian and American patients) but absent in, or at exceedingly low levels in, the normal breast tissue corresponding to each cancer patient. RT-PCR analysis has also shown that the B18Ag1 transcript is not present in various normal tissues (including lymph node, myocardium and liver) and present at 5 relatively low levels in PBMC and lung tissue. The presence of B18Ag1 mRNA in breast tumor samples, and its absence from normal breast tissue, has been confirmed by Northern blot analysis, as shown in Figure 2.

The differential expression of B18Ag1 in breast tumor tissue was also confirmed by RNase protection assays. Figure 3 shows the level of B18Ag1 mRNA in 10 various tissue types as determined in four different RNase protection assays. Lanes 1-12 represent various normal breast tissue samples, lanes 13-25 represent various breast tumor samples; lanes 26-27 represent normal prostate samples; lanes 28-29 represent prostate tumor samples; lanes 30-32 represent colon tumor samples; lane 33 represents normal aorta; lane 34 represents normal small intestine; lane 35 represents normal skin, lane 36 15 represents normal lymph node; lane 37 represents normal ovary; lane 38 represents normal liver; lane 39 represents normal skeletal muscle; lane 40 represents a first normal stomach sample, lane 41 represents a second normal stomach sample; lane 42 represents a normal lung; lane 43 represents normal kidney; and lane 44 represents normal pancreas. Interexperimental comparison was facilitated by including a positive control RNA of 20 known β -actin message abundance in each assay and normalizing the results of the different assays with respect to this positive control.

RT-PCR and Southern Blot analysis has shown the B18Ag1 locus to be present in human genomic DNA as a single copy endogenous retroviral element. A genomic clone of approximately 12-18 kb was isolated using the initial B18Ag1 sequence 25 as a probe. Four additional subclones were also isolated by XbaI digestion. Additional retroviral sequences obtained from the ends of the XbaI digests of these clones (located as shown in Figure 4) are shown as SEQ ID NO:3 - SEQ ID NO:10, where SEQ ID NO:3 shows the location of the sequence labeled 10 in Figure 4, SEQ ID NO:4 shows the location of the sequence labeled 11-29, SEQ ID NO:5 shows the location of the sequence

labeled 3, SEQ ID NO:6 shows the location of the sequence labeled 6, SEQ ID NO:7 shows the location of the sequence labeled 12, SEQ ID NO:8 shows the location of the sequence labeled 13, SEQ ID NO:9 shows the location of the sequence labeled 14 and SEQ ID NO:10 shows the location of the sequence labeled 11-22.

5 Subsequent studies demonstrated that the 12-18 kb genomic clone contains a retroviral element of about 7.75 kb, as shown in Figures 5A and 5B. The sequence of this retroviral element is shown in SEQ ID NO:141. The numbered line at the top of Figure 5A represents the sense strand sequence of the retroviral genomic clone. The box below this line shows the position of selected restriction sites. The arrows depict the different
10 overlapping clones used to sequence the retroviral element. The direction of the arrow shows whether the single-pass subclone sequence corresponded to the sense or anti-sense strand. Figure 5B is a schematic diagram of the retroviral element containing B18Ag1 depicting the organization of viral genes within the element. The open boxes correspond to predicted reading frames, starting with a methionine, found throughout the element. Each
15 of the six likely reading frames is shown, as indicated to the left of the boxes, with frames 1-3 corresponding to those found on the sense strand.

Using the cDNA of SEQ ID NO:1 as a probe, a longer cDNA was obtained (SEQ ID NO:227) which contains minor nucleotide differences (less than 1%) compared to the genomic sequence shown in SEQ ID NO:141.

20 B. Preparation of cDNA Molecules Encoding Other Breast Tumor-Specific Polypeptides

Normal RNA and tumor RNA was prepared and mRNA was isolated and converted into cDNA using a (dT)₁₂AG anchored 3' primer, as described above. Differential display PCR was then executed using the randomly chosen primers of SEQ ID
25 NOs:87-125. Amplification conditions were as noted above, and bands observed to be specific to the RNA fingerprint pattern of the tumor were cut out of a silver stained gel, subcloned into either the T-vector (Novagen, Madison, WI) or the pCRII vector (Invitrogen, San Diego, CA) and sequenced. The sequences are provided in SEQ ID

NO:11 - SEQ ID NO:86. Of the 79 sequences isolated, 67 were found to be novel (SEQ ID NOs:11-26 and 28-77) (*see also* Figures 6-20).

An extended DNA sequence (SEQ ID NO:290) for the antigen B15Ag1 (originally identified partial sequence provided in SEQ ID NO:27) was obtained in further studies. Comparison of the sequence of SEQ ID NO:290 with those in the gene bank as described above, revealed homology to the known human β -A activin gene. Further studies led to the isolation of the full-length cDNA sequence for the antigen B21GT2 (also referred to as B311D; originally identified partial cDNA sequence provided in SEQ ID NOs:56). The full-length sequence is provided in SEQ ID NO:307, with the corresponding amino acid sequence being provided in SEQ ID NO:308. Further studies led to the isolation of a splice variant of B311D. The B311D clone of SEQ ID NO:316 was sequenced and a XhoI/NotI fragment from this clone was gel purified and 32P-cDTP labeled by random priming for use as a probe for further screening to obtain additional B311D gene sequence. Two fractions of a human breast tumor cDNA bacterial library were screened using standard techniques. One of the clones isolated in this manner yielded additional sequence which includes a poly A+ tail. The determined cDNA sequence of this clone (referred to as B311D_BT1_1A) is provided in SEQ ID NO:317. The sequences of SEQ ID NOs:316 and 317 were found to share identity over a 464 bp region, with the sequences diverging near the poly A+ sequence of SEQ ID NO:317.

Subsequent studies identified an additional 146 sequences (SEQ ID NOs:142-289), of which 115 appeared to be novel (SEQ ID NOs:142, 143, 146-152, 154-166, 168-176, 178-192, 194-198, 200-204, 206, 207, 209-214, 216, 218, 219, 221-240, 243-245, 247, 250, 251, 253, 255, 257-266, 268, 269, 271-273, 275, 276, 278, 280, 281, 284, 288 and 291). To the best of the inventors' knowledge none of the previously identified sequences have heretofore been shown to be expressed at a greater level in human breast tumor tissue than in normal breast tissue.

In further studies, several different splice forms of the antigen B11Ag1 (also referred to as B305D) were isolated, with each of the various splice forms containing slightly different versions of the B11Ag1 coding frame. Splice junction sequences define individual exons

which, in various patterns and arrangements, make up the various splice forms. Primers were designed to examine the expression pattern of each of the exons using RT-PCR as described below. Each exon was found to show the same expression pattern as the original B11Ag1 clone, with expression being breast tumor-, normal prostate- and normal testis-specific. The determined cDNA sequences for the isolated protein coding exons are provided in SEQ ID NOs:292-298, respectively. The predicted amino acid sequences corresponding to the sequences of SEQ ID NOs:292 and 298 are provided in SEQ ID NOs:299 and 300. Additional studies using rapid amplification of cDNA ends (RACE), a 5' specific primer to one of the splice forms of B11Ag1 provided above and a breast adenocarcinoma, led to the isolation of three additional, related, splice forms referred to as isoforms B11C-15, B11C-8 and B11C-9,16. The determined cDNA sequences for these isoforms are provided in SEQ ID NO: 301-303, with the corresponding predicted amino acid sequences being provided in SEQ ID NOs:304-306.

The protein coding region of B11C-15 (SEQ ID NO: 301; also referred to as B305D isoform C) was used as a query sequence in a BLASTN search of the Genbank DNA database. A match was found to a genomic clone from chromosome 21 (Accesssion no. AP001465). The pairwise alignments provided in the BLASTN output were used to identify the putative exon, or coding, sequence of the chromosome 21 sequence that corresponds to the B305D sequence. Based on the BlastN pairwise alignments, the following pieces of GenBank record AP001465 were put together: base pairs 67978-68499, 72870-72987, 73144-73335, 76085-76206, 77905-78085, 80520-80624, 87602-87633. This sequence was then aligned with the B305D isoform C sequence using the DNA Star Seqman program and excess sequence was deleted in such a way as to maintain the sequence most similar to B305D. The final edited form of the chromosome 21 sequence was 96.5% identical to B305D. This resulting edited sequence from chromosome 21 was then translated and found to contain no stop codons other than the final stop codon in the same position as that for B305D. As with B305D, the chromosome 21 sequence (provided in SEQ ID NO: 325) encoded a protein (SEQ ID NO: 326)with 384 amino acids. An

alignment of this protein with the B305D isoform C protein (SEQ ID NO: 304) showed 90% amino acid identity.

In subsequent studies on B305D isoform A (cDNA sequence provided in SEQ ID NO:292), the cDNA sequence (provided in SEQ ID NO:313) was found to contain 5 an additional guanine residue at position 884, leading to a frameshift in the open reading frame. The determined DNA sequence of this ORF is provided in SEQ ID NO:314. This frameshift generates a protein sequence (provided in SEQ ID NO:315) of 293 amino acids that contains the C-terminal domain common to the other isoforms of B305D but that differs in the N-terminal region.

10

EXAMPLE 2

PREPARATION OF B18Ag1 DNA FROM HUMAN GENOMIC DNA

This Example illustrates the preparation of B18Ag1 DNA by amplification 15 from human genomic DNA.

B18Ag1 DNA may be prepared from 250 ng human genomic DNA using 20 pmol of B18Ag1 specific primers, 500 pmol dNTPS and 1 unit of *Taq* DNA polymerase (Perkin Elmer, Branchburg, NJ) using the following amplification parameters: 94°C for 30 seconds denaturing, 30 seconds 60°C to 42°C touchdown annealing in 2°C increments 20 every two cycles and 72°C extension for 30 seconds. The last increment (a 42°C annealing temperature) should cycle 25 times. Primers were selected using computer analysis. Primers synthesized were B18Ag1-1, B18Ag1-2, B18Ag1-3, and B18Ag1-4. Primer pairs that may be used are 1+3, 1+4, 2+3, and 2+4.

Following gel electrophoresis, the band corresponding to B18Ag1 DNA 25 may be excised and cloned into a suitable vector.

EXAMPLE 3
PREPARATION OF B18Ag1 DNA FROM BREAST TUMOR cDNA

This Example illustrates the preparation of B18Ag1 DNA by amplification
 5 from human breast tumor cDNA.

First strand cDNA is synthesized from RNA prepared from human breast tumor tissue in a reaction mixture containing 500 ng poly A+ RNA, 200 pmol of the primer (T)₁₂AG (*i.e.*, TTT TTT TTT TTT AG) (SEQ ID NO:130), 1X first strand reverse transcriptase buffer, 6.7 mM DTT, 500 mmol dNTPs, and 1 unit AMV or MMLV reverse
 10 transcriptase (from any supplier, such as Gibco-BRL (Grand Island, NY)) in a final volume of 30 µl. After first strand synthesis, the cDNA is diluted approximately 25 fold and 1 µl is used for amplification as described in Example 2. While some primer pairs can result in a heterogeneous population of transcripts, the primers B18Ag1-2 (5'ATG GCT ATT TTC GGG GGC TGA CA) (SEQ ID NO:126) and B18Ag1-3 (5'CCG GTA TCT CCT CGT
 15 GGG TAT T) (SEQ ID NO:127) yield a single 151 bp amplification product.

EXAMPLE 4
IDENTIFICATION OF B-CELL AND T-CELL EPITOPES OF B18Ag1

20 This Example illustrates the identification of B18Ag1 epitopes.

The B18Ag1 sequence can be screened using a variety of computer algorithms. To determine B-cell epitopes, the sequence can be screened for hydrophobicity and hydrophilicity values using the method of Hopp, *Prog. Clin. Biol. Res.* 172B:367-77 (1985) or, alternatively, Cease et al., *J. Exp. Med.* 164:1779-84 (1986) or Spouge et al., *J.
 25 Immunol.* 138:204-12 (1987). Additional Class II MHC (antibody or B-cell) epitopes can be predicted using programs such as AMPHI (*e.g.*, Margalit et al., *J. Immunol.* 138:2213 (1987)) or the methods of Rothbard and Taylor (*e.g.*, *EMBO J.* 7:93 (1988)).

Once peptides (15-20 amino acids long) are identified using these techniques, individual peptides can be synthesized using automated peptide synthesis

equipment (available from manufacturers such as Perkin Elmer/Applied Biosystems Division, Foster City, CA) and techniques such as Merrifield synthesis. Following synthesis, the peptides can be used to screen sera harvested from either normal or breast cancer patients to determine whether patients with breast cancer possess antibodies reactive with the peptides. Presence of such antibodies in breast cancer patient would confirm the immunogenicity of the specific B-cell epitope in question. The peptides can also be tested for their ability to generate a serologic or humoral immune in animals (mice, rats, rabbits, chimps etc.) following immunization *in vivo*. Generation of a peptide-specific antiserum following such immunization further confirms the immunogenicity of the specific B-cell epitope in question.

To identify T-cell epitopes, the B18Ag1 sequence can be screened using different computer algorithms which are useful in identifying 8-10 amino acid motifs within the B18Ag1 sequence which are capable of binding to HLA Class I MHC molecules. (see, e.g., Rammensee et al., *Immunogenetics* 41:178-228 (1995)). Following synthesis such peptides can be tested for their ability to bind to class I MHC using standard binding assays (e.g., Sette et al., *J. Immunol.* 153:5586-92 (1994)) and more importantly can be tested for their ability to generate antigen reactive cytotoxic T-cells following *in vitro* stimulation of patient or normal peripheral mononuclear cells using, for example, the methods of Bakker et al., *Cancer Res.* 55:5330-34 (1995); Visseren et al., *J. Immunol.* 154:3991-98 (1995); Kawakami et al., *J. Immunol.* 154:3961-68 (1995); and Kast et al., *J. Immunol.* 152:3904-12 (1994). Successful *in vitro* generation of T-cells capable of killing autologous (bearing the same Class I MHC molecules) tumor cells following *in vitro* peptide stimulation further confirms the immunogenicity of the B18Ag1 antigen. Furthermore, such peptides may be used to generate murine peptide and B18Ag1 reactive cytotoxic T-cells following *in vivo* immunization in mice rendered transgenic for expression of a particular human MHC Class I haplotype (Vitiello et al., *J. Exp. Med.* 173:1007-15 (1991)).

A representative list of predicted B18Ag1 B-cell and T-cell epitopes, broken down according to predicted HLA Class I MHC binding antigen, is shown below:

Predicted Th Motifs (B-cell epitopes) (SEQ ID NOS.: 131-133)

SSGGRTFDDFHRYLLVGI
QGAAQKPINLSKXIEVVQGHDE
SPGVFLEHLQEAYRIYTPFDLSA

5

Predicted HLA A2.1 Motifs (T-cell epitopes) (SEQ ID NOS.: 134-140)

YLLVGIQGA
GAAQKPINL
10 NLSKXIEVV
EVVQGHDES
HLQEAYRIY
NLAFVAQAA
FVAQAAPDS

15

EXAMPLE 5

IDENTIFICATION OF T-CELL EPITOPES OF B11Ag1

This Example illustrates the identification of B11Ag1 (also referred to as B305D) epitopes. Four peptides, referred to as B11-8, B11-1, B11-5 and B11-12 (SEQ ID NOs:309-312, respectfully) were derived from the B11Ag1 gene.

Human CD8 T cells were primed *in vitro* to the peptide B11-8 using dendritic cells according to the protocol of Van Tsai et al. (*Critical Reviews in Immunology* 18:65-75, 1998). The resulting CD8 T cell cultures were tested for their ability to recognize the B11-8 peptide or a negative control peptide, presented by the B-LCL line, JY. Briefly, T cells were incubated with autologous monocytes in the presence of 10 ug/ml peptide, 10 ng/ml IL-7 and 10 ug/ml IL-2, and assayed for their ability to specifically lyse target cells in a standard 51-Cr release assay. As shown in Fig. 22, the bulk culture line demonstrated strong recognition of the B11-8 peptide with weaker recognition of the peptide B11-1.

A clone from this CTL line was isolated following rapid expansion using the monoclonal antibody OKT3 and human IL-2. As shown in Fig. 23, this clone (referred to as A1), in addition to being able to recognize specific peptide, recognized JY LCL transduced with the B11Ag1 gene. This data demonstrates that B11-8 is a naturally processed epitope of the B11Ag1 gene. In addition these T cells were further found to recognize and lyse, in an HLA-A2 restricted manner, an established tumor cell line naturally expressing B11Ag1 (Fig. 24). The T cells strongly recognize a lung adenocarcinoma (LT-140-22) naturally expressing B11Ag1 transduced with HLA-A2, as well as an A2+ breast carcinoma (CAMA-1) transduced with B11Ag1, but not 10 untransduced lines or another negative tumor line (SW620).

These data clearly demonstrate that these human T cells recognize not only B11-specific peptides but also transduced cells, as well as naturally expressing tumor lines.

CTL lines raised against the antigens B11-5 and B11-12, using the procedures described above, were found to recognize corresponding peptide-coated targets.

15

EXAMPLE 6
CHARACTERIZATION OF BREAST TUMOR GENES DISCOVERED BY
DIFFERENTIAL DISPLAY PCR

5 The specificity and sensitivity of the breast tumor genes discovered by differential display PCR were determined using RT-PCR. This procedure enabled the rapid evaluation of breast tumor gene mRNA expression semiquantitatively without using large amounts of RNA. Using gene specific primers, mRNA expression levels in a variety of tissues were examined, including 8 breast tumors, 5 normal breasts, 2 prostate tumors, 2
10 colon tumors, 1 lung tumor, and 14 other normal adult human tissues, including normal prostate, colon, kidney, liver, lung, ovary, pancreas, skeletal muscle, skin, stomach and testes.

15 To ensure the semiquantitative nature of the RT-PCR, β -actin was used as internal control for each of the tissues examined. Serial dilutions of the first strand cDNAs were prepared and RT-PCR assays performed using β -actin specific primers. A dilution was then selected that enabled the linear range amplification of β -actin template, and which was sensitive enough to reflect the difference in the initial copy number. Using this condition, the β -actin levels were determined for each reverse transcription reaction from each tissue. DNA contamination was minimized by DNase treatment and by assuring a
20 negative result when using first strand cDNA that was prepared without adding reverse transcriptase.

Using gene specific primers, the mRNA expression levels were determined in a variety of tissues. To date, 38 genes have been successfully examined by RT-PCR, five of which exhibit good specificity and sensitivity for breast tumors (B15AG-1,
25 B31GA1b, B38GA2a, B11A1a and B18AG1a). Figures 21A and 21B depict the results for three of these genes: B15AG-1 (SEQ ID NO:27), B31GA1b (SEQ ID NO:148) and B38GA2a (SEQ ID NO:157). Table I summarizes the expression level of all the genes tested in normal breast tissue and breast tumors, and also in other tissues.

TABLE I
Percentage of Breast Cancer Antigens that are Expressed in Various Tissues

5	Breast Tissues	Over-expressed in Breast Tumors	84%
		Equally Expressed in Normals and Tumor	16%
10			
		Over-expressed in Breast Tumors but not in any Normal Tissues	9%
15	Other Tissues	Over-expressed in Breast Tumors but Expressed in Some Normal Tissues	30%
		Over-expressed in Breast Tumors but Equally Expressed in All Other Tissues	61%

20 EXAMPLE 7

PREPARATION AND CHARACTERIZATION OF ANTIBODIES AGAINST BREAST TUMOR
POLYPEPTIDES

25 Polyclonal antibodies against the breast tumor antigen B305D were prepared as follows.

The breast tumor antigen expressed in an *E. coli* recombinant expression system was grown overnight in LB broth with the appropriate antibiotics at 37 °C in a shaking incubator. The next morning, 10 ml of the overnight culture was added to 500 ml to 2x YT plus appropriate antibiotics in a 2L-baffled Erlenmeyer flask. When the Optical Density (at 560 nm) of the culture reached 0.4-0.6, the cells were induced with IPTG (1 mM). Four hours after induction with IPTG, the cells were harvested by centrifugation. The cells were then washed with phosphate buffered saline and centrifuged again. The supernatant was discarded and the cells were either frozen for future use or immediately processed. Twenty ml of lysis buffer was added to the cell pellets and vortexed. To break

open the *E. coli* cells, this mixture was then run through the French Press at a pressure of 16,000 psi. The cells were then centrifuged again and the supernatant and pellet were checked by SDS-PAGE for the partitioning of the recombinant protein. For proteins that localized to the cell pellet, the pellet was resuspended in 10 mM Tris pH 8.0, 1% CHAPS
5 and the inclusion body pellet was washed and centrifuged again. This procedure was repeated twice more. The washed inclusion body pellet was solubilized with either 8 M urea or 6 M guanidine HCl containing 10 mM Tris pH 8.0 plus 10 mM imidazole. The solubilized protein was added to 5 ml of nickel-chelate resin (Qiagen) and incubated for 45 min to 1 hour at room temperature with continuous agitation. After incubation, the resin
10 and protein mixture were poured through a disposable column and the flow through was collected. The column was then washed with 10-20 column volumes of the solubilization buffer. The antigen was then eluted from the column using 8M urea, 10 mM Tris pH 8.0 and 300 mM imidazole and collected in 3 ml fractions. A SDS-PAGE gel was run to determine which fractions to pool for further purification.

15 As a final purification step, a strong anion exchange resin such as HiPrepQ (Biorad) was equilibrated with the appropriate buffer and the pooled fractions from above were loaded onto the column. Antigen was eluted off the column with a increasing salt gradient. Fractions were collected as the column was run and another SDS-PAGE gel was run to determine which fractions from the column to pool. The pooled fractions were
20 dialyzed against 10 mM Tris pH 8.0. The protein was then vialled after filtration through a 0.22 micron filter and the antigens were frozen until needed for immunization.

Four hundred micrograms of B305D antigen was combined with 100 micrograms of muramyl dipeptide (MDP). Every four weeks rabbits were boosted with 100 micrograms mixed with an equal volume of Incomplete Freund's Adjuvant (IFA). Seven
25 days following each boost, the animal was bled. Sera was generated by incubating the blood at 4 °C for 12-24 hours followed by centrifugation.

Ninety-six well plates were coated with B305D antigen by incubating with 50 microliters (typically 1 microgram) of recombinant protein at 4 °C for 20 hours. 250 microliters of BSA blocking buffer was added to the wells and incubated at room

temperature for 2 hours. Plates were washed 6 times with PBS/0.01% Tween. Rabbit sera was diluted in PBS. Fifty microliters of diluted sera was added to each well and incubated at room temperature for 30 min. Plates were washed as described above before 50 microliters of goat anti-rabbit horse radish peroxidase (HRP) at a 1:10000 dilution was 5 added and incubated at room temperature for 30 min. Plates were again washed as described above and 100 microliters of TMB microwell peroxidase substrate was added to each well. Following a 15 min incubation in the dark at room temperature, the colorimetric reaction was stopped with 100 microliters of 1N H₂SO₄ and read immediately at 450 nm. The polyclonal antibodies showed immunoreactivity to B305D.

10 Immunohistochemical (IHC) analysis of B305D expression in breast cancer and normal breast specimens was performed as follows. Paraffin-embedded formal fixed tissue was sliced into 8 micron sections. Steam heat induced epitope retrieval (SHIER) in 0.1 M sodium citrate buffer (pH 6.0) was used for optimal staining conditions. Sections were incubated with 10% serum/PBS for 5 minutes. Primary antibody was added to each 15 section for 25 min at indicated concentrations followed by a 25 min incubation with either an anti-rabbit or anti-mouse biotinylated antibody. Endogenous peroxidase activity was blocked by three 1.5 min incubations with hydrogen peroxide. The avidin biotin complex/horseradish peroxidase (ABC/HRP) systems was used along with DAB chromagen to visualize antigen expression. Slides were counterstained with hematoxylin.

20 B305D expression was detected in both breast tumor and normal breast tissue. However, the intensity of staining was much less in normal samples than in tumor samples and surface expression of B305D was observed only in breast tumor tissues.

25 A summary of real-time PCR and immunohistochemical analysis of B305D expression in an extensive panel of normal tissues is presented in Table II below. These results demonstrate minimal expression of B305D in testis, inconclusive results in gall bladder, and no detection in all other tissues tested.

TABLE II

mRNA	IHC staining	Tissue type	Summary
Moderately positive	Positive	Testis	Nuclear staining of small minority of spermatids; spermatozoa negative; siminoma negative
Negative	Negative	Thymus	No expression
N/A	Negative	Artery	No expression
Negative	Negative	Skeletal muscle	No expression
Negative	Positive (weak staining)	Small bowel	No expression
Negative	Positive (weak staining)	Ovary	No expression
Negative		Pituitary	No expression
Negative	Positive (weak staining)	Stomach	No expression
Negative	Negative	Spinal cord	No expression
Negative	Negative	Spleen	No expression
Negative	Negative	Ureter	No expression
N/A	Negative	Gall bladder	Inconclusive
N/A	Negative	Placenta	No expression
Negative	Negative	Thyroid	No expression
Negative	Negative	Heart	No expression
Negative	Negative	Kidney	No expression
Negative	Negative	Liver	No expression
Negative	Negative	Brain-cerebellum	No expression
Negative	Negative	Colon	No expression
Negative	Negative	Skin	No expression
Negative	Negative	Bone marrow	No expression
N/A	Negative	Parathyroid	No expression
Negative	Negative	Lung	No expression
Negative	Negative	Esophagus	No expression
Negative	Positive (weak staining)	Uterus	No expression
Negative	Negative	Adrenal	No expression
Negative	Negative	Pancreas	No expression
N/A	Negative	Lymph node	No expression
Negative	Negative	Brain-cortex	No expression
N/A	Negative	Fallopian tube	No expression
Negative	Positive (weak staining)	Bladder	No expression
Negative	N/A	Bone	No expression
Negative	N/A	Salivary gland	No expression
Negative	N/A	Activated PBMC	No expression
Negative	N/A	Resting PBMC	No expression

Negative	N/A	Trachea	No expression
Negative	N/A	Vena cava	No expression
Negative	N/A	Retina	No expression
Negative	N/A	Cartilage	No expression

EXAMPLE 8

PROTEIN EXPRESSION OF BREAST TUMOR ANTIGENS

5 This example describes the expression and purification of the breast tumor antigen B305D in *E. coli* and in mammalian cells.

Expression of B305D isoform C-15 (SEQ ID NO:301; translated to 384 amino acids) in *E. coli* was achieved by cloning the open reading frame of B305D isoform C-15 downstream of the first 30 amino acids of the *M. tuberculosis* antigen Ra12 (SEQ ID 10 NO:318) in pET17b. First, the internal EcoRI site in the B305D ORF was mutated without changing the protein sequence so that the gene could be cloned at the EcoRI site with Ra12. The PCR primers used for site-directed mutagenesis are shown in SEQ ID NO:319 (referred to as AW012) and SEQ ID NO:320 (referred to as AW013). The ORF of EcoRI site-modified B305D was then amplified by PCR using the primers AW014 (SEQ ID 15 NO:321) and AW015 (SEQ ID NO:322). The PCR product was digested with EcoRI and ligated to the Ra12/pET17b vector at the EcoRI site. The sequence of the resulting fusion construct (referred to as Ra12mB11C) was confirmed by DNA sequencing. The determined cDNA sequence for the fusion construct is provided in SEQ ID NO:323, with the amino acid sequence being provided in SEQ ID NO:324.

20 The fusion construct was transformed into BL21(DE3)CodonPlus-RIL *E. coli* (Stratagene) and grown overnight in LB broth with kanamycin. The resulting culture was induced with IPTG. Protein was transferred to PVDF membrane and blocked with 5% non-fat milk (in PBS-Tween buffer), washed three times and incubated with mouse anti-His tag antibody (Clontech) for 1 hour. The membrane was washed 3 times and probed 25 with HRP-Protein A (Zymed) for 30 min. Finally, the membrane was washed 3 times and developed with ECL (Amersham). Expression was detected by Western blot.

For recombinant expression in mammalian cells, B305D isoform C-15 (SEQ ID NO:301; translated to 384 amino acids) was subcloned into the mammalian expression vectors pCEP4 and pcDNA3.1 (Invitrogen). These constructs were transfected into HEK293 cells (ATCC) using Fugene 6 reagent (Roche). Briefly, the HEK cells were plated at a density of 100,000 cells/ml in DMEM (Gibco) containing 10% FBS (Hyclone) and grown overnight. The following day, 2 ul of Fugene 6 was added to 100 ul of DMEM containing no FBS and incubated for 15 minutes at room temperature. The Fugene 6/DMEM mixture was added to 1 ug of B305D/pCEP4 or B305D/pcDNA plasmid DNA and incubated for 15 minutes at room temperature. The Fugene/DNA mix was then added to the HEK293 cells and incubated for 48-72 hours at 37 °C with 7% CO₂. Cells were rinsed with PBS, the collected and pelleted by centrifugation.

For Western blot analysis, whole cell lysates were generated by incubating the cells in Triton-X100 containing lysis buffer for 30 minutes on ice. Lysates were then cleared by centrifugation at 10,000 rpm for 5 minutes at 4 °C. Samples were diluted with SDS_PAGE loading buffer containing beta-mercaptoethanol, and boiled for 10 minutes prior to loading the SDS_PAGE gel. Proteins were transferred to nitrocellulose and probed using Protein A purified anti-B305D rabbit polyclonal sera (prepared as described above) at a concentration of 1 ug/ml. The blot was revealed with a goat anti-rabbit Ig coupled to HRP followed by incubation in ECL substrate. Expression of B305D was detected in the HEK293 lysates transfected with B305D, but not in control HEK293 cells transfected with vector alone.

For FACS analysis, cells were washed further with ice cold staining buffer and then incubated with a 1:100 dilution of a goat anti-rabbit Ig (H+L)-FITC reagent (Southern Biotechnology) for 30 minutes on ice. Following 3 washes, the cells were resuspended in staining buffer containing Propidium Iodide (PI), a vital stain that allows for identification of permeable cells, and then analyzed by FACS. The FACS analysis showed surface expression of B305D protein.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

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CLAIMS

What is Claimed:

1. An isolated polynucleotide comprising a sequence selected from the group consisting of:
 - (a) sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (b) complements of the sequences provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (d) sequences that hybridize to a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325, under moderately stringent conditions;
 - (e) sequences having at least 75% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325;
 - (f) sequences having at least 90% identity to a sequence of SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325; and
 - (g) degenerate variants of a sequence provided in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325.
2. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:
 - (a) sequences encoded by a polynucleotide of claim 1; and
 - (b) sequences having at least 70% identity to a sequence encoded by a polynucleotide of claim 1; and

(c) sequences having at least 90% identity to a sequence encoded by a polynucleotide of claim 1.

3. An expression vector comprising a polynucleotide of claim 1 operably linked to an expression control sequence.

4. A host cell transformed or transfected with an expression vector according to claim 3.

5. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a polypeptide of claim 2.

6. A method for detecting the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with a binding agent that binds to a polypeptide of claim 2;
- (c) detecting in the sample an amount of polypeptide that binds to the binding agent; and
- (d) comparing the amount of polypeptide to a predetermined cut-off value and therefrom determining the presence of a cancer in the patient.

7. A fusion protein comprising at least one polypeptide according to claim 2.

8. An oligonucleotide that hybridizes to a sequence recited in SEQ ID NO: 1, 3-86, 142-298, 301-303, 307, 313, 314, 316, 317 and 325 under moderately stringent conditions.

9. A method for stimulating and/or expanding T cells specific for a tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1; and
- (c) antigen-presenting cells that express a polypeptide according to claim 1,

under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

10. An isolated T cell population, comprising T cells prepared according to the method of claim 9.

11. A composition comprising a first component selected from the group consisting of physiologically acceptable carriers and immunostimulants, and a second component selected from the group consisting of:

- (a) polypeptides according to claim 2;
- (b) polynucleotides according to claim 1;
- (c) antibodies according to claim 5;
- (d) fusion proteins according to claim 7;
- (e) T cell populations according to claim 10; and
- (f) antigen presenting cells that express a polypeptide according to claim 2.

12. A method for stimulating an immune response in a patient, comprising administering to the patient a composition of claim 11.

13. A method for the treatment of a cancer in a patient, comprising administering to the patient a composition of claim 11.

14. A method for determining the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with an oligonucleotide according to claim 8;
- (c) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and
- (d) compare the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence of the cancer in the patient.

15. A diagnostic kit comprising at least one oligonucleotide according to claim 8.

16. A diagnostic kit comprising at least one antibody according to claim 5 and a detection reagent, wherein the detection reagent comprises a reporter group.

17. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

- (a) incubating CD4+ and/or CD8+ T cells isolated from a patient with at least one component selected from the group consisting of: (i) polypeptides according to claim 2; (ii) polynucleotides according to claim 1; and (iii) antigen presenting cells that express a polypeptide of claim 2, such that T cell proliferate;
- (b) administering to the patient an effective amount of the proliferated T cells,

and thereby inhibiting the development of a cancer in the patient.

COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF
BREAST CANCER

ABSTRACT OF THE DISCLOSURE

Compositions and methods for the therapy and diagnosis of cancer, particularly breast cancer, are disclosed. Illustrative compositions comprise one or more breast tumor polypeptides, immunogenic portions thereof, polynucleotides that encode such polypeptides, antigen presenting cell that expresses such polypeptides, and T cells that are specific for cells expressing such polypeptides. The disclosed compositions are useful, for example, in the diagnosis, prevention and/or treatment of diseases, particularly breast cancer.

WPN\210121 - corixa\419c10\419c10-app.doc

cDNA PREPARED FROM
NORMAL BREAST TISSUE
FROM THE SAME PATIENT

cDNA PREPARED
FROM BREAST TUMOR

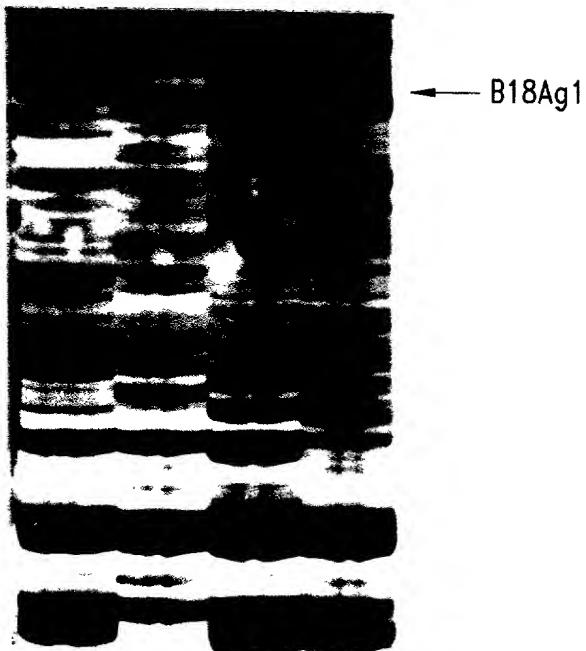


Fig. 1

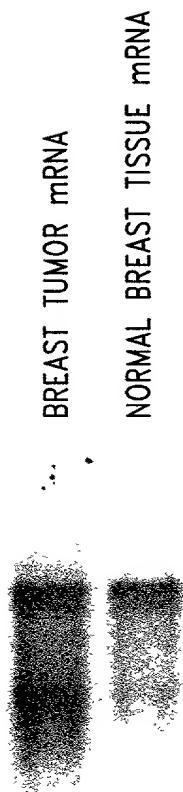


Fig. 2

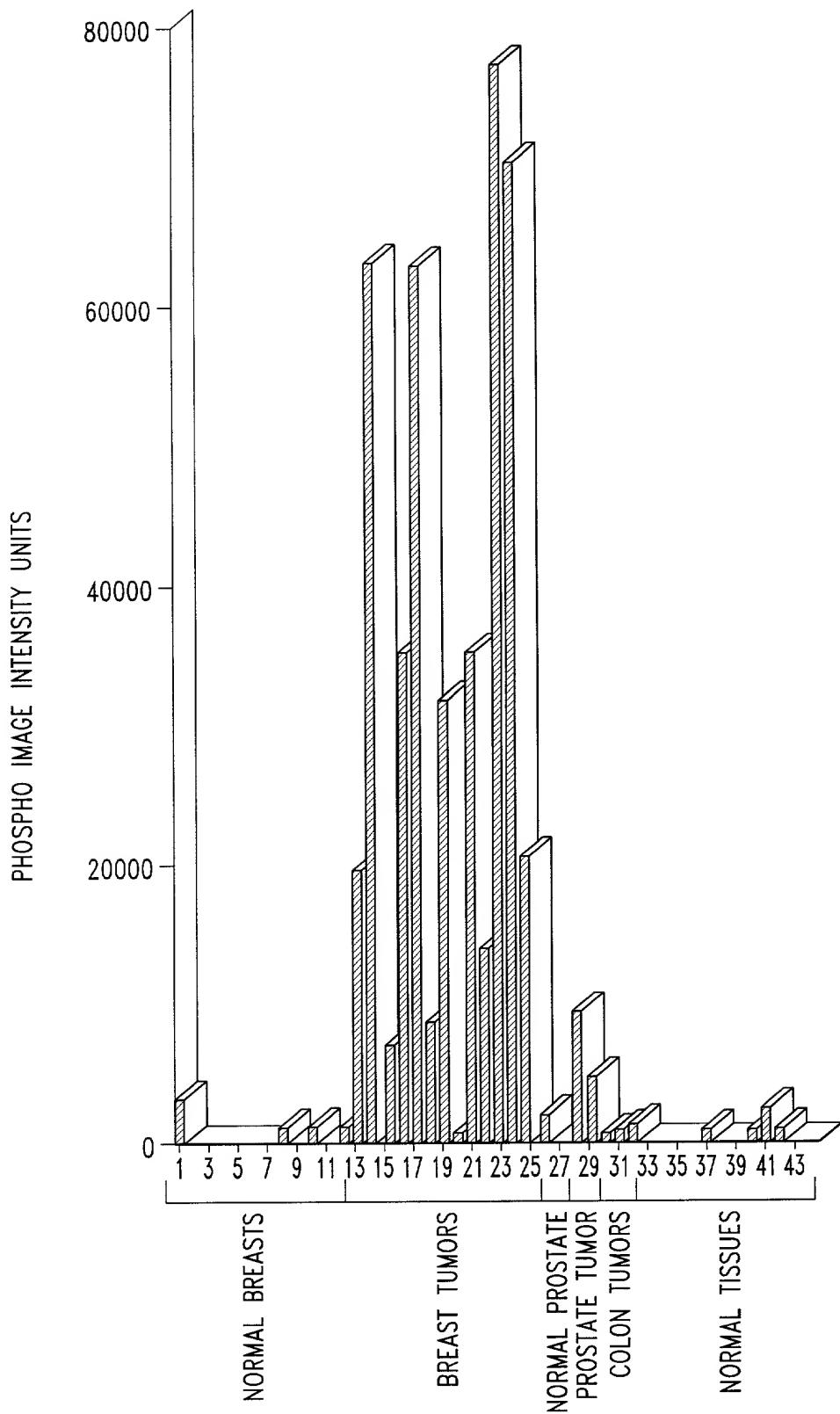


Fig. 3

GENOMIC CLONE MAP

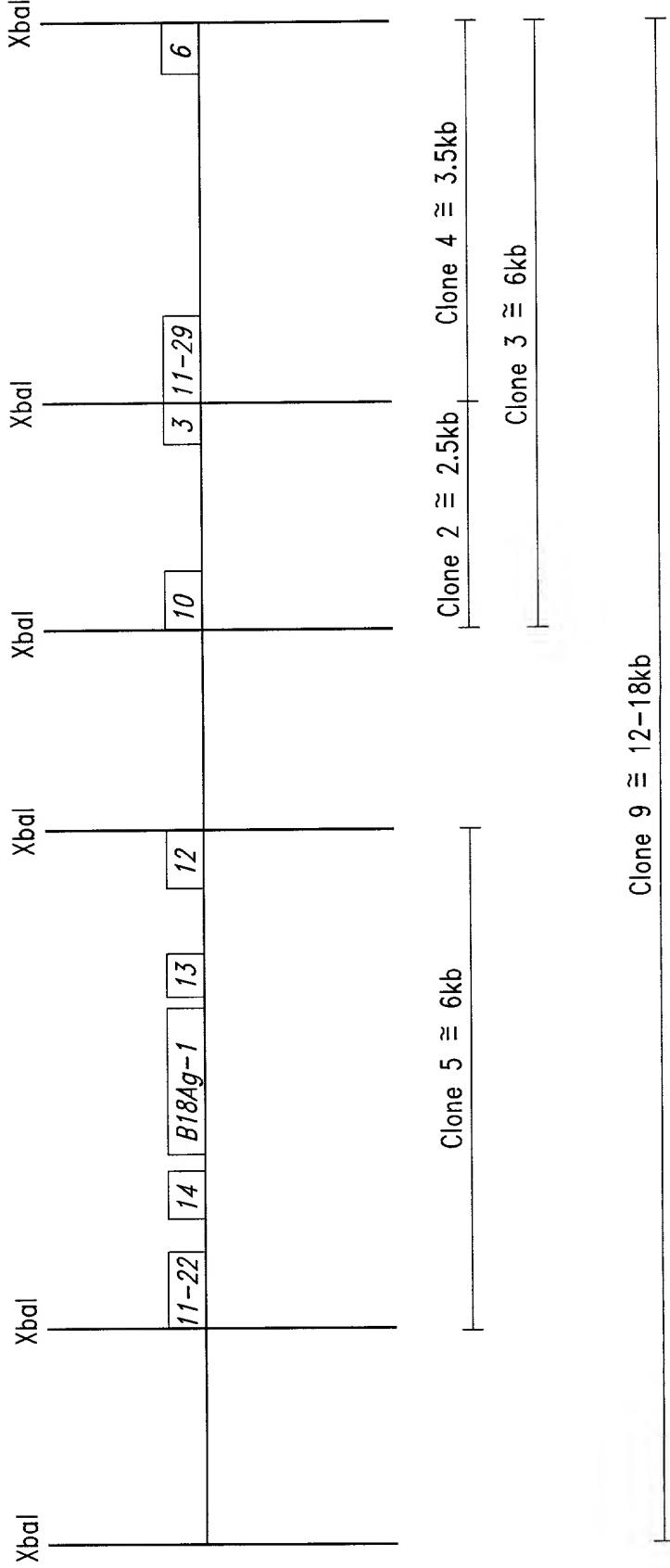


Fig. 4

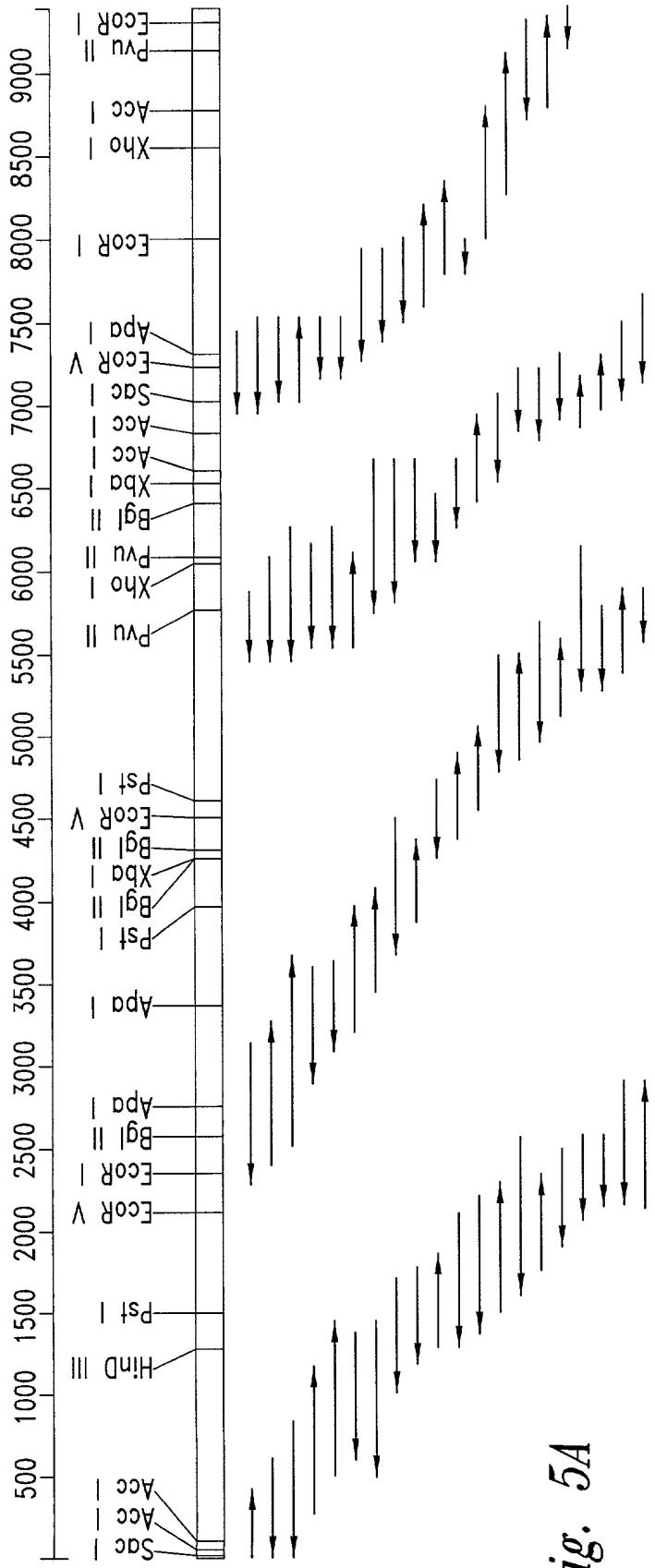


Fig. 5A

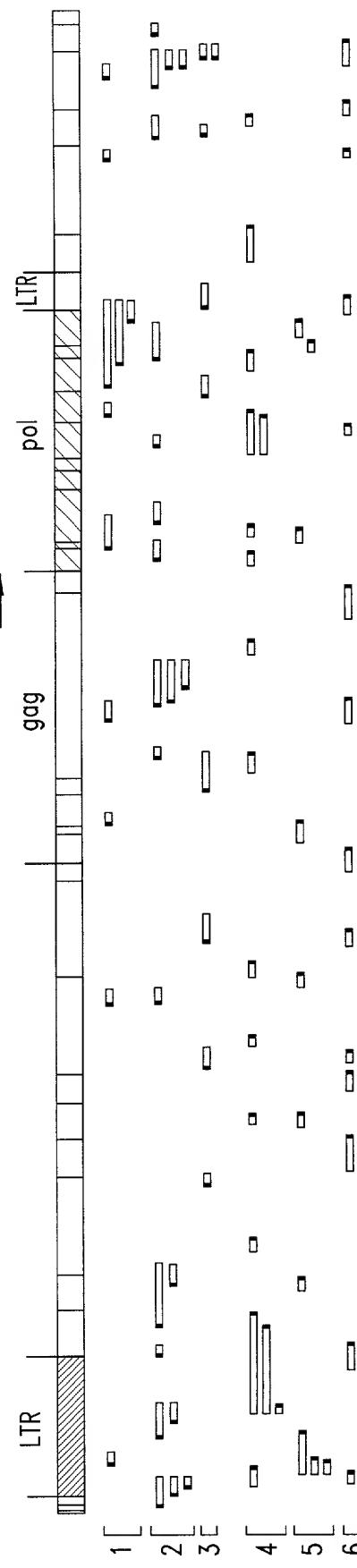


Fig. 5B

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B18Ag1

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Gly Arg Thr Phe Asp Asp Phe His Arg Tyr Leu Leu Val Gly Ile Gln	
20 25 30	
GGA GCT GCC CAG AAA CCT ATA AAC TTG TCT AAG GCG ATT GAA GTC GTC	144
Gly Ala Ala Gln Lys Pro Ile Asn Leu Ser Lys Ala Ile Glu Val Val	
35 40 45	
CAG GGG CAT GAT GAG TCA CCA GGA GTG TTT TTA GAG CAC CTC CAG GAG	192
Gln Gly His Asp Glu Ser Pro Gly Val Phe Leu Glu His Leu Gln Glu	
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Ala Tyr Arg Ile Tyr Thr Pro Phe Asp Leu Ala Ala Pro Glu Asn Ser	
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CAT GCT CTT AAT TTG GCA TTT GTG GCT CAG GCA GCC CCA GAT AGT AAA	288
His Ala Leu Asn Leu Ala Phe Val Ala Gln Ala Ala Pro Asp Ser Lys	
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Arg Lys Leu Gln Lys Leu Glu Gly Phe Cys Trp Asn Glu Tyr Gln Ser	
100 105 110	
GCT TTT AGA GAT AGC CTA AAA GGT TTT	363
Ala Phe Arg Asp Ser Leu Lys Gly Phe	
115 120	

Fig. 6

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B17Ag1

GC TGGGCACAGT GGCTCATACC TGTAATCCTG ACCGTTTCAG AGGCTCAGGT	60
CG CTTGAGGCCA AGATTCAAG ACTAGTCTGG GTAACATAGT GAGACCCTAT	120
AA AAATAAAAAAA ATGAGCCTGG TGTAGTGGCA CACACCAGCT GAGGAGGGAG	180
CT AGGAGA	196

Fig. 7

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B17Ag2

GC TTGGGGGCTC TGACTAGAAA TTCAAGGAAC CTGGGATTCA AGTCCAAC TG	60
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CA AGAAGACTGA ACAGTACTAC TGTGAAAAGC CCGAAGNGGC AATATGTTCA	300
TT GAAGGATGGC TGGGAGAATG AATGCTCTGT CCCCCAGTCC CAAGCTCACT	360
CT CCTTATAGC CTAGGAGA	388

Fig. 8

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag2 α

GC CTATAATCAT GTTTCTCATT ATTTCACAT TTTATTAACC AATTCTGTT	60
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CT ATTTTTCCA TATTTGGGCA ACTACTA	337

Fig. 9

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag1b

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GC TGGTGTTTT TACTGTAAAC AATAAGGAGA CTTTGCTCTT CATTAAACC	120
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TT TAAGTCGTTT GGAACAAGAT ATTTTTCTT TCCTGGCAGC TTTAACATT	240
TT TGTGTCTGGG GGACTGCTGG TCACTGTTTC TCACAGTTGC AAATCAAGGC	300
CC AAGAAAAAAA AATTTTTTG TTTTATTGA AACTGGACCG GATAAACGGT	360
CG GCTGCTGTAT ATAGTTTAA ATGGTTTATT GCACCTCCTT AAGTTGCACT	420
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NA CTGAGCTAAA AAGGGCTGNT TTTCGGGTGG GGGCAGATGA AGGCTCACAG	540
TC TCTTAGAGGG GGGAACTNCT A	571

Fig. 10

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B13Ag1a

TA ATAACCTAAA TATATTTGA TCACCCACTG GGGTGATAAG ACAATAGATA	60
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	548

Fig. 11

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B11Ag1

TG CACATGCAGA ATATTCTATC GGTACTTCAG CTATTACTCA TTTTGATGGC	60
AG CCTATCCTCA AGATGAGTAT TTAGAAAGAA TTGATTTAGC GATAGACCAA	120
GC ACTCTGACTA CACGAAATTG TTCAGATGTG ATGGATTTAT GACAGTTGAT	180
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Fig. 12

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA3c

ACTGATGGAT GTCGCCGGAG GCGAGGGGCC TTATCTGATG CTCGGCTGCC TGTTCGTGAT	60
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GATGGCGGCT TCGGC GGCGT TTATGACCCC GGTCTCCTCG CCGGTTAACCA CCCTGGTGCT	240
TGGCCCTGGC AAGTACTCAT TTAGCGATT TGTCAAAATA GGCGTG	286

Fig. 13

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B9CG1

AG CAGCCCCCTTC TTCTCAATT CATCTGTCAC TACCCTGGTG TAGTATCTCA	60
CA TTTTTATAGC CTCCCTCCCTG GTCTGTCTTT TGATTTCT GCCTGTAATC	120
AC ATAAC TGCAA GTAAACATTT CTAAAGTG TG GTTATGCTCA TGTCACTCCT	180
AA ATAGTTCCA TTACCGTCTT AATAAAATTC GGATTGTTG TTTNCTATTN	240
CA CCTATGACCG AA	262

Fig. 14

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B9CG3

AG CAAAGCCAGT GGTTTGAGCT CTCTACTGTG TAAACTCCTA AACCAAGGCC	60
TA AATGGTGGCA GGATTTTAT TATAAACATG TACCCATGCA AATTTCCTAT	120
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AG TTAGGTAAGA GTGTTAATG AGAGGGTATA AGGTATAAAT CACCAGTCAA	240
TG CCTATGACCG A	261

Fig. 15

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B2CA2

GG GCATGGACGC AGACGCCCTGA CGTTTGGCTG AAAATCTTC ATTGATTCGT	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCTGT TGCTGCCAG TTTTTNTGTT	120
GG ANAAGGCAAN GAGCTCTTCA GACTATTGGN ATTNTCGTTC GGTCTTCTGC	180
CG NCTTGCNANG ATCTTCAT	208

Fig. 16

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA1

GG GCATGGACGC AGACGCCCTGA CGTTTGGCTG AAAATCTTTC ATTGATTCGT	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCCTGT TGCTCGCCAG TTTTTNTGTT	120
GG ANAAGGCAAN GAGCTCTTCA GACTATTGGN ATTNTCGTTC GGTCTTCTGC	180
CG NCTTGCNANG ATCTTCAT	208

Fig. 17

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA2

GG GCATGGACGC AGACGCCCTGA CGTTTGGCTG AAAATCTTTC ATTGATTCGT	60
AT AGGAAAATTC CCAAAGAGGG AATGTCCCTGT TGCTGCCAG TTTTTNTGTT	120
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Fig. 18

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B3CA3

AG GGAGCAAGGA GAAGGCATGG AGAGGCTCAN GCTGGTCCTG GCCTACGACT	60
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Fig. 19

NUCLEOTIDE SEQUENCE OF THE REPRESENTATIVE
BREAST-TUMOR SPECIFIC cDNA B4CA1

TC AGGAGCGGGT AGAGTGGCAC CATTGAGGGG ATATTCAAAA ATATTATTTT	60
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Fig. 20

Fig. 21A

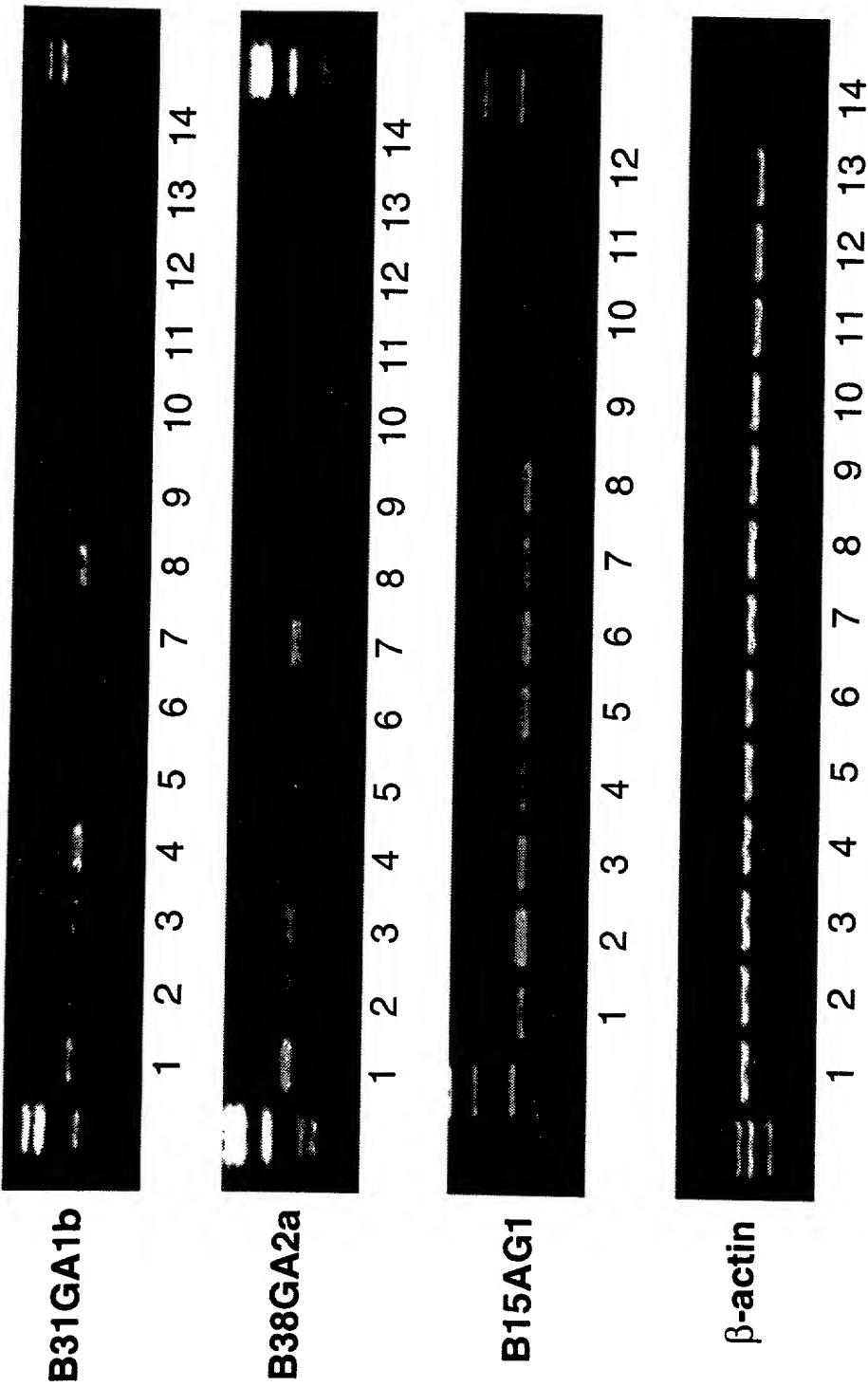


Fig. 21B



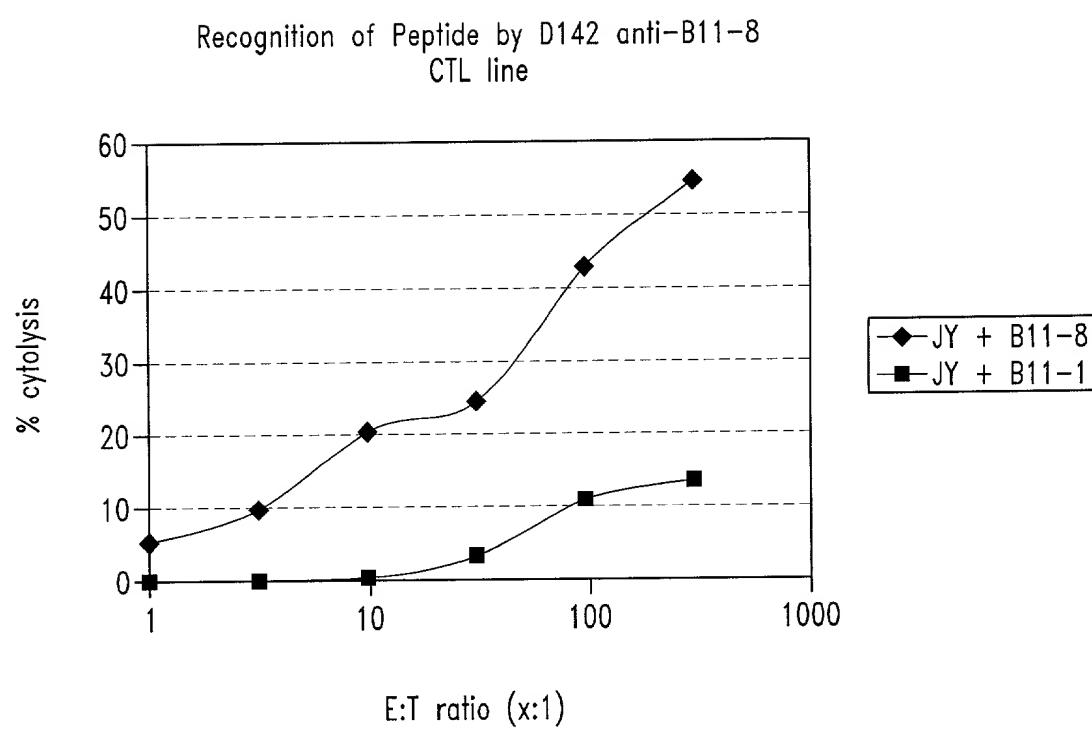


Fig. 22

Recognition of B11 Transductant by B11-8
Specific Clone A1

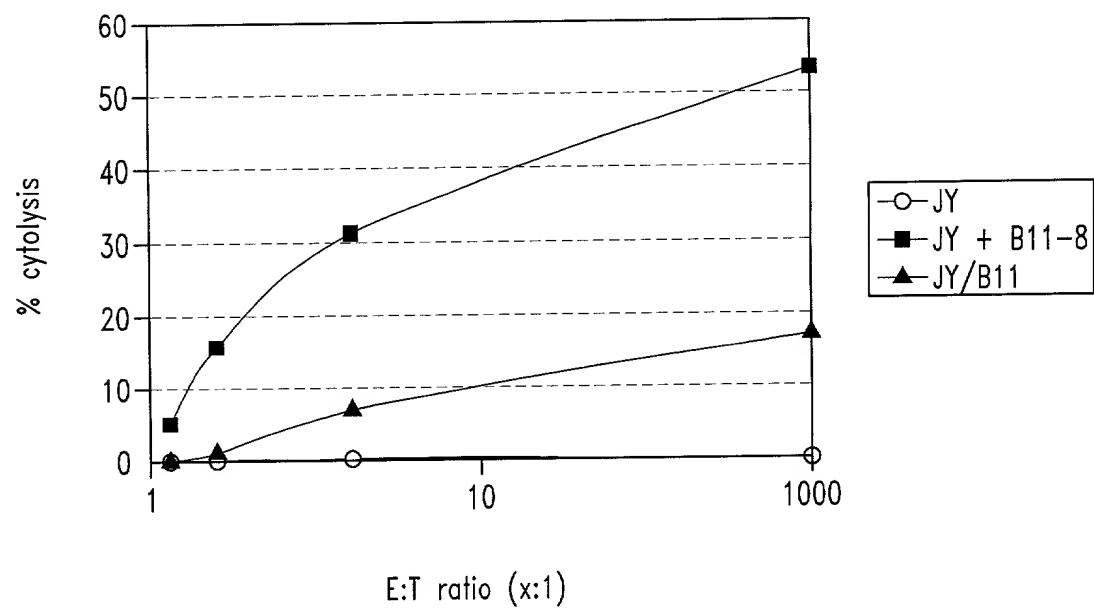


Fig. 23

Recognition of Tumor Cell Lines by Clone A1

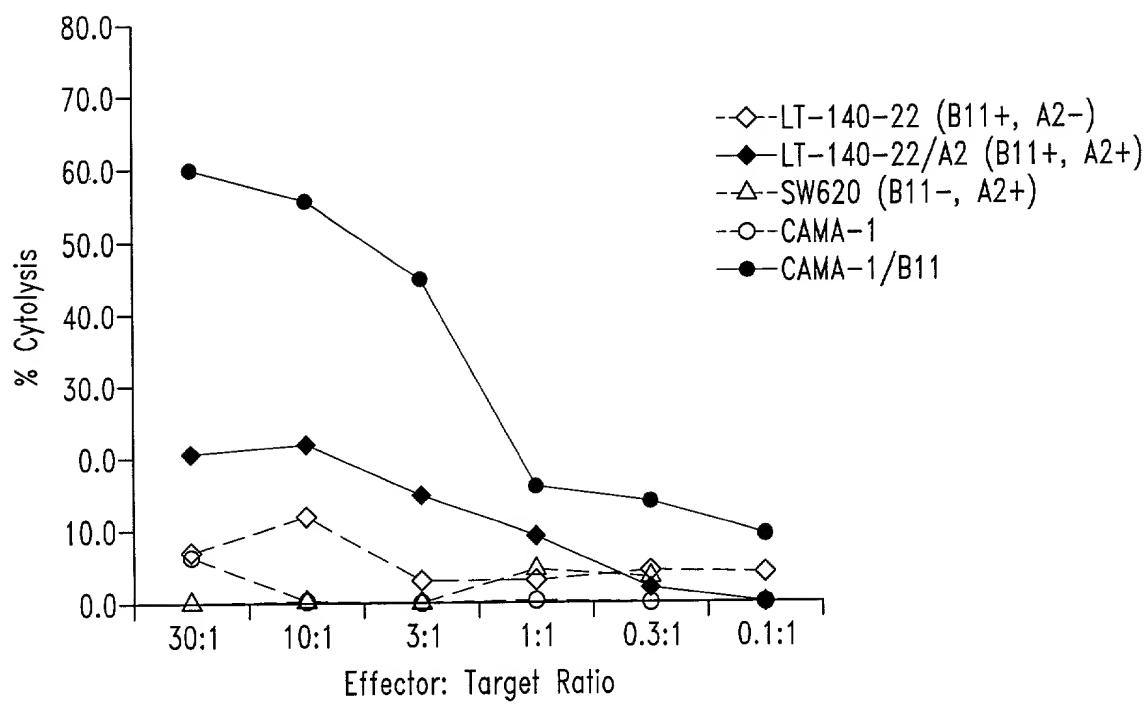


Fig. 24

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicants : Tony N. Frudakis et al.
Filed : October 26, 2000
For : COMPOSITIONS AND METHODS FOR THE THERAPY AND
DIAGNOSIS OF BREAST CANCER
Docket No. : 210121.419C10
Date : October 26, 2000

Box Patent Application
Assistant Commissioner for Patents
Washington, D.C. 20231

DECLARATION

Sir:

I, Monica Steinborn, in accordance with 37 C.F.R. § 1.821(f) do hereby declare that, to the best of my knowledge, the content of the paper entitled "Sequence Listing" and the computer readable copy contained within the floppy disk are the same.

I declare further that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Dated this 26th day of October, 2000.



Monica Steinborn
Biotechnology Paralegal

701 Fifth Avenue, Suite 6300
Seattle, WA 98104-7092
(206) 622-4900
FAX (206) 682-6031

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SEQUENCE LISTING

<110> Frudakis, Tony N.
 Reed, Steven G.
 Smith, John M.
 Misher, Linda E.
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 <222> (1)...(1010)
 <223> n = A,T,C or G

<400> 5

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aggtaacaca catactatct cccaaatacc taccacaaag ctcaacaatt tttaactgtt	180
aggatcactg gctctaattca ccatgacatg aggtcaccac caaaccatca agcgctaaac	240
agacagaatg ttccactcc tgatccactg tgtggaaaga agcaccgaac ttacccactg	300
gggggcctgc ntcanananaa aagcccatgc ccccggtnt nccttnaac cgaaacgaat	360
naacccacca tccccacanc tcctctgtc ntggccctg catctgtgg cctcntntnc	420
tttnggggan acntgggaa ggtacccat ttcnntgacc ccncnanaaa accccngtgg	480
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aatgtcccn gaaccctctc cntnctgccc aaaacctacc taaattntc nctangnntt	600
ttcttgggtgt tnctttcaa aggtacccat ncctgttcan ncccnacnaa aatttttcc	660
ntatnntgn cccnnaaaaa nnatcnnc cnaattgccc gaattggtn ggaaaaatcc	720
nctggggaaa accctttaaa ttccccctt ggcggcccc cctttttcc ccccttngaa	780
aggcaggnggg ttcttcccga acttccatt ncaacagccn tgcccatgtn tgaaaccctt	840
ttccttaaaat taaaaaatccn ccggtnnngg nnggcctt tcccttcng gnnggnngng	900
aaantccta ccccnnaaaaa ggttgcctttag ccccnngtcc ccactccccc ngaaaaatn	960
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<210> 6
 <211> 950
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature

<222> (1)...(950)

<223> n = A,T,C or G

<400> 6

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ctgggattac	aggcggtcaa	caccacaccc	ggctaatttt	gtatTTTAA	tagagatggg	180
gtttccctt	gttggccann	atggtctcna	accctgacc	tcnngtgatc	ccccnccnn	240
nganctnna	ctgctggga	tnnccgnnn	nnncctccn	ncncnnnnnn	ncncnntccn	300
tnntcctnc	tcnnnnnnnn	cnntcnntcc	nncttctcnc	nnnntnttnt	cnncnccnn	360
cnnnccnct	ncccnnnnt	tencntncnn	tntccnnnn	nttcnnnnnn	cnnnnccnnt	420
cnnntacntc	tnnnncnnnt	cntctntnn	cctcnnccnnt	cnctncnent	tntctcctcn	480
tnnnnnnnct	ccnnnnntct	cntcncnnn	tncctcnntn	ncncnccccc	ncctcncnn	540
ctnnttnnn	cnncnntcc	ntncnttcn	nttccnnnn	cnncntcncn	nncttnttc	600
ccnccnnttc	cttncnentn	nnntntcnm	cnctcnntc	ntttntctct	nnntccnn	660
tcnnttcncc	cnnntccncc	ccccnccnt	ctctcncnn	ntnnnnntntn	nnncntccnc	720
tntcncnttc	ntcnntncnt	ntctntemc	nnenntncnc	tncntntnt	ctnntcncn	780
tcnntntcn	ccntccntn	ctntctccn	tntccctccc	ctcncctnct	cnttccncnc	840
cnnntntntn	tnncncnnt	ntcnnncnnc	cntcncntn	tctctnctnn	nnntnnccctc	900
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<210> 7

<211> 1086

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(1086)

<223> n = A,T,C or G

<400> 7

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agaaaaattc	ttctgccttg	agatgcttt	aatctgttaac	cctagcccc	accctgtgt	180
cacagagaca	tgtgtgtgt	tgactcaagg	ttcaatggat	ttagggttat	gtttgtttaa	240
aaaagtgttt	gaagataata	tgcttgtaa	aagtcatcac	cattctctaa	tctcaagtac	300
ccagggacac	aatacactgc	ggaaggccgc	agggacctct	gtctaggaaa	gccaggtatt	360
gtccaagatt	tctcccatg	tgatagcctg	agatatggcc	tcatggaaag	ggtaagacct	420
gactgtcccc	cagccgaca	tcccccagcc	cgacatcccc	cagccgaca	cccggaaagg	480
gtctgtgt	aggaagatta	ntaaaagagg	aaggctttt	gcattgaagt	aagaagaagg	540
ctctgtctcc	tgctcgtccc	tggcaataa	aatgtcttgg	tgttaaaccc	aatgtatgt	600
tctacttact	gagaatagga	aaaaacatcc	ttagggctgg	aggtgagaca	ccctggcgcc	660
atactgtct	ttaatgcacg	agatgttgt	ntaattgca	tccagggcca	nncccttcc	720
ttaacttttt	atganacaaa	aactttgtc	nttttcctg	cgaacctctc	cccttattan	780
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tccaaaacnt	tttcccggtt	gtcccccttc	caaccccgta	cctggccnn	tttctcccc	900
aacntgtccc	ggntcctcn	ttcccncnn	cttcccnngan	aaaaaacc	gtntganggn	960
gccccctcaa	attataacct	ttccnaaaaca	aannggttcn	aaggtggtt	gnntccgg	1020
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ntcccc						1086

<210> 8

<211> 1177

<212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1177)
 <223> n = A,T,C or G

<400> 8

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atgggtttta	aatccagcta	cactactcc	tgactcaaac	tccactattc	ctgttcatga	180
ctgtcaggaa	ctgttgaaa	ctactgaaac	tggccgacct	gatcttcaaa	atgtccccct	240
aggaaagggtg	gatgccaccg	tgttcacaga	cagtaccncc	ttcctcgaga	agggactacg	300
agggggccgtt	gcanctgtta	ccaaggagac	tnatgtttg	tgggctcagg	cttaccanc	360
aaacacacctca	ncncnnaagg	ctgaatttat	cgcctctact	caggctctcg	gatggggtaa	420
gggatattaa	cgttaaacact	gacagcaggt	acgccttgc	tactgtgcat	gtacgtggag	480
ccatctacca	ggagcgtggg	ctactcactc	ggcaggtggc	tgtnatccac	tgtaaangga	540
catcaaagg	aaaacnnngc	tgttgcccgt	ggttaaccana	aanctgaten	ncagctcnaa	600
gatgctgtgt	tgactttcac	tcncncctct	taaacttgct	gcccacantc	tccttccca	660
accagatctg	cctgacaatc	cccatactca	aaaaaaaaan	aanactggcc	ccgaaccnna	720
accaataaaa	acggggangg	tnngtnganc	nncctgaccc	aaaaataatg	gatccccccgg	780
gctcagggaa	ttcaattcan	ccttatcnat	accccaacn	ngnggggggg	ggccngtncc	840
catnccctt	ntattnattc	tttnnccccc	ccccggcnt	cctttttnaa	ctcgtgaaag	900
gaaaaacctg	ncttaccaan	ttatcnccctg	gacntcccc	ttccncggtn	gnntanaaaa	960
aaaagccnc	antccntcc	naaatttgca	cngaaaggna	aggaattttaa	ccttttatttt	1020
ttnntcctt	antttgtnnn	ccccctttta	cccaggcgaa	cngccatcnt	ttaanaaaaaa	1080
aaanagaang	tttatttttc	cttnngaacca	tcccaatana	aancacccgc	nggggaacgg	1140
gnggnaggc	cnctcaccc	cttntgtn	ngggnc			1177

<210> 9
 <211> 1146
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(1146)
 <223> n = A,T,C or G

<400> 9

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agactccatc	agtggaggtca	aagcctgggg	ctttcagag	aagggaggat	tatgggtttt	180
ccaattatac	aagtccagaag	tagaaagaag	ggacataaaac	caggaagggg	gtggagcact	240
catcacccag	agggacttgt	gcctctctca	gtggtagtag	aggggtact	tcctcccacc	300
acgggttgc	ccaagaggca	atgggtgatg	agcctacagg	ggacatancc	gaggagacat	360
ggatgaccc	taagggagta	ggctgggttt	aaggcggtgg	gactgggtga	ggaaactct	420
cctcttcttc	agagagaagc	agtacagggc	gagctgaacc	ggctgaaggt	cgaggcgaaa	480
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atagan	1146
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<211> 545	
<212> DNA	
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ggtagatggc tccacgtaca tgcacagtag caaaggcgta cctgctgtca gtgttaacgt	420
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<210> 11	
<211> 196	
<212> DNA	
<213> Homo sapien	
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aatcgagcct aggaga	196
<210> 12	
<211> 388	
<212> DNA	
<213> Homo sapien	
<220>	
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<222> (1)...(388)	
<223> n = A,T,C or G	
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aataaaaataa ggaaaacgat gtctgtgtat agccaagtca gntatcctaa aaggagatac	180
taagtgcacat taaatatcag aatgtaaaac ctggaaacca ggttcccacg ctggattaa	240
actgacagca agaagactga acagtactac tgtgaaaagc ccgaagnngc aatatgtca	300
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tactataacct ctttatagc cttaggaga	388

<210> 13		
<211> 337		
<212> DNA		
<213> Homo sapien		
<400> 13		
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acaagatatg atttctacat cagatgctct ttcccttcct gtttatttcc ttttatttc	180	
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<210> 14		
<211> 571		
<212> DNA		
<213> Homo sapien		
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<221> misc_feature		
<222> (1)...(571)		
<223> n = A,T,C or G		
<400> 14		
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<210> 15		
<211> 548		
<212> DNA		
<213> Homo sapien		
<220>		
<221> misc_feature		
<222> (1)...(548)		
<223> n = A,T,C or G		
<400> 15		
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aactacta	548
<210> 16	
<211> 638	
<212> DNA	
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<222> (1)...(638)	
<223> n = A,T,C or G	
<400> 16	
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<210> 17	
<211> 286	
<212> DNA	
<213> Homo sapien	
<400> 17	
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tgccttagcg gcggcgaagt caatgggcgt ctcaccctat cttttgccca tgggtggc	180
gatggcggct tcggcggcgt ttatgacccc ggtctcctcg ccggtaaca ccctgggtct	240
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<210> 18	
<211> 262	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(262)	
<223> n = A,T,C or G	
<400> 18	
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<211> 261	
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<400> 19	
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<210> 20	
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<222> (1)...(294)	
<223> n = A,T,C or G	
<400> 20	
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<210> 21	
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<223> n = A,T,C or G	
<400> 21	
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<211> 287	
<212> DNA	
<213> Homo sapien	
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<222> (1)...(287)

<223> n = A,T,C or G

<400> 22

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<210> 23

<211> 204

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(204)

<223> n = A,T,C or G

<400> 23

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<210> 24

<211> 264

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(264)

<223> n = A,T,C or G

<400> 24

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<210> 25

<211> 376

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(376)

<223> n = A,T,C or G

<400> 25

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<210> 26	
<211> 372	
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<223> n = A,T,C or G	
<400> 26	
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ggtaagggtt gcatgagtca tgatcgcc actgcactcc agcctgggtg acagactgag	180
accctgcctc aaaagaaaaa gaataggaag ttcagaaacc ctgggtgtgg ngcccagcaa	240
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gactttttt gggaaaaaat agtcgaaaat gtcaattgg tccataaaat acatgttact	420
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<210> 28	
<211> 438	
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<220>	
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cacaaatgcc aaattaagag catggctatt ttggggct gacaggtaa aagggtgtaa	180
aatccgataa gcctcctgga ggtgctctaa aaacactct ggtgactcat catgcccctg	240
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gaggagatac cggtgaaat cgtcaaaaat tctccctcca cttgagaaat ttgggtcccc	360
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<211> 620	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(620)	
<223> n = A,T,C or G	
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cctaaccacct agatattcag aaaaaatgtt actacaggaa tgaagcttc acggaaaacc	180
tctactagga aagtacagaa gagaatgtt ggtttggagc ccccaaacag aatcccctct	240
agaacactgc ctaatgaaac tgtgagaaga tggccactgt catccagaca ccagaatgtat	300
agacccacca aaaacttatg ccatattgcc tataaaacctt acagacactc aatgccagcc	360
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nangaaatcn ttttaanact tccacggtnn aatgactgcc ctattanatt cngaaacttan	540
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cccatgcctg tnccctctta	620
<210> 30	
<211> 100	
<212> DNA	
<213> Homo sapien	
<400> 30	
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<210> 31	
<211> 762	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(762)	
<223> n = A,T,C or G	
<400> 31	
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acactcttcc tgaaaagaga aagaaaagag gcaggaaaga gtttaggatt tcatttcaa	120
gagtcaagcta attaggagag cagagtttag acagcagtag gcaccccatg atacaacca	180

tggacaaagt ccctgttag taactgccag acatgatcct gctcaggatt taaaatctct	240
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tctcaggag acaagggtat caaaaaacaa gattcttaat gggaaaggaaa tcaaaccaaa	360
aaatttagatt ttctctaca tatataat atacagatat ttaacacatt attccagagg	420
tggctccagt ccttgggct tgagagatgg tgaaaacttt tgttccacat taactctgc	480
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agacccaaat gggtctgtgc cccagaaga gaagccccaa agacatgaag gatgcttaag	600
gggggttggg aaagccaaat tggtantatc ttttcctcct gcctgtgtc cngaaagtctc	660
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<211> 276	
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cacaaccagt aaattggcag agtcagattt gaatccatgg agtctggct gcacttcaa	180
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<210> 33	
<211> 477	
<212> DNA	
<213> Homo sapien	
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caagccccattt atctttttc ccccccgaat ctgaaaattt caggggacag agggaaagtt	360
tcccaattaaa aaattgtaaa tatgttcaat ttatgtttaa aaatgcacaa aacataagaa	420
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<210> 34	
<211> 631	
<212> DNA	
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ggcctgacat ctggaaagg ctcagatcca cttactgctc cttgtctgtt gattgtttt	540
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<210> 35	
<211> 578	
<212> DNA	
<213> Homo sapien	
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actgataccat tgaaacctac ttggagcaga cattgcacag tttctgtgg taaaaactaa	180
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tacataagct tgcttggatc gcctgggtt taaaaggact atcttggcc tcagttcac	420
aagaatgggc aaagtgttgc cttatgttct gtagttctca ataaaagatt gccaggggcc	480
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<210> 36	
<211> 583	
<212> DNA	
<213> Homo sapien	
<400> 36	
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agttagattc catctcaaaa aaaaaaaaaa gaaaaagaaaa agaaaaggaa aaaacgtata	180
aaccaggcca aaacaaaaatg atcattctt taataagcaa gactaattta atgtgtttat	240
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ggttcttact tgggtgaacg tttgatgtc acagttata aaatggtaa caagaaaaat	360
gatgcataaa gaatttata aactactaaa aataaataaa atataaatgg ataggtgcta	420
tggatggagt ttttgttaa tttaaaatct tgaagtgcatt ttggatgctc attgggtgtc	480
tggtaatttc cattagggaaa aggttatgtat atggggaaac tggttctgga aattgcggaa	540
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<210> 37	
<211> 716	
<212> DNA	
<213> Homo sapien	
<220>	
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<222> (1)...(716)	
<223> n = A,T,C or G	
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aatggacca aaactgtat agaaaaatca gaggaagaga ggaacaaata tttactgagt	360
cctagaatgt acaaggcttt ttaattacat attttatgtt aggcctgcaaa aaaacagggt	420

agtaatcaac atttgtccca ttttacatat aaggaaaactg aagcttaaat tgaataattt	480
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<210> 38	
<211> 688	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(688)	
<223> n = A,T,C or G	
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cgagaaattt ctttcccatt tgaataacag ggtggcttga tgggtacggt gggtgaccca	660
acgaagaaaa taaaattctg ccctttcc	688
<210> 39	
<211> 585	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(585)	
<223> n = A,T,C or G	
<400> 39	
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caacatcactc tcactgccc ccagcctgg ggacaggaac aagantcccg tcctcanaaa	480
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<210> 40	
<211> 475	

<212> DNA

<213> Homo sapien

<400> 40

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gaatttcagc acactgagtt gggaaattct tatcccagaa gaccaaccaa ttcatat	180
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gaacaaatac ttccatattt ttttccacca ttgtgggatt ggactttaag aggtgact	300
aaaaaaacag agaacaataa tgtctcagg gtattaagca cggacccata ttatcatat	360
cactaaaaaa aatgatttcc tgtgcaccc ttggcaactt cttttcaa tgttagggaaa	420
aacttagtca ccctgaaaac ccacaaaata aataaaaactt gtatgtgg acaga	475

<210> 41

<211> 423

<212> DNA

<213> Homo sapien

<400> 41

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ttgttttaag atcctgtta gtgcattaa aagtcatgt tatatcaaacc gctctaaaac	360
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cta	423

<210> 42

<211> 527

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(527)

<223> n = A,T,C or G

<400> 42

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<210> 43

<211> 331

<212> DNA

<213> Homo sapien

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<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(908)

<223> n = A,T,C or G

<400> 46

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cttgatgcat	agctttagt	tctatagtg	cactaaatag	cctggcgta	tcatggtcat	840
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cataaaagt						908

<210> 47

<211> 480

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(480)

<223> n = A,T,C or G

<400> 47

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cacagataka	atattacaca	gataaaagag	gagttgatct	aaagtaraga	tagtggggg	180
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ttatTTTAA	aaagtaggtg	acatTTGA	gagagaaaag	ggcttgggt	agatgaagtc	360
cccccccccc	ctttttttt	tttagctga	aatagatacc	ctatgttnaa	rgaarggatt	420
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<210> 48

<211> 591

<212> DNA

<213> Homo sapien

<400> 48

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gaggatggtg	gcgtgaattc	tggccttct	ttgccgtgg	atcgtagcc	gccatcatcg	180
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actccgaaaa	cgtccgggtg	ctgctgtcg	gtgactccca	aatcttgat	aacaacaagg	420
taaccgaatc	gchgctaagg	accccgcat	ctcggtact	ctgcataatgc	gtacccctta	480
agccgaattc	cagcacactg	gcccgcgtt	ctaattggat	ccgaactccg	taaccaagcc	540
tgatgcgtaa	tttgcgttat	tctatagtgt	ccctaaaata	acctggcgtt	a	591
<210> 49						
<211> 454						
<212> DNA						
<213> Homo sapien						
<400> 49						
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aagaaaagctg	ctgtggggaa	aggaggata	aatactgaag	ggatttacta	aacaaatgtc	180
catcacagag	ttttcccttt	ttttttttt	agacagatc	ttgctctgtc	acccaggctg	240
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catgcctcag	cctcctgagc	agctgggact	ataggcgat	gctaccatgc	caggctaatt	360
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<211> 463						
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<213> Homo sapien						
<400> 50						
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ccatctgcat	ctgcataaggg	tattggggcg	tttgcattccat	atagccatga	ttgctgtgg	420
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<400> 51						
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tgattacaat	aatggaaactt	agatttatata	attacaattt	tttccttagc	atgttggttc	180
cataattatt	aagagtatgg	acttacttag	aaatgagctt	tcattttaaag	aatttcatct	240
ttgaccttct	ctattagtct	gagcagtatg	acactatacg	tatTTTATT	aactaaccta	300
ccttgagcta	ttactttta	aaaggctata	tacatgaatg	tgtattgtca	actgtaaagc	360
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<210> 52						

<211> 392

<212> DNA

<213> Homo sapien

<400> 52

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ttgaaggata	tttgaataat	tcaaaagcg	aatcagtagt	180
tagctagaac	gttggacc	tggatctaag	atcagccgaa	240
ttttgtgtaa	acctcctaca	tcctgggct	tccactaacc	300
aaataggaag	cgcttcgctc	tggtcgcctc	agctgattgg	360
acaaagaggc	atataatgaa	atttgtcaaa	gtaaaggctg	
ctgtattccc	gtgtctttt	ggtctctttt	ccatccat	392
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<210> 53

<211> 179

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(179)

<223> n = A,T,C or G

<400> 53

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tttcagattc	ctgttaaacct	ctaaagaaaa	ggagtgcgc	ctcaactgtat	120
ctagttcagc	atacngagac	acntctgact	ccgattctag	aggactgagt	179
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<210> 54

<211> 112

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(112)

<223> n = A,T,C or G

<400> 54

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tgcattttcc	cacanacaaa	attcaaata	ntgaaagaaa	ttggganagt	112
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<210> 55

<211> 225

<212> DNA

<213> Homo sapien

<400> 55

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aaaggagtat	atccaaatgc	caataaaacat	ataaaaagga	agggattcac	120
gaagwatgca	aattaaaacc	ataatgagaa	accactatgt	atccactagaa	180
cttaaaagac	tggtaaaacc	aagtgttgt	tagataaaaat	gagca	225

<210> 56

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<211> 175		
<212> DNA		
<213> Homo sapien		
<400> 56		
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tttagtattgg gatTTACCC ctgtcctata aagatgttat gtaccaaaaa tgaagtggag	120	
ggccatACCC tgagggaggg gagggatctc tagtgttgTC agaAGCGAA gCTCA	175	
<210> 57		
<211> 223		
<212> DNA		
<213> Homo sapien		
<400> 57		
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tcccagttgc tcctggTCAC tccCTTATAA gCcattactg tCTTGTtCT tgtaactcAG	180	
gttaggtttt ggtctcttCT gCTCCACTGC aaaaaaaaaaaa AAA	223	
<210> 58		
<211> 211		
<212> DNA		
<213> Homo sapien		
<400> 58		
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aactgacttg gatcaatcaa atgtgactGA ggaaacacctt gaagggtgaag aacatcatcc	120	
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<210> 59		
<211> 208		
<212> DNA		
<213> Homo sapien		
<400> 59		
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<210> 60		
<211> 171		
<212> DNA		
<213> Homo sapien		
<400> 60		
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aaccactgac accAGTTGGC aataGCTTCT tccttCTTA acCTTCTAGA gtatttATGG	120	
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<210> 61		
<211> 134		

<212> DNA		
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<220>		
<221> misc_feature		
<222> (1)...(134)		
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canaatcatc nggc	134	
<210> 62		
<211> 145		
<212> DNA		
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gccatttaat caataagtga tagagtcaag gctcaaccca ggtgtgacgg attccaggc	120	
ccaagctcct tactggtacc ctctt	145	
<210> 63		
<211> 297		
<212> DNA		
<213> Homo sapien		
<400> 63		
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ccacagtcag cgccatggtg gtccggtaaa gcatttggtc aggccaggct cgttcaggt	180	
agacgggcac acatcagctt tctggaaaaa cttttgtage tctggagctt tgaaaaa	240	
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<210> 64		
<211> 300		
<212> DNA		
<213> Homo sapien		
<400> 64		
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aatgttttac cattttctgt cttgcctgtt tttctgtgtt tttgttggtc tcttcattct	180	
ccattttttag gcctttacat gtttagaaata tattttttt aatgataactt caccttgg	240	
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<210> 65		
<211> 203		
<212> DNA		
<213> Homo sapien		
<400> 65		
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<210> 66	
<211> 344	
<212> DNA	
<213> Homo sapien	
<400> 66	
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<210> 67	
<211> 157	
<212> DNA	
<213> Homo sapien	
<400> 67	
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<210> 68	
<211> 137	
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<213> Homo sapien	
<400> 68	
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<210> 69	
<211> 137	
<212> DNA	
<213> Homo sapien	
<400> 69	
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<210> 70	
<211> 220	
<212> DNA	
<213> Homo sapien	
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<222> (1)...(220)

<223> n = A,T,C or G

<400> 70

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cgagggcant	ctcatwgaca	gttccaccc	accaaactgc	aaggagctca	nnaagtactr	180
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<210> 71

<211> 353

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

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<223> n = A,T,C or G

<400> 71

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tcccanctaa	atatgccaag	tgacttcaca	tgtttatctt	aaagatgtcc	aaaacgcaac	120
tgattttctc	ccctaaacct	gtgatggtgg	gatgattaan	cctgagtgt	ctacagcaag	180
ttaagtgcaa	ggtgctaaat	gaangtgacc	tgagatacag	catctacaag	gcagtagcctc	240
tcaacncagg	gcaactttgc	ttctcanagg	gcatttagca	gtgtctgaag	taatttctgt	300
attacaactc	acggggcggg	gggtgaatat	ctantggana	gnagacccta	acg	353

<210> 72

<211> 343

<212> DNA

<213> Homo sapien

<400> 72

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aaaatgttyg	caatctctcc	atctgacaaa	aggctaata	ccagawtcta	awaggaactt	120
aaacaaattt	atgaaaaaag	aacaracaac	ctcawcaaaa	agtgggtgaa	ggawatgcts	180
aaargaagac	atyatttcag	ccagtaaaca	yataaaaaa	aggctcatsa	tcactgawca	240
ttagagaaat	gcaaataaaa	accacaatga	gataccatct	yayrczagtt	agaayggtg	300
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<210> 73

<211> 321

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

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<223> n = A,T,C or G

<400> 73

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agaaggttag	aaagtctttg	gttctgaagc	agcttctaag	atctttcat	ttgcttcatt	120
tcaaagttcc	catgtgccca	aagtgccatc	cttgggtta	ctgtttctg	agctccagtg	180

ataactcatt tataacaaggg agatacccg aaaaaaaagtg agcaaatctt aaaaaggtgg	240
cttgagttca gccttaaata ccatcttcaa atgacacaga gaaagaanga tgttgggtgg	300
gagtggatag agaccctaac g	321
<210> 74	
<211> 321	
<212> DNA	
<213> Homo sapien	
<400> 74	
gcactgagag gaacttcaga gagagagaga gagttccacc ctgtacttgg ggagagaaac	60
agaaggttag aaagtctttg gttctgaagc agcttctaag atctttcat ttgcttcatt	120
tcaaagttcc catgtgccca aagtgccatc ctttgggtta ctgtttctg agctccagtg	180
ataactcatt tataacaaggg agatacccg aaaaaaaagtg agcaaatctt aaaaaggtgg	240
cttgagttca gccttaaata ccatcttcaa atgamacaga gaaagaaggaa tgttgggtgg	300
gagtggatag agaccctaac g	321
<210> 75	
<211> 317	
<212> DNA	
<213> Homo sapien	
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gagtggatag agaccct	317
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<211> 244	
<212> DNA	
<213> Homo sapien	
<400> 76	
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ttgcctatggt ggtttgtgc acccatcaat ccatcatctta cattaggtat ttctcttaat	180
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gtgc	244
<210> 77	
<211> 254	
<212> DNA	
<213> Homo sapien	
<400> 77	
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gatggcaagt tcwtttacca cactctttaa catttgcattt agtttaacc ttatattatg	120
gataataaaag gttaatatta ataatgattt attttaaggc attccraat ttgcataatt	180
ctccttttgg agataccctt ttatctccag tgcaagtctg gatcaaagtg atasamagaa	240
gttcctctca gtgc	254

<210> 78		
<211> 355		
<212> DNA		
<213> Homo sapien		
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<221> misc_feature		
<222> (1)...(355)		
<223> n = A,T,C or G		
<400> 78		
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cctgaggggg cgcaggaccct ttagtaccct cagaatcttc acaacgggag atggactgg	180	
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ctcaacaggt tccgatggct gtgatggca tagtcatant taacntgtn tcgaa	355	
<210> 79		
<211> 406		
<212> DNA		
<213> Homo sapien		
<400> 79		
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ccccagtgc acgttaggtg tgcattctcc agccatcaag agactgagtc aagttgttcc	180	
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cgaaatctg tcgatttagac tggacagctt gtggcaagtg aatttgcctg taacaagcca	300	
gattttttaa aatttatattt gtaaataatg ttttgtgtgt ttttgtgtata tatatatata	360	
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<211> 327		
<212> DNA		
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<400> 80		
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tgttagggctc atggtaggggg taaaaggagg gcaatttcta gatcaaataa taagaaggta	180	
atagtacta agaagaattt tatggagaaaa gggacgcggg cgggggatata agggtcgaag	240	
cgcactcgtt aagggttggaa tttttctatg tagcgttga gtttgtttag tcaaaaatgtt	300	
ataatttattttt ttagtaagcc taggaga	327	
<210> 81		
<211> 318		
<212> DNA		
<213> Homo sapien		
<400> 81		
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attgcattca taatttattttt tgcattttatg cttgtatctc ctaagtcatg gtatataatc	120	
catgctttttt atgtttttgtc tgacataaaac tcttattcaga gccctttgca cacaggatt	180	

caataaat taacacagtc tacatttatt tggtaatat tgcataatctg ctgtactgaa
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atgattgcgc atagacta

<210> 82
<211> 338
<212> DNA
<213> Homo sapien

<400> 82

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ctgatcaaattt atcactctcc tacttacagg actcaacata ctatcacag ccctataactc
cctctacata tttaccacaa cacaatgggg ctcactcacc caccacatta acaacataaaa
accctcattt acacgagaaa acaccctcat gttcatacac ctatccccca ttctcctcct
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<210> 83
<211> 111
<212> DNA
<213> Homo sapien

<400> 83

agccatttac caccatcca caaaaaaaaaaaaaaaaag aaaaatatca aggaataaaaa
atagactttt aacaaaaagg aacatttgct ggccctgagga ggcacacccc g

<210> 84
<211> 224
<212> DNA
<213> Homo sapien

<400> 84

tcgggtgatg cctcctcagg ccaagaagat aaagtttcag acccctaaca catttccaaa
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tgaggtggat tcacgagttt cggacaactc ctttgcatttgc aagcgagggtg cagccggaga
ctggggagag cgagccaaatc aggtttgaa gttcctctca gtgc

<210> 85
<211> 348
<212> DNA
<213> Homo sapien

<400> 85

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ctcagtaact tccttgtgtt gtgtgtattt aactcacasa gttgaacat cctttacaca
gagcagactt gtaacactct twttgtggaa tttgcaagtg gagatttcag scgctttgaa
gtsaaaggta gaaaaggaaa tatcttccta taaaaacttag acagaatgt tctcagaaac
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<210> 86
<211> 293
<212> DNA
<213> Homo sapien

<400> 86	
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tgggtggat tcacgagttc cgacacaactc cttgatgcc aagcgaggtg cagccggaga
ctggggagag cgagccaatc aggtttgaa gttcctctca gtgc 180

<210> 145 224
<211> 111
<212> DNA
<213> Homo sapien

<400> 145 111
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atagactttt aacaaaaagg aacatttgct ggcctgagga ggcacacccc g 60

<210> 146
<211> 585
<212> DNA
<213> Homo sapien

<400> 146 120
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cagaggatgc tttgcagaaa cttcataaat atatgcagggt gatttcctt ttcctcctag
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acattactt agtaaggcat tatggaaagt ttcttttag gtatagttt tcctaattgg
gtttgacatt gcttcatagt gcctctgtt ttgtccataa tcgaaaagtaa agatagctgt
gagaaaacta ttacctaaat ttggtagtt gttttgagaa atgtcctt agggagctca
cctgggtggtt tttaaattt tggtagttc ataattggc taattataaa aaccttttgc
agacatattt taaaattgtct ttccctgtaa tactgatgtat gatgtttct catgcatttt
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<210> 147
<211> 579
<212> DNA
<213> Homo sapien

<220>
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<222> (1)...(579)
<223> n = A,T,C or G 240
<224> 300
<225> 360
<226> 402

<400> 147	
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ggagtgaatg ttccacgact ttgcaggag tgtcagaag ccaggtgcaa cttggttgc	180
ttgtgttcat caccctcaa gatatgcaca ctgcttcca aataaagcat caactgtcat	240
ctccagatgg ggaagacttt ttctccaacc agcaggcagg tccccatcca ctcagacacc	300
agcacgtcca ccttctcggg cagcaccacg tcctccacct tctgctggta cacgtgtatg	360
atgtcagcaa agccgttctg cangaccacg tgccccgtgt gctgtgccat ctcactggcc	420
tccaccgcgt acaccgcgtc aggccgcgca tantgtcac agaanaaaatg atgatccagt	480
cccacagccc acgtccaaga ngactttatc cgtcaggat tctttattct gcaggatgac	540
ctgtggtatt aattgttcgt gtctggcgc aacatgcta	579
<210> 148	
<211> 249	
<212> DNA	
<213> Homo sapien	
<400> 148	
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ttggcaccag gatctggac ttccaatctc cagaactgtg agaaataagt atttgcgtct	120
aaataaaatct ttgtggtttc agatatttag ctatagcaga tcaggctgac taagagaaac	180
cccataagag ttacatactc attaatctcc gtctctatcc ccaggtctca gatgctggac	240
aagggtgtca	249
<210> 149	
<211> 255	
<212> DNA	
<213> Homo sapien	
<400> 149	
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gtgagatgtt aactcattgt gggTTGGC tgcatttctc taatgatcat tggatTTAAG	120
CTTTTTAAATGCTTGTTGACCTTGTGACCTGATGATCATCTTTGAGAAGT GTCTGTTCT	180
ATCCCTTGCCTACTTTAAATGCTTGTTATC TTGTAATTCTTACAGATGCTTGACAGATG	240
CTGGACAAGGTGTCA	255
<210> 150	
<211> 318	
<212> DNA	
<213> Homo sapien	
<400> 150	
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ggaaagtca agtccacgac agggTGGGTG ggttagacagt ggcactcaga aatgtcagct	120
ggacccctgt ccccgcatag gcaggacacg aaggctgtgg ctctccaggg ccagctgaa	180
aacaggacac tgcacccgtt gcccacaAGC gtcagagact cccatTTG aagcacggcc	240
ttcttggctt tcctgcactt ccctgttctg tttagagacct ggttatagac aaggcttctc	300
cacagtgtt cagcgtaa	318
<210> 151	
<211> 323	
<212> DNA	
<213> Homo sapien	

<220>
 <221> misc_feature
 <222> (1)...(323)
 <223> n = A,T,C or G

<400> 151

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tcnngccta aaagcnnntc cactacatgc ntcancactg tntgtgtac ntcatnaact	180
gtcngnaata gggcncata actacagaaa tgcanttcat actgcttcca ntgcacatcng	240
cgtgtggcct tncctactct tcttntattc caagtagcat ctctggantg cttccccact	300
ctccacattt ttgcagcat aat	323

<210> 152
 <211> 311
 <212> DNA
 <213> Homo sapien

<400> 152

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ggagagagct gtagtttga gggttgcaaa gacttaggat ggagttggtg ggtgtggta	120
gtctctaagg ttgattttgt tcataaattt catcccctga atgccttgct tgcctcaccc	180
ttgtccaaagc cttagtgaac acctaaaagt ctctgtcttc ttgctctcca aacttctcct	240
gaggatttcc tcaagattgtc tacattcaga tcgaagccag ttggcaaaca agatgcagtc	300
cagagggtca g	311

<210> 153
 <211> 332
 <212> DNA
 <213> Homo sapien

<400> 153

caagattcca taggctgacc aggaggctat tcaagatctc tggcagttga ggaagtctct	60
ttaagaaaat agttaaaca atttgtaaa attttctgt cttaatccat ttctgttagca	120
gtttagatct ggctgtcctt ttataatgc agagttggaa ctttccctac catgtttat	180
aatgttgtc caggctccat tgccaataat gtgtgtcctt aaatgcctgt ttatgtttt	240
aagacggAAC tccaccctt gcttggctt aagtatgtat ggaatgttat gataggacat	300
agttagtagcg gtggcagcc tatggatatct tg	332

<210> 154
 <211> 345
 <212> DNA
 <213> Homo sapien

<220>
 <221> misc_feature
 <222> (1)...(345)
 <223> n = A,T,C or G

<400> 154

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acattgcata tcctcagaga gggaggagat gtangtctgg gcttccacag ggacctggta	180

ttttaggatc agggtaccgc tggcctgagg cttggatcat tcanagcctg ggggtggaat	240
gctggcagc ctgtggcccc attgaaatacg ctctggggc actccctctg ttcctanttg	300
aacttgggta aggaacagga atgtggtcan cctatggaat cttga	345
<210> 155	
<211> 295	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(295)	
<223> n = A,T,C or G	
<400> 155	
gacgcttggc cacttgacac attaaacagt tttgcataat cactancatg tatttctagt	60
ttgctgtctg ctgtgatgcc ctgcccgtat tctctggcgt taatgatggc aagcataatc	120
aaacgcgttt ctgttaattc caagttataa ctggcattga ttaaaggcatt atctttcaca	180
actaaaactgt tcttcatana acagcccata ttatttatcaa attaagagac aatgtattcc	240
aatatccctt angccaata tatttnatgt cccttaatta agagctactg tccgt	295
<210> 156	
<211> 406	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(406)	
<223> n = A,T,C or G	
<400> 156	
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cctcgaagcc caggcagagg accagccatc ccagcctgca ggtaaagtgt gtcacctgtc	120
aggtgtggctt ggggtgagtg ggtggggaa gtgtgtgtgc aaagggggtg tnaatgtnta	180
tgcgtgtgag catgagtgtat ggctagtgtg actgcatgtc agggagtgtg aacaagcgtg	240
cgggggtgtg tgtcaagtgtcgtatg tgagaatatg tgtctgtgga tgagtgcatt	300
tgaaagtctg tgtgtgtgcg tgtggcatg angtaantt antgactgcg caggatgtgt	360
gagtgtgcattt ggaacactca ntgtgtgtgt caagtggccn ancgat	406
<210> 157	
<211> 208	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(208)	
<223> n = A,T,C or G	
<400> 157	
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ggcatgtgatgatgatcacttggcac tcatttgcattt ggcagtact gtaanccana	120
tctgtatgcattt acaccagctt gtaaattgaa taaatgttca taataactatg tgctcacaat	180

anggtanggg tgaggagaag gggagaga	208
<210> 158	
<211> 547	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(547)	
<223> n = A,T,C or G	
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agggttcat catgttgccc tggctggct caaactcctg acctaaggca atgtgcccac	180
ctcagcctcc caaagtgctg ggattacagg cataagccac catgcccagt ccatntttaa	240
tcttcctac cacatttta ccacacttcc ttttatgttt agatacataa atgcttacca	300
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gaacagtaggg caataccaca tagcttaggt gtgtggtaga ctataccatc taggtttgtg	420
taagttcacac ttatgtctgt ttacacaatg acaaaaccat ctaatgatgc atttctcaga	480
atgtatcctt gtcagtaagc tatgtatgtac agggAACACT gcccaaggac acagatattg	540
tacctgt	547
<210> 159	
<211> 203	
<212> DNA	
<213> Homo sapien	
<400> 159	
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aacagcctgt atccaaacac ttaacacact cacctgaaaa gttcaggcaa caatcgccctt	120
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tcgatagaag ttctctctcag tgc	203
<210> 160	
<211> 402	
<212> DNA	
<213> Homo sapien	
<400> 160	
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taaacaataa taataatatt tagcatttat agaggacttt atatcttcaa agtacttgca	120
aacattayct aattaaatac cctctctgtat tataatctgg atacaatgc acttaaaactc	180
aggacaggggt catgagaraa gtagcattt gaaagttggt gctagctatg ctttaaaaac	240
ctataacaatg atgggraagt tagagttcag attctgttgg actgttttg tgcatttcag	300
ttcagcctga tggcagaatt agatcatatc tgcactcgat gactytgctt gataacttat	360
cactgaaatc tgagtgttga tcatcacact gctcgactta ca	402
<210> 161	
<211> 193	
<212> DNA	
<213> Homo sapien	

<400> 161		
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ttagcggaca aggacatgaa aacagctatt gtaagagcgg atatagttgt gtgtgtctgg	180	
gctcaacatg cta	193	
<210> 162		
<211> 147		
<212> DNA		
<213> Homo sapien		
<400> 162		
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tggtgtgtgt ctggctcaa catgcta	147	
<210> 163		
<211> 294		
<212> DNA		
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<400> 163		
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ttgggttctta tctccctcac attatcttca tttctatcat tgacctctta tcccagagac	180	
tctcaaaactt ttatgttata caaatcacat tctgtctcaa aaaatatctc acccacttct	240	
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<210> 164		
<211> 412		
<212> DNA		
<213> Homo sapien		
<220>		
<221> misc_feature		
<222> (1)...(412)		
<223> n = A,T,C or G		
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cacctggctg caagtgcgcc agagccgccccc tgactacgtg ctgctgtggg gctggggcgt	120	
gatgaactcc accgcccctga aggaagccca ggcacccga taccccccgcg acaagatgt	180	
ccggcgtgtgg tggggccgtg cggagcccgta tgcgtgtac gtggggcgaag ggcaccaagg	240	
ctacaacgcg ctggctctga acggctacgg cacgcagtcc aaggtgatcc angacatcct	300	
gaaacacgtg cacgacaagg gccaggccac gggccccaaa gacgaagtgg gctcggtgct	360	
gtacacccgc ggcgtgatca tccagatgtt ggacaagggtg tcaatcacta at	412	
<210> 165		
<211> 361		
<212> DNA		
<213> Homo sapien		
<400> 165		
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gaaggcaaag gagaacaggc attgtatggc aagaaaggaa gaaagagaga ggggagaaaag gtgctaggtt ctttcaaca accagttctt gatggactg agagtaagag ctcaaggcca ggtgtgtgtgta ctccaaaccag taatcccaac attttaggag gctgaggcag gcagatgtct tgaccccattg agtttgtgac cagcctgaac aacatcatga gactccatct ctacaataat tacaaaaattt aatcaggcat tgtgttatgc cctgttagtcc cagatgctgg acaaggtgtc a	120 180 240 300 360 361
<210> 166 <211> 427 <212> DNA <213> Homo sapien	
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aaaaaaaagaa tgatcagagc cacaataca gaaaacctg agtcaccgag cgatgaaa	358
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<211> 1265	
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<213> Homo sapien	
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aacatgtatt ttatggacca aattgacatt ttgcactatt tttcccaa aaaagtcaagg	120
tgaatttcag cacactgagt tgggatttc ttatcccaga agwcggcacg agcaatttca	180
tatttattta agattgattc catactccgt tttcaaggag aatccctgca gtctccctaa	240
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actctaaaaa aacagagaac aaatatgtct cagttgtatt aagcacggac ccatattatc	360
atattcattt aaaaaaatga tttcctgtgc acctttggc aacttctt ttcaatgttag	420
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atgggttcat attttctt aaaaagaaaaa tataattatg aaagccaaga taatctgaag	780
cctgttttat ttaaaaactt tttatgttct gtgggttcat ttgtttgtt gtttgggttct	840
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aagacaatg tgaaaaggat aatatagttg gatcaaacaa aaacaacaca atttgtcccg	300
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aacatcacac tgctcgactt aca	383
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aagaacataaa tgaagtaaca tttaattac tcaaggacta cttttggtt aagtttataaa	180

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<212> DNA	
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cattagcagt ggaagaagaa atgttgatattttatgtcatttata atcaccagag	180
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<213> Homo sapien

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<210> 199

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<212> DNA

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<220>

<221> misc_feature

<222> (1)...(1027)

<223> n = A,T,C or G

<400> 199

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tttsgaccag tgaaccatt gwacaggacc tcatttccty tgagatgrta gccataatca	300
gataaaaagrt tagaagtytt tctgcacgtt aacagcatca ttaaatggag tggcatcacc	360
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gcccccgct gcaggtcaac atatngggaaa accccccacc ccttnngagc ntaccttga	900
ttttccatat gtcccnntaaa ttanctngnc ttanctngc cntaacctnt tccggttaa	960
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<210> 200

<211> 207

<212> DNA

<213> Homo sapien

<400> 200

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tatactacca gcgtcgtaat gtcacta	207

<210> 201

<211> 209

<212> DNA

<213> Homo sapien

<400> 201

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gaggttacat ctggagtctt cgatataatca ggaaaaatgg aagtgaacat tcacagagttt	120
ttaacttctttt gggaaactcaa atgctagaaa agaaaagggt gcccttttc tctggcttcc	180
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<210> 202

<211> 349

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(349)

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<400> 202

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tgcgggaccc cgacgagcgt cactgggtac agaccagatt cagccggaaag agaaaggcgcc	180
gcagggagag actcgaactc cactccgctg gtgagcagcc ccatgttttc aactcgaagt	240
tcaaacggca ttgggttata taccatcagc tgaacttcaac acacatctcc ttgaacccac	300
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<210> 203

<211> 241

<212> DNA

<213> Homo sapien

<400> 203

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acaactgcta ccaccaccac caaccttaggg atttaggatt ctccacagac cagaaattat	180
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<210> 204

<211> 248

<212> DNA

<213> Homo sapien

<400> 204		
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cagaactaaa tactcaatgc tatgtgttca tgtctgtgt tatgtgtgt taatgttca	180	
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agaggagc	248	
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<211> 505		
<212> DNA		
<213> Homo sapien		
<220>		
<221> misc_feature		
<222> (1)...(505)		
<223> n = A,T,C or G		
<400> 205		
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ctctataact ggtgatgcta gaggtgatgt ttttgtaaa caggcggtgt aagatttgcc	180	
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ttttaatctg acgcaggcatt atgcggagga gaatgtttt atgttactta tactaacatt	360	
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<212> DNA		
<213> Homo sapien		
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<211> 176		
<212> DNA		
<213> Homo sapien		
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<211> 196		
<212> DNA		
<213> Homo sapien		
<400> 208		

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<210> 209	
<211> 345	
<212> DNA	
<213> Homo sapien	
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<221> misc_feature	
<222> (1)...(345)	
<223> n = A,T,C or G	
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<213> Homo sapien	
<400> 212	

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tgatcacctg ggTTTCTTA tttatcgact gtgtcatgac aaggAAACTT acaaACTGCA	180
acgcagagaa actattaaAG gtattcagaa acgtGAAGCC agcaattgtt tcgcaATTCG	240
gcattttgaa aacaAAATTG ccgtggAAAC tttaATTGT tcTTGAACAG tcaagaaaaAA	300
cattattttag gaaaattaAT atcacAGCAT aacggaa	337
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<211> 715	
<212> DNA	
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agtatCCTC ttgtcacCTT gcagACTTAA aacataAAAA tactCATGG tttAAAGG	180
aaaaAAAGTAT acattAGCAC tattaAGCTT ggcCTTGAA catTTCTAT CTTTATTAA	240
atgtcggTTA gctGAACAGA attcATTtTA caatGCAGAG tgAGAAAAGA agggAGCTAT	300
atgcATTtGA gaATGCAAGC attGTCAAAT aaACATTtTA aATGCTTCT taaAGTgAGC	360
acatacAGAA atacATTAAG atATTAGAA gtgtTTTGC ttgtGTACTA ctaATTAGGG	420
aAGCACCTG tataGTTCTT CTTCTAAAT tGAAGTAGAT tttaAAACC catGTAATT	480
aATTGAGCTC tcAGTTcaga ttttaggAGA ATTtTAACAG ggatttGGTT ttGTCTAAAT	540
tttGTCAATT tNTTtagtta ATCTGTATAA TTTATAAAAT gtCAAACtGT ATTAGTCCG	600
ttttCATGCT gctATGAAAG AAATACCCAN gACAGGGTTA ttTATAAANG gAAAGANGTT	660
aATTGACTC ccAGTTcaca ggcCTGAGGA ngnATCnCCC gAAATCCTTA ttGCG	715
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<211> 345	
<212> DNA	
<213> Homo sapien	
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<222> (1)...(345)	
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tcccacCTGc ctgattCTTC atATGTTGGG tggccCTGTT ttTCTGGTGC tatttCCTGA	180
ctgtGTtca gctGCCACTG tcctGCAAAG CCTGCCTTT taaATGCCTC accattCCTT	240
catttGTTc ttaaatatGG gaAGTGAAG tgccACCTGA ggCCGGGAC agtggcTAC	300
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<210> 215	
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<212> DNA	
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tgtAACATCC attcccaagc aagcacaact tcacataata ctttccagaa gttcatttgct	480
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345

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<211> 347
<212> DNA
<213> Homo sapien

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180
240
300
347

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180
240
281

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<211> 3646
<212> DNA
<213> Homo sapien

<400> 227 60
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720
780

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<210> 228
<211> 419
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(419)
<223> n = A,T,C or G

<400> 228

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tggtgacggt	cccagatggc	ttacagaaga	aagtgtcctg	agatgagttt	ttaagaatga	180
ataaggatag	acacaagtga	ggactgactt	ggcagtggtg	aatggtggtt	ggcaaaaaac	240
ttcgcatgt	tggaaactgc	acgtacagga	atgaagaatg	agactgtgt	gtgttaatg	300
agctgcaat	actaatttt	tcctgaaaat	tttgaagagt	taactaaaaa	gtatTTTta	360
gtaaggaaat	aaccctacat	ttcagggtt	ttgtttgtt	anatattgaa	ggtgcccaa	419

<210> 229
<211> 148
<212> DNA
<213> Homo sapien

<400> 229

aagagggtac	ctgtatgttag	ccatggtggc	aatgagagac	tgattactac	ctgctggaga	60
ttgtttaagt	gagttaatat	attaaggata	aaggagcca	ggtttttga	ctgttgaga	120
aggaaattac	agatattgaa	ggtcccaa				148

<210> 230
<211> 257
<212> DNA
<213> Homo sapien

<400> 230

taagagggta	cmaaaaaaaa	aaaatagaac	gaatgagtaa	gacctactat	ttgatagtagc	60
acagggtga	ctatagtcaa	tgataactt	attatacatt	taacatagag	tgtaattgga	120
ttgtttgtaa	ctcgaaggat	aatgcttga	gaggatggat	accccattct	ccatgtatgt	180
cttatttcac	attacatgcc	tgtatcaaag	catctcatat	accctataaaa	tatgtacacc	240
tactatgtac	cctctta					257

<210> 231
<211> 260
<212> DNA
<213> Homo sapien

<400> 231

taagagggta	cgggttattt	ctgatggat	tttttttct	ttctttttct	ttggaaaaca	60
aaatgaaagc	cagaacaaaa	ttattgaaca	aaagacaggg	actaaatctg	gagaaaatgaa	120
gtccccctcac	ctgactgcca	tttcattctt	tctgaccc	cagtcttagt	taggagaata	180
gggggtggag	gggattaatc	tgatacaggt	atatttaag	caactctgca	tgtgtgccag	240
aagtccatgg	taccctctta					260

<210> 232
<211> 596
<212> DNA
<213> Homo sapien

<220>

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<221> misc_feature
<222> (1)...(596)
<223> n = A,T,C or G

<400> 232
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gtgggattaa cattatttaaaa aaaaatcgaa gtattgacaa ggatgtgaag aaatttagaac
atctgtgcac ttttgtggg aatgtaaaaa aggtgtggcc actatggta acagcatgaa
ggttcctcaaa aaaaaattttt ttttaatcta ctctatgatc gatcttgagg ttgttatgc
aaaagaactg aaatcaggat tttgaggaaa tattcacatt cccacatcca tttctgcttt
attcataata ctcagagat gaaacaacc taaatgtcca tcccggatg aatggataaa
cacagtgtgg tatatgcata caatgaaata ttathtagtc tttaaaaaga aaaattctat
cataactac aacttanatn aaccttgagg acacaatgct nagtgaata agccacggaa
gacgaaatac tgcatatttc ctttatatga agtatactaaa gtggtaaaac tcttanagca
naaagtaaaa atgggtgggtt gccanacagt tggtaggcn agaaganaan cctant 60
120
180
240
300
360
420
480
540
596

<210> 233
<211> 96
<212> DNA
<213> Homo sapien

<400> 233
tcttctgaag acctttcgac actcttaagc tcgtggttgg taaggcaaga ggagcggttgg 60
taaggcaaga ggagcggttgg taaggcaaga ggagca 96

<210> 234
<211> 313
<212> DNA
<213> Homo sapien

<400> 234
tgtaagtcga gcagtgtgat gataaaactt gaatggatca atagttgctt cttatggatg 60
agcaaaagaaa gtatgtttctt gtatgtggaaat ctgtcttgg caaaaatgct gtgaacgttg 120
ttgaaaagac aacaaagagt ttagagtagt acataaattt agaatagtagc ataaacttag 180
aatagtacat aaacttagta cataaataat gcacgaagca ggggcagggc ttgagagaat 240
tgacttcaat ttggaaagag tatctactgt aggttagatg ctctcaaaaca gcatcacact 300
gctcgactta caa 313

<210> 235
<211> 550
<212> DNA
<213> Homo sapien

<400> 235
aacgaggaca gatccttaaaa aagaatgttg agtggaaaaaaa gtagaaaaata agataatctc 60
caaagtccag tagcattatt taaacatttt taaaaaaatc actgataaaaa attttgatca 120
tttccccaaa atacatatgg aagcacagca gcatgaatgc ctatggrrt gaggataggg 180
gttgggagta gggatggggta taaagggggaa aataaaaacc agagaggagt cttacacatt 240
tcatgaacca aggagtataa ttatttcaac tatttgatcc wgaagtccag aaagagtgg 300
ggcagaaggg ggagaagagg gcgaagaaac gtttttgggaa gaggggtccc asaagagaga 360
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ttctcatcac taatatttaga ttaaaccctt tgaagacagc gtctgtgggtt tctctacttc 480
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qaqattattc 550

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<210> 236	
<211> 325	
<212> DNA	
<213> Homo sapien	
<400> 236	
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aggaaactcac tattgaatac ataaatggaa ttatttcagc cttaaaaagt ttggaggaa	120
attctgacat atgctaaaac atggatgaa cttgaagact ttatgataag taaaagaaggc	180
cagtcataaa aggaaaaata ttgcattgatt ccacttat gaggtaccta gagtagtcaa	240
tttcataaaaa acacaaaaata gaatgggttt tgccaggct tttgaggaaa aggaaatgac	300
aagtttagggg acatgagtca gtcta	325
<210> 237	
<211> 373	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(373)	
<223> n = A,T,C or G	
<400> 237	
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agacttatct tgcctccaaag caaactctttt atttcttttc atccttagtct ttatttcttg	120
tgcgtctta cccatctcaa aagagtgc aaatccacca agttgtgaa acagaaatct	180
aagaaatatac cttgattctt cttttccca tctacttcac ttcttaattca ttagtaaata	240
atctgtttca gaaaacccaa cacctcatgt tctcactcat aagggggagt tgaacaatga	300
gaacacacac acacagggag gggAACATCA cacaccacgg cccgtcagg agtangggac	360
atgagtcagt cta	373
<210> 238	
<211> 492	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(492)	
<223> n = A,T,C or G	
<400> 238	
tagactgact catgtccctt ataatgctcc caggcatcg aaagcatctc aaactggagg	60
tgacaccatg gcagagggtt caggtaaatc acaaaaagggg tcctaaagaa ttggccctca	120
atatacgtt gattagaaga agtggacaga gctacccaag taaaacatata gcgagataaa	180
aaaaatattgg cacttgtgaa cacacactac aggaggaaaa taaggaacat aatagcatat	240
tgtgttatta tgatgtgaa gaacctctt anaagaaaaac ataacccaaag aaacaaagaa	300
aattcctgcn aatgttaat gctatagaag aaattaacaa aaacatataat tcaatgaatt	360
cagaaaagggtt agcaggtcan aagaaaacaa atcaaagacc agaataatcc catttttagat	420
tgtcgagtaa actanaacag aaagaatacc actggaaatt gaattcctac gtangggaca	480
tgantcanc ta	492

<210> 239	
<211> 482	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(482)	
<223> n = A,T,C or G	
<400> 239	
tggaaagtat ttaatgatgg gcaacttgct gtttacttcc tacatatccc atcatcttct	60
gtatTTTTT aaataacttt tttttggatt tttaaagtaa ccttattctg agaggttaaca	120
tggattacat acttctaaggc cattaggaga ctctatgtta aaccaaaagg aaatgttact	180
agatcttcat ttgatcaata ggatgtgata atcatcatct ttctgctcta atggaaaagt	240
actanaaaaca tggaaaccata atcttagatg aacaacgtta gaatttgac taattctacg	300
gaatttcagt aattcggcaa atgtcgggca gtgacacaac atttcatgac ggggacgcac	360
ctaccaacctt ctggcgataa gggccacccct tccctctgta cttacagtcc catttcatac	420
acagtcttgc attaaatatt cacattttt ctctacctaa agaccttcaa gaccagtacg	480
ta	482
<210> 240	
<211> 519	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(519)	
<223> n = A,T,C or G	
<400> 240	
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gctgtcccccc agcccgacac ccgtaaaggg tctgtctga ggtggattag taaaagagga	120
aaggccttgcgat gttgagatag aggaaggcgtt ctgtctctgt cctgccccctg ggaactgaat	180
gtctcggtat aaaacccgat tgtacatttg ttcaattctg agataggaga aaaaccacccc	240
tatggcgccgat ggcgagacat gttggcagca atgctgcctt gttatgcctt actccacaga	300
tgtttggccgat gagggaaaca taaatctggc ctacgtgcac atccaggcat agtacccc	360
tttgaactta attatgacac agattcctt gctcacatgt tttttgtctg accttctcct	420
tattatcacc ctgctctctt accgcattcc ttgtgtcgat ataatgaaaa taatatcaat	480
aaaaacttgcg nggaactcg agaccactac gtcgataca	519
<210> 241	
<211> 771	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(771)	
<223> n = A,T,C or G	
<400> 241	
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actgtcacgg ctcccggtta gaagtcactt atgagacaca ccagtgtggc cttgttgct	120
tgaagctcct cagaggaggg tgggaacaga gtgaccgagg gggcagcctt gggctgacct	180
aggacggtca gtttgtccc tccgc当地 acgagagtgc tgctgcttgt atatgagctg	240
cagtaataat cagcctcgta ctcagcctgg agccc当地 tggtcagggg gcccggttg	300
ccanacttgg agccagagaa gcgattagaa acccctgagg gccgattacg gacctcataa	360
atcatgaatt tggggcttt gcctgggtgc tttgttacc angagacatt attataacca	420
ccaacgtcac tgctggtcc antgcaggaa aaatgggtga tcnaactgtc caagaaaacc	480
actacgtcca taccaatcca ctaattgccc gccgc当地 ggttcaacca tattggggaa	540
naactccccn ccgc当地tttgg gattgnat naaccttga aatttttcc tattanttgt	600
ccccctaaaaa taaaccnttgc gcncnttacc cattgggtcc atancttntt tncccggtt	660
ttaaaaanttgc ttatcccgc cnccc当地tatttcccccaac tttccaaaac ccgaaaccnt	720
ttnaaatttnt tnaaccctg gggggttccc nnaatnnan ttnaanctnc c	771
<210> 242	
<211> 167	
<212> DNA	
<213> Homo sapien	
<400> 242	
tgggcaccc ttccatcggtt acatcacgt gctgatgtgc ctgttgctgg	60
tcctctctag gaacctctgg attttcaat tctttgagga attcatccaa attatctgcc	120
tctc当地tctt tctaagggtct tctggtaacca gcggta	167
<210> 243	
<211> 338	
<212> DNA	
<213> Homo sapien	
<400> 243	
ttgggc当地tcaatatcta ctgatctaaa tagtgtggtt tgaggcctct tgttc当地tggc	60
taaaaatcct tggcaaggt caatctccac tttacaatag agttaaaaat cttacaatgg	120
atattcttga caaagcttagc atagagacag caatttaca caaggtattt tt当地ctgtt	180
taataacagt ggtt当地tctt caccctatgg gtgc当地ccaa gggaggagtg cacagttgca	240
gaaaccaaatt aagatactga agacaacact acttaccatt tccc当地tatag ctaaccacca	300
gttcaactgt acatgtatgt tcttatggc aatcaaga	338
<210> 244	
<211> 346	
<212> DNA	
<213> Homo sapien	
<400> 244	
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tgcaaaaatc atcaatatac ttgaagatcc cc当地gttaagg tacaatgtat ttaatattat	120
cactgataca attgatccaa taccagttt agtctggcat tgaatcaat cactgtttt	180
gttgtataaa aagagaaata ttagcttattt atttaagtac catattgtaa gaaaaaaat	240
gcttatctt acatgtaaa atcatgatct gtacattggt gcagtgataa ttactgtaaa	300
agggaagaag gaatgaagac gagctaagga tattgaaggt gccc当地	346
<210> 245	
<211> 521	
<212> DNA	
<213> Homo sapien	

<220>
<221> misc_feature
<222> (1)...(521)
<223> n = A,T,C or G

<400> 245

accaatccca cacggatact gagggacaag tatatcatcc catttcatcc ctacagcagc	60
aacttcatga ggcaggagtt attagtcccc ttttacagaa gaggaaactg agacttaggg	120
agatcaagta atttgcccaag gtcgcacaat tagtgataga gccagggtt gaagcgacgt	180
ctgtcttaag ccaatgaccc ctgcagatta ttagagcaac tggtctccac aacagtgtaa	240
gcctcttgc anaagctcag gtccacaagg gcagagattt ttgtctgtt tgctcattgc	300
tccttccccca ttgcttagag cagggtctgc cacgaancag gttctcaatg catagttatt	360
aaatgtataat aagagcaaac atatgttaca gagaacttgc tgtatgttgc tcacttacat	420
gaatcacctg tganatgggt atgcttggc cccantgtt cagatnaaga tattgaangt	480
gcccaaatca ctanttgcgg gcgcctgcan gtccancata t	521

<210> 246
<211> 482
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(482)
<223> n = A,T,C or G

<400> 246

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ACCATGAAAT ACTATGCAGC CATAAAAAAG GATGAGTTCA TATCCTTGC AGGGACATGG	120
ATGAAGCTGG AGACCATCAT TCTCAGCAA CTAACAAGGG AACAGAAAAC CAAACACTGC	180
ATGTTCTCAC TCTTAAGTGG GAGCTGAACA ATGAGAACAC ATGGACACAG GGAGGGAAAC	240
ATCACACAGT GGGGCCTGCT GGTGGGTAGG GGTCTAGGGG AGGGATAGCA TTAGGAGAAA	300
TACCTAATGT AGATGACGGG TTGATGGGT CAGCAAACCA CCATGACACG TGTATACCTA	360
TGTAACAAAC CTGCAATGTT CTCACATGTA CCCCCAGAACT TAAAGTGTAA ATAAGGAAAT	420
TAAGGAAAAAA GTTAAGTATG TCATAGATAC ATAAGGAAATT GTANATATTG AAGGTGCCCA	480
AA	482

<210> 247
<211> 474
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(474)
<223> n = A,T,C or G

<400> 247

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aagttagaga ggggcagaga agacaaggc atatgcaggg ggtgattata acaggtggtt	120
gtgctggaa gtgagggtac tcggggatga ggaacagtga aaaagtggca aaaagtggta	180
agatcagtga attgtacttc tccagaattt gatttctggg ggagtcaaat aactatccag	240
tttgggttat catangcaa cagttgaggt ataggaggtt gaagtcncag tggataatt	300
gaggttatga anggttttgtt actgactgtt actgacaang tctgggttat gaccatggga	360

atgaatgact gtanaagcgt anaggatgaa actattccac ganaaagggg tccnaaaaact aaaaannnaa gnnnnngggg aatattattt atgtggatat tgaangtgcc caaa	420 474
<210> 248	
<211> 355	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(355)	
<223> n = A,T,C or G	
<400> 248	
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<210> 249	
<211> 434	
<212> DNA	
<213> Homo sapien	
<400> 249	
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<210> 250	
<211> 430	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(430)	
<223> n = A,T,C or G	
<400> 250	
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aaaaaaaaan	430
<210> 251	
<211> 329	
<212> DNA	
<213> Homo sapien	
<400> 251	
tggtaactcca ccatyatggg gtcaaccgcc atcctcgccc tcctcctggc tgttctccaa	60
ggagtctgtg ccgaggtgca gctgrtgca tctggagcag aggtgaaaaa gtccggggag	120
tctctgaaga tctcctgtaa gggttctgga tacacctta agatctactg gatgcctgg	180
gtgcgccagt tgcccgaa aggctggag tggatgggc tcatcttcc ttagtactct	240
gataccagat acagccgtc cttccaaggc caggtcacca tctcagtcga taagtccatc	300
agcaccgcct atctgcagtg gagtaccaa	329
<210> 252	
<211> 536	
<212> DNA	
<213> Homo sapien	
<400> 252	
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caggctcctc tgctctaacc aggcttctgg gacagtatta gaaaaggatg tctcaacaag	120
tatgttagatc ctgtactggc ctaagaagtt aaactgagaa tagcataaat cagaccaaac	180
ttaatggtcg ttgagacttg tgcctggag cagctggat aggaaaactt ttggcagca	240
agaggaagaa ctgcctggaa gggggcatca tgtaaaaat tacaagggaa acccacacca	300
gcccccttc ccagctctca gcctagagta tttagcatttc tcagctagag actcacaact	360
tccttgctta gaatgtgcca cggggggag tccctgtggg ttagtggact ctcaagagtg	420
agagtggcat cctatcttct gtgtccccac aggacctgg cccgagactt agcagggtgaa	480
gtttctggc caggcttgc ctttgactca ctatgtgacc tctgggtggag taccaa	536
<210> 253	
<211> 507	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(507)	
<223> n = A,T,C or G	
<400> 253	
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tgaggccgca gtgagccgg accacgccc tacactccag cctggggcat agagtggagac	120
cctccaagac agaaaaagaaa agaaaggaa gggaaaggaa agggaaaagg aaaaggaaaa	180
ggaaaaaggaa aagggaaaaga caagacaaaa caagacttga atttggatct cctgacttca	240
attttatgtt ctttctacac cacaatttct ctgttacta agatgataat ttagaaaccc	300
ctcggtccat tctttacagec aagctggaa tttggtcaag taattacaat aatagtaaca	360
aatttgaata ttatatgcca ggtgttttc attcctgctc tcacttaatt ctaccactc	420
tgatataaat acaattgctg ccgggtgtgg tggctcatgc ctgtaatccc ggcactttgg	480
gagaccgagg tggcgaggats gcaacaa	507
<210> 254	
<211> 222	

<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(222)
<223> n = A,T,C or G

<400> 254

ttggatttgt cactgtgagg aagccaaatc ggatccgaga gtcttttct aaaggccagt	60
actggccaca ctttctcctg ccgccttcct caaagctgaa gacacacaga gcaaggcgct	120
tctgtttac tccccaatgg taactccaaa ccatagatgg tttagctnccc tgctcatctt	180
tccacatccc tgctattcag tatagtcgt ggaccaatcc aa	222

<210> 255
<211> 463
<212> DNA
<213> Homo sapien

<400> 255

tgttgcgtc cataaaatgct gaaatggaaa taaaacaacat gatgagggag gattaagttg	60
gggagggagc acattaagggt ggccatgaag tttgttgaa gaagtgactt ttgaacaagg	120
ccttgggttt aagagctgat gagagtgtcc cagacagagg ggccactggt acaatagacg	180
agatgggaga gggcttgaa ggtgtgcgaa ataggaagga gtttgttctg gtatgagtct	240
agtgaacaca gaggcgagag gccctgggg gtgcagctgg agagttatgc agaataacat	300
taggcccgtt gggggactgt agactgtcag caataatcca cagtttgat tttattctaa	360
gagtgtatgg aagccgtgaa aagggggtta agcaaggagt gaaattatca gattacagt	420
gataaaaaata aattggtctg gctactgggg aaaaaaaaaaaa aaa	463

<210> 256
<211> 262
<212> DNA
<213> Homo sapien

<400> 256

ttggatttgt caacctgctc aactctacyt ttccctccttc ttccctaaaaa attaatgaat	60
ccaatacatt aatgccaaaa cccttgggtt ttatcaatat ttctgttaaa aagtattatc	120
cagaactgga cataatacta cataataata cataacaacc ccttcattctg gatgcaaaca	180
tctattaata tagcttaaga tcactttcac tttacagaag caacatcctg ttgatgttat	240
tttgatgttt ggaccaatcc aa	262

<210> 257
<211> 461
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(461)
<223> n = A,T,C or G

<400> 257

gnngnnnnnn nnnaattcg actcngrttcc cttgtancc ggtcgacatg gccgcgggat	60
taccgcttgtt nnctgggggt gtatggggta ctatgaccgc ttgttagctgg gggtgtatgg	120

gggactatga ccgtttagt mtggkgtgt atggggact atgaccgctt gtcgggttgt	180
cggataaaccc gacgcaaggg acgtgatcga agctcggttc ccgcctttc gcatcggtag	240
ggatcatgga cagcaataatc cgcattcgyc tgaaggcggtt cgaccatcgc gtgctcgatc	300
aggcgaccgg cgacatcgcc gacaccgcac gccgtaccgg cgcgctcatc cgcggtccga	360
tcccgttcc cacgcgcata gagaagttca cggtaaccc tgccccgac gtcgacaaga	420
agtcgcgcga cgagttcgag gtgcgtacct acaagcggtc a	461
<210> 258	
<211> 332	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(332)	
<223> n = A,T,C or G	
<400> 258	
tgaccgcttg tagctggggg tgtatgggg actacgaccg ctttagctg ggggtgtatg	60
ggggactatg accgcttcta gctgggggtg tatggggac tatgaccgct ttagctgg	120
gggttatggg ggactaggac cgctttagc tgggggtgt tggggacta tgaccgcttg	180
tagctggggg tgtatgggg actacgaccg ctttagctg ggggtgtatg gggactatg	240
accgcttcta nctgggggtg tatggggac tatgaccgct ttagctgcct gggggatggg	300
aggagagttt tggggggaaa aaaaaaaaaaa aa	332
<210> 259	
<211> 291	
<212> DNA	
<213> Homo sapien	
<220>	
<221> misc_feature	
<222> (1)...(291)	
<223> n = A,T,C or G	
<400> 259	
taccgcttgt gaccgcttgt gaccgcttgt gaccgcttgt gaccgcttgt	60
gaccgcttgt gaccgcttgt gaccgcttgt gaccgcttgt gaccgcttgt	120
gaccgcttgt gaccgcttgt naaaaaaaaaa gtctggggaa ctatgannga ntgtactgg	180
gggtgtctgg gggncatgaa nngantgtna cnnggggtgt ctggggact atganngact	240
gtgcnnctg ggggatcnga ggagantnngn ggntagngat ggttngggan a	291
<210> 260	
<211> 238	
<212> DNA	
<213> Homo sapien	
<400> 260	
taagagggtt ctggttaaaa tacagggaaat ctggggtaat gaggcagaga accaggatac	60
tttgagggtca gggatgaaaa ctagaatttt tttctttttt ttgcctgag aaacttgctg	120
ctctgaagag gcccattgtat taattgcttt gattttcctt ttcttacagc ctttcaagg	180
gcagagccct ctttacctt aaggaatctt atccttagct atagatgtt ccctcttta	238
<210> 261	

<211> 746
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(746)
<223> n = A,T,C or G

<400> 261

ttgggcaccc	tcaatatcaa	tagctaaca	ttatttagtg	tttatcgat	cataaaacac	60
tgttctaagg	ctttaaacgt	actaattcat	ttaatgctca	taatcactt	agaaggtggg	120
tactagtatt	agtctcattt	acagatgcaa	catgcaggca	cagagaggtt	aattaacttg	180
cccaaggtaa	cacagctaag	aaatagaaaaa	aatattgaat	ctggaaagtt	gggcttctgg	240
gtaaccacaa	gagtcttcaa	tgagcctggg	gcctcactca	gttgctttt	acaaaagcgaa	300
ttagtaacat	cacttaattc	agttagttagg	ccaaatggag	gtcagctacg	agtttctgct	360
gttcttgcag	tggactgaca	gatgtttaca	acgtctggcc	atcagtwaat	ggactgatta	420
tcattggaw	gtgggtgggc	tgaatgttgg	ccagtgaaat	ttattcawgc	catattttta	480
tgtttaggat	gactttggc	ttgtccttagg	gcaagctctg	tctgscacgg	aacacagaat	540
wacacaggga	ccccctcaat	ttctggtgtg	gctagaacca	tgaaccactg	gttgggggaa	600
caagcggctca	aaacctaagt	gcggccggct	ggcagggtcc	accatatacg	ggaaaactcc	660
cnacgcgttt	ggaatgcctn	agctngaatt	attctaana	ttgtccncnt	aaaatttagcc	720
ttggcgtaa	tcanggtcn	naagcc				746

<210> 262

<211> 588
<212> DNA
<213> Homo sapien

<220>

<221> misc_feature
<222> (1)...(588)
<223> n = A,T,C or G

<400> 262

tgaccgcttg	tcatctcaca	tgggtcctg	cacgctttt	cctttgtagg	aaacctgaca	60
tttgtctgtt	tcttctttct	ctttccttc	ccatatcctc	ctaatttacg	tttgacttgt	120
ttgctgagga	ggcaggagct	agagactgt	gtgagctcat	aggggtggga	agtttatcct	180
tcaagtcccg	cccactcatc	actgcttctc	accttcccct	gaccaggctt	acaagtgggt	240
tcttgccctgc	tttccctttg	gacccaacaa	gcccctgtaa	tgagtgtgca	tgactctgac	300
agctgtggac	tcagggctct	tggctacagc	tgccatgtaa	aatatctcat	ccagttctcg	360
caaattgtta	aaataaccac	atttcttaga	ttccagtacc	caaatcatgt	ctttacgaac	420
tgctcctcac	acccagaagt	ggcacaataa	ttcttgggaa	attattactt	ttttttttct	480
ctctntnnnc	gnnnnnnnng	gnnnngnccag	gaattaccac	nttggaaagac	ctggccngaa	540
tttattattan	aggggagccg	attnttttc	ctaacacaaa	gcgggtca		588

<210> 263

<211> 730
<212> DNA
<213> Homo sapien

<220>

<221> misc_feature
<222> (1)...(730)

<223> n = A,T,C or G

<400> 263

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agactgc当地	aagattaaat gtaaaagt	tcttgatac agtaatgtt	120
ttanatttat aaatgaaaaa	ttagggcatt tggatatac	agttgaaaat tcaggagt	180
ggttgggctg gctgggtata	tactgaaaac tgcgtac	cagatgacat ctaaaaccac	240
aatctgggtt ttatTTAGC	agtatgtgt gtcactccc	caaaagcctt cccaattggc	300
ctcagcatac acaacaagtc	acctccccac agccctctac	acataaaacaa attccttagt	360
ttagttcagg aggaaatgcg	ccctttcct tccgctctag	gtgaccgcaa ggcccagttc	420
tcgtcaccaa gatgttaagg	aaaggttgcg aaagaggcat	ctgaaaggaa ataaggggaa	480
tgggagtgac cacaaggaa	agccaaggan aaactttgga	gaccgttct agancctgg	540
catttcacaa caaaactcng	gaacaaaacct tgtctcatca	atcatttaag cccttcgtt	600
ggannagact ttctgaactg	ggcgctgaac ataancctca	ttgaatgtct tcacagtctc	660
ccagctgaag gcacacettg	ggccagaagg ggaatcttcc	aggtcctcaa nacagggctc	720
gccctttgnc			730

<210> 264

<211> 715

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(715)

<223> n = A,T,C or G

<400> 264

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tacacttaat gtggatatag	atgcgttgta gcttacttct	accacctgc tatttctccc	120
gtctctttt tgttcccttt	ctttctttt cctcccttat	tttataattt aatttttttag	180
gattctattt tatatagatt	tatcagctat aacactttgt	atttttttgt tttgtggttc	240
ttctgtcatt tcaatgtgca	tcttaaactc atcacaatct	atttcaaat aatatcatat	300
aacttacat ataatgtaa	aatctaccac catatatttc	catttctccc ttccatccta	360
tgtntgtcat atttttcct	ttatataatgt tttaaagaca	taatagtata tgggaggttt	420
ttgcttaaaa tgcgtcaat	attccttcaa ngaaacgtaa	aaattcaaaa taaatntctg	480
tttattctca aatnnaccta	atatttccta ccatntctna	tacnttcaa gaatctgaag	540
gcattggttt ttccggctt	aagaacctcc tctaaagcac	tctaaggcaga attaagtctt	600
ctgggagagg aattctccca	agcttgggcc ttnanntgta	ctccntrang gttaaanttt	660
ggccgggaaa tagaaattcc	aagttAACAG gntantttt	nttttnttn tncc	715

<210> 265

<211> 152

<212> DNA

<213> Homo sapien

<400> 265

tttttttttt tttcccaaca	caaagcacca ttatcttcc	tcacaatttt caacatagtt	60
tgattcccat gaagaggta	tgatttctaa agaaaacatg	gctactatac tatcaatcag	120
ggttaaatct ttttttttg	agacggagtt ta		152

<210> 266

<211> 193

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(193)

<223> n = A,T,C or G

<400> 266

taaactccgt ccccttctta atcaatatgg aggctaccca ctccacattha ctttcttttc	60
aagggactgt ttccgtaact gttgtgggta ttcacgacca ggcttctaaa cctcttaaaa	120
ctccccaaatt ctggtgccaa cttggacaac atgctttttt tttttttttt ttttttttn	180
gagacggagt tta	193

<210> 267

<211> 460

<212> DNA

<213> Homo sapien

<400> 267

tgttgcgate ccttaaggcat gggtgctatt aaaaaaatgg tggagaagaa aataacctgga	60
atttacgtct tatctttaga gattgggaag accctgatgg aggacgtgga gaacagcttc	120
ttcttgaatg tcaattccca agtaacaaca gtgtgtcagg cacttgctaa ggatcctaaa	180
ttgcagcaag gctacaatgc tatgggatc tcccaggag gccaatttct gagggcagtg	240
gctcagagat gcccttcacc tcccatgatc aatctgatct cggttgggg acaacatcaa	300
ggtgttttg gactccctcg atgcccagga gagagctctc acatctgtga cttcatccga	360
aaaacactga atgctggggc gtactccaaa gttgttcagg aacgcctcgt gcaagccgaa	420
tactggcatg acccataaaaa ggaggatgtg gatcgcaaca	460

<210> 268

<211> 533

<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(533)

<223> n = A,T,C or G

<400> 268

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accttcgccc gtggggaccc cgagtagcgtc tacggcgtcg tcacttagag taccctctgg	120
acgcccggc gcgttcgatt taccggaagc gcgagctgca gtgggcttgc gccccggcc	180
aaattcttg gggggtttaa ggccgcggg aatttgaggt atctctatca gtatgtagcc	240
aagttggAAC agtcgccatt cccgaaatcg ctttcttga atccgcaccc cctccagcat	300
tgcctcattc atcaacctga aggacacgt aagtgacggt tgtgtcttca gcagctccac	360
tccataacta gcgcgctcga cctcgtcttc gtacgcgcca ggtccgtgc tgcaattcc	420
caactccggt gagttgcgca tttcaagttt cgaaaactgtt cgccctccacn atttggcatg	480
ttcacgcattt acacggata aactcgcca gtaccggaa tggatcgca aca	533

<210> 269

<211> 50

<212> DNA

<213> Homo sapien

<400> 269		
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<210> 270		
<211> 519		
<212> DNA		
<213> Homo sapien		
<400> 270		
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tgtccaccac ctcctggcac tcttccgaca gggacttcgg cagcttcgag cacattttgt	240	
caaaaaggcgtc gagtatttct ttctcagtc ttttttgtc aatcagcttgc acatccct	300	
tcaccaggaa ttcacacacc tcacagtaaa catcagactt tgctgggacc tcgtgcttct	360	
taatggggtc caccagttcc agggcaggga tgacattttt ggaggccact ttggcgggga	420	
ccagagtcgtc catggcata tctttcacct catcacagaa cccaaaccage gcacagatct	480	
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<211> 457		
<212> DNA		
<213> Homo sapien		
<400> 271		
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gaacagcaca atggcaagac cattttcgcc tactttacgg gttctaagga cgccccgggg	180	
aaaagcttgtt gccccgactg cgtgcagggtt gaaccagtcg tacgagaggg gctgaagcac	240	
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ccaaataatg acttcagaaa aaacttggaa gtaacagcag tgcctacact acttaagtat	360	
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ttctctgaag attaagattt taggatggca atcaaga	457	
<210> 272		
<211> 102		
<212> DNA		
<213> Homo sapien		
<400> 272		
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cgcaggggaa atgcaactgg ccaggtcaca gggcaatcaa ga	102	
<210> 273		
<211> 455		
<212> DNA		
<213> Homo sapien		
<220>		
<221> misc_feature		
<222> (1)...(455)		
<223> n = A,T,C or G		
<400> 273		

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ggcaatcaa aggttaagt cttcgccga agttaatctc gtgttttgga caatcaacag	120
gtttaagtct tcggccgaag ttaatctcggtt gttttggca atcaacaggtt ttaagtcttc	180
ggccgaagtt aatctcggtt ttttggcaat caacaggaaa aagtcttcgg ccgaagttaa	240
tctcggttt ttggcaatca acaggtaaa gtctcgcc gaagttaatc tcgtgtttt	300
ggcaatcaa aggttaagt cttcgccga agttaatctc gtgttttgga caatcaacag	360
gtttaagtct tcggccgaan ttaatctcggtt gttttggca atcaacaggtt ttaantcttc	420
ggccgaagtt aatctcggtt ttttggcaat caana	455
<210> 274	
<211> 461	
<212> DNA	
<213> Homo sapien	
<400> 274	
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tggcaaacca aatccagcag cacatcaaaa agcttatcca ccatgatcaa gtgggcttca	120
ccccctggat gcaaggctgg ttcaacataa gaaaatcaat aaatgtaatc catcacataa	180
acagaaccaa agacaaaaac cacatgatta tctcaataga tgcagaaaag gccttggaca	240
aattcaacag cccttcatgc taaacactct taataaaacta gatattgtatg gaatgtatct	300
caaaaataata agagctattt atgacaaacc cacagccaat atcatactga atggcщаag	360
actggaaagca ttcccttga aaactggcac aagacaagga tgccctctct caccgctcct	420
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<210> 275	
<211> 729	
<212> DNA	
<213> Homo sapien	
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<221> misc_feature	
<222> (1)...(729)	
<223> n = A,T,C or G	
<400> 275	
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ctccccaac cccaccttca cagcctctc cacacgtctc ccanagattt ttgtccttca	180
tttgc当地 canggatgtt ggaagtngac atttnnagtn gcnngaaccc catcgtgaa	240
ncantaagca gaantacgt gactttgana nacanctgtat gaagaacaacn ctacnganaa	300
ccctttctnt cgttgttanga tctcnngtcc ntcaactatg cggccccctg cnngtccacc	360
atttgggaga actccccccn cggtggatcc ccccttgagt ntcccattct ngtcccccan	420
accngncttg ngngncantn cnncctcnca ccntgttcc ctgnngtnaa aatnngttt	480
nccgcnccc naattccac cnaatcaca gegaancng aaggccttcn naagtgttta	540
angcccnngng gtttctctnt ntanttgac cctacccttc cnctnnnnnt tncngttgg	600
tcgcgcctg gncncgcctn gttcctctt nnggnacaa cctngntcnn nggcncntcn	660
nnnctntcc tnnnactagc tngcctntcc ncncgcngn ncanngcaca ttncncnnac	720
tntgtnncc	729
<210> 276	
<211> 339	
<212> DNA	
<213> Homo sapien	

<400> 276
tgacctgaca tgttagtagat acttaataaa tatttgtga atgaatggat gaagtggagt 60
tacagagaaa aatagaaaag tacaaattgt tgtcagtgtt ttgaaggaaa attatgatct 120
ttcccaaagt tctgacttca ttctaagaca gggtagtat ctccatacat aattttactt 180
gctttgaaa atcaaattgag ataattctatt tagattgata atttatttag actggctata 240
aactattaag tgctagcaaa tatacattt aatctcattt tccacctt gtgatatacg 300
tatgttaggtg ttgactttaa tggatgtcag gtcaatccc 339

<210> 277
<211> 664
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(664)
<223> n = A,T,C or G

<400> 277
tgacctgaca tccataacaa aatcttttc cattatattc ttctagggga atttcttgc 60
aagcatccaa agaaacaaa ttagtggtaag accgtgcca gtggggagca gacaccaaag 120
taagaccaca gatttacat tcaacaggtt gtcacagta cttggccca cactgtggc 180
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aagagggttt tgcatcctgg tcagatnaag aagcaatggt gtgctgagga aatcccatac 300
gaataagtga gcattcagaa cttgagctag caggaggagg actaagatga tgtgtgagca 360
actctttgtt atggcttca tctaaaataa catgttacgt gccaccagg tcacgagcaa 420
gtacagtgca aacgcgaact tctgcagaca atccaataac agataactcta atttttagctg 480
cctttagggt ctgttataaa tcataaataat tagatggatc gcaagttgtt aggnntgctaa 540
aagatgatataa gtacttctcg acttgtatgt ccagcatgt ttgtttaan tctgccttag 600
nccctgctta gggaaatttt taaagaagat ggctctccat gttcanggtc aatcacnaat 660
tgcc 664

<210> 278
<211> 452
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(452)
<223> n = A,T,C or G

<400> 278
tgacctgaca ttgaggaaga gcacacacct ctgaaattcc tttaggttcag aaggcattt 60
gacacagagt gggcctctga taattcatga aatgcattct gaagtcatcc agaatggagg 120
ctgcaatctg ctgtgtttt ggggttgct cactgtgctc ctggatataca cacaaaagct 180
gcaatccttc ttcttcaact aacattttgc agtatttgct gggatttta ctgcagacat 240
gatacatagc ccatagtgcc cagagctgaa cctctgggtt agagaagttg ccaaggagcg 300
ggaaaaaatgt ctgttataaa ctataggtca ccaatgctgtt catcttacaa cttgaacttg 360
gccaaattctg tatgttgca tgcagatctt ggagaagagt acgcctctgg aagtcacggg 420
atatccaaan ctgtctgtca gatgtcaggtt ca 452

<210> 279
<211> 274

<212> DNA
<213> Homo sapien

<400> 279
ttttttttt ttcggcaagg caaatttact tctgaaaag ggtgctgctt gcactttgg 60
ccactgcgag agcacaccaa acaaagttagg gaaggggittt ttatccctaa cgcggttatt 120
ccctggttct gtgtcgtgtc cccatggct ggagtcagac tgcacaatct acactgaccc 180
aactggctac tgtttaaat tgaatatgaa taattaggta ggaaggggga ggctgttgt 240
tacggtacaa gacgtgtttg ggcatgtcag gtca 274

<210> 280
<211> 272
<212> DNA
<213> Homo sapien

<400> 280
tacctgacat ggagaaataa cttgttagtat tttgcgtgca atgaaatact atatgagggt 60
gaaaatgaat gaactagcaa tgcgtgtatc aacatgaata aatccccaaa acataataat 120
gttgaatgga aaaggtgagt ttcaagaagga tatatatgcc ctctaaatcc atttatgtaa 180
acctttaaaa aactacatta tttatggtca taagtccatc cagaaaatat taaaaaacct 240
acatgggatt gataactact gatgtcaggt ca 272

<210> 281
<211> 431
<212> DNA
<213> Homo sapien

<220>
<221> misc_feature
<222> (1)...(431)
<223> n = A,T,C or G

<400> 281
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tagcattaaat cagaaaatat tgcatacgctt ctaggcctt tagagtaggt gtgcgtcttc 180
aaatataatca tagtcccaca gtttatttca tgtatatttt ctgcctgaat cacatagaca 240
tttgaatttg caacgcctga tgtaaatata taaattctt ccaatcagaa acatagcaag 300
aaattcaggg acttggcatc yatcagggtt tgacagcana tccctgtara aacactgata 360
cacactcaca cacgtatgca acgtggagat gtcgcyttww kkktwywcwm rmrycrwcn 420
aatcacttan n 431

<210> 282
<211> 98
<212> DNA
<213> Homo sapien

<400> 282
attcgattcg atgcttggc ccaggagttc aagactgcag tgagccactg cacttcaggc 60
tggacaacag agcgagtccc tgtgccaaaa aaaaaaaaaa 98

<210> 283
<211> 764
<212> DNA

<213> Homo sapien

<220>

<221> misc_feature

<222> (1)...(764)

<223> n = A,T,C or G

<400> 283

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gggccascat tgcacagtgg astgcaaagg ttgcaggota tggcgcccta ctavtaaccc	180
cgtttttcct gtattatctg taacataata tggtagactg tcacagagcc gaatwccart	240
hacasgatga atccaawggc caygaggatg cccasaatca gggcccasat sttcaggcac	300
ttggcggtgg gggcatasgc ctgkgtccccg gtcacgtcsc caaccwtcty cctgtcccta	360
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cnctntnctn cnnatcggtc cnccnntaa ctacnctttn nacnannct cactnatncc	600
ngnnanttct ttcccttcct cccnaacgcn tgcgtgegcc cgtctngcct nnctncgna	660
cccnactt atttacctt ncacccttagc nctctacttn acccancnc tcctacctcc	720
nggnccaccc nnccctnatac nctnnctctn tcnnctcntt cccc	764

<210> 284

<211> 157

<212> DNA

<213> Homo sapien

<400> 284

caagtgttagg cacagtgtatg aaaggctgga gcaaacacaa tctgtggta attaacgttt	60
atttctcccc ttccaggaac gtcttgcatg gatgataaa gatcagctcc tggtaacat	120
aaataagcta gtttaagata cgttccctta cacttga	157

<210> 285

<211> 150

<212> DNA

<213> Homo sapien

<400> 285

attcgattgt actcagacaa caatatgcta agtggaaagaa gtcagtcaca aaagaccaca	60
tactgtatga cttcatttac attaagtgtc cagaataggc aaatccgtag agacagaaag	120
tagatgagca gctgcctagg tctgagtaca	150

<210> 286

<211> 219

<212> DNA

<213> Homo sapien

<400> 286

attcgatttt tttttttttt gccatgatga aattcttact ccctcagatt ttttgtctgg	60
ataaatgcaa gtctcaccac cagatgtgaa attacagtaa actttgaagg aatctcctga	120
gcaaccttgg ttaggatcaa tccaatattc accatctggg aagtcaggat ggctgagttg	180
caggtcttta caagttcggg ctggatttgtt ctgagtaca	219

<210> 287

<211> 196
<212> DNA
<213> Homo sapien

<400> 287
attcgattct tgaggctacc aggagctagg agaaggaggca tggaaacaaat tttccctcat 60
atccatactc agaagaacc aaccctgctg acaccttaat ttcaagcttct ggcctctaga 120
actgtgagag agtacatttc tcttggttta agccaagaga atctgtcttt tggtacttta 180
tatcatagcc tcaaga 196

<210> 288
<211> 199
<212> DNA
<213> Homo sapien

<400> 288
attcgatttc agtccagtcc cagaacccac attgtcaatt actactctgt araagattca 60
tttgtgaaa ttcatggagt aaaacattta tgatccctta atatatgc当地 attaccatgc 120
taggtactga agattaagt gaccgagatg ctggcccttg gggtcaagtg atccctctcc 180
cagagtgcac tggactgaa 199

<210> 289
<211> 182
<212> DNA
<213> Homo sapien

<400> 289
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tagtaataca gaagcaagta tctgtatatg taaacattaa aaaggtacag tggaaacttca 120
gtattataat cttagggacc accattatat atgtggtcca tcattggcca aaaaaaaaaaa 180
aa 182

<210> 290
<211> 1646
<212> DNA
<213> Homo sapien

<400> 290
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ttgaaataga agtataagtt gctaccattt ttgtataaca ttgaaagata gtatTTTacc 180
atcttaatc atctggaaa atacaagtcc tgtgaacaac cactcttca cctagcagca 240
tgaggccaaa agtaaaggct ttaaattata acatatggga ttcttagtag tatgttttt 300
tcttgaaact cagtgctct atctaaccctt actatctcct cactcttct ctaagactaa 360
actcttaggt cttaaaaatc tgcccacacc aatcttagaa gctctgaaaa gaatttgc 420
ttaaatatct tttaatgtatc acatgtatTTT tatggaccaa attgacattt tcgactatTTT 480
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gagcagaagc aaaccacatg tctcagctat attattattt atttttatg cataaaagtga	1020
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cagtgcattg acaatgggtt gatattttc tttaaaagaa aaataataatt atgaaagcca	1140
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kgcatactac atgcagttct ttaaccaatg tctgtttggc taatgttaatt aaagttgtta	1320
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ctggtaattt ttttatttac aatatgttta aagagataac agtttgatat gtttcatgt	1440
gtttatagca gaagtttattt atttctatgg cattccagcg gatattttgg tggttgcgag	1500
gcatgcagtc aatattttgt acagttatgt gacagtattc agcaacgcct gatagttct	1560
ttggccttat gttaaataaaa aagacctgtt tggtatgtat tttttatttt taaaaaaaaa	1620
aaaaaaaaaaa aaaaaaaaaaa aaaaaaaaaa	1646

<210> 291

<211> 1851

<212> DNA

<213> Homo sapien

<400> 291

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tcacttcctt taaggccttgc tgactcttcc tctgatgtca gctttaagtc ttgttctgga	180
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caaattacat gatgtatgact agaaaacagca tactctctgg ccgtcttcc agatcttgag	300
aagatacatc aacattttgc tcaagtagag ggctgactat acttgcgtat ccacaacata	360
cagcaagtat gagagcagtt cttccatatac tatccagcgc atttaaatcc gctttttct	420
tgattaaaaa tttcaccact tgctgtttt gctcatgtat accaagttagc agtgggtgtga	480
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cacaggtact gaaatcatgt catctgcggc aacatggtgg aacctaccca atcacacatc	1320
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cttttccccca ttttagtatta tggtggctgtt gggcttgcata taggtgggtt ttattacttt	1800
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<210> 292

<211> 1851

<212> DNA

<213> Homo sapien

<400> 292

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tcacttcctt	taaggcttt	tgactcttc	tctgatgtca	gtttaagt	ttgttctgga	180
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cagcaagtat	gagagcagtt	cttccatatac	tatccagcgc	atttaaattc	gttttttct	420
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ggccatgctt	gtttttgtat	tgcataatcag	caccgtataa	gagcagtgt	ttggccatta	540
atttatcttc	atttagaca	gcatagtgt	gagtggatt	tccatactca	tctgaatat	600
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ccttgc	agctgtcc	ttttgttgt	caagacatt	aagttgacat	cgtctgtcca	720
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tttgacaaa	tccagcatcc	ttgtattt	tgttg	ctc	agaggaa	1740
ctttccca	tttagtatta	ttggctgt	ggc	tttgc	tttacttt	1800
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<210> 293

<211> 668

<212> DNA

<213> Homo sapien

<400> 293

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accrtataag	agcagtgc	tggccattaa	tttatcttc	attrtagaca	gcrtagtya	180
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catatcttag	gaattcaaaa	taacattca	cagctt	caactagtt	tattaaagg	360
agaaaactca	tttttatg	atgtattgaa	atcaaacc	cctcatgc	atatagttg	420
ctactgcata	cctttatc	agctgtcc	ttttgtt	caaggacatt	aagttgacat	480
cg	gc	aggagtt	tactact	gaattcc	tggcagag	540
gcagtccat	gagagtgaga	agactttt	gaaattgt	gtgcact	tacagccata	600
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aaaaaaaaa						668

<210> 294
 <211> 1512
 <212> DNA
 <213> Homo sapien

<400> 294

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tgatctcg	cc					1512

<210> 295

<211> 1853
 <212> DNA
 <213> Homo sapien

<400> 295

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ttcaaacaga	ttggaaaccc	ggagttacct	gctagtttgtt	gaaactgggtt	ggtagacgcg	180
atctgttggc	tactactggc	ttctcctggc	tgttaaaagc	agatggtgtt	tgaggttgat	240
tccatgccgg	ctgcttcttc	tgtgaagaag	ccatttggtc	tcaggagcaa	gatgggcaag	300
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ggagaccacg	acgactctgc	tatgaagaca	ctcaggagca	agatgggcaa	gtggtgcgc	420
cactgcttcc	cctgctgcag	ggggagtggc	aagagcaacg	tggcgcttc	tggagaccac	480
gacgaytcg	ctatgaagac	actcaggaac	aagatggca	agtggtgctg	ccactgcttc	540
ccctgctgca	ggggagcrg	caagagcaag	gtggcgctt	ggggagacta	cgatgacagy	600
gccttcatgg	akcccaggta	ccacgtccrt	ggagaagatc	tggacaagct	ccacagagct	660
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aacaagargg	acaagcaaaa	gaggactgt	ctacatctgg	cctctgccaa	tggaaattca	780
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gtsgtgaard	tttaatyaa	gaaaaaaagcg	aatttaaat	gcrctggata	gatatggaag	1140
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<210> 296

<211> 2184

<212> DNA

<213> Homo sapien

<400> 296

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tttcctctga	gaactgcaac	aataaaataca	aggatgctgg	attttgcctaa	atgcctttc	180
tgtgtctgtt	gagatgctta	tgtgactttg	cttttaattc	tgtttatgtg	attatcacat	240
ttattgactt	gcctgtgtta	gaccggaaga	gctgggggtgt	ttctcaggag	ccaccgtgt	300
ctgcggcagc	ttcgggataa	cttgaggctg	catcaactggg	gaagaaaacac	aytcctgtcc	360
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taataatatt	agatagtc	aaatgaaatw	acctatgaga	ctaggcttgc	agaatcaata	1860

gatttttt ttaagaatct tttggctagg agcgggtgtct cacgcctgta attccagcac	1920
cttgagaggc tgaggtggc agatcacgag atcaggagat cgagaccatc ctggctaaca	1980
cggtaaaacc ccatctctac taaaaataca aaaacttagc tgggtgttgt ggcgggtgcc	2040
tgttagtcca gctactcagg argctgaggc aggagaatgg catgaacccg ggaggtggag	2100
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<210> 297
 <211> 1855
 <212> DNA
 <213> Homo sapien

 <220>
 <221> misc_feature
 <222> (1)...(1855)
 <223> n = A,T,C or G

<400> 297	
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gcccccccg cataaccgtc agactggcct gtaacggctt gcaggcgcac gccgcacgc	180
cgtaacggct tggctgcct gtaacggctt gcacgtgcac gctgcacgcg cgttaacggc	240
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tcttgattt acgcttcctc cttggatkga cgttccctcc ttggatkgac gtttctyty	360
tcgcgttctt ttgcgtggact tgacctttt tctgcgtgggt ttggcattcc tttgggggtgg	420
gctgggtgtt ttctccgggg gggktkgccc ttccctgggt gggcgtggk cgccccccagg	480
gggcgtggc tttcccccggg tgggtgtggg ttttctggg gtgggtgtgg ctgtgtgtgg	540
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gtaacntgtt agtttgttac actgggttggt agacgcgtac tgctgttact actgttttcc	660
ctggctgtt aaagcagatg ttggctgagg ttgattcaat gcccgtgtct tcttctgtga	720
agaagccatt tggcttcagg agcaagatgg gcaagtgggtg cgccactgtct tccctgtgt	780
caggggggagc ggcaagagca acgtgggcac ttctggagac cacaacgtact cctctgtgaa	840
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gtaccacgtc crtggagaag atctggacaa gtcacacaga gtcgttgggtt gggtaaagt	1020
ccccagaaag gatctcatcg tcatgtctcg ggacactgay gtgaacaaga rggacaagca	1080
aaagaggact gctctacatc tggctctgc caatggaaat tcagaagtag taaaactcg	1140
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aaatatttcca gatgagtatg gaaataccac tctacactat gctgtctaca atgaagataa	1320
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gatctactaa ttttatcttc aaaatactga aatgcattca ttttaacatt gacgtgtgt	1440
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acctaattt ctaagacttt attttaata ttgttatttt caaagaagca ttagagggtt	1560
cagttttttt tttttaaatg cacttctggt aaatactttt gttgaaaaca ctgaatttgc	1620
aaaaggtaat acttactatt tttcaatttt tccctcttag gattttttc ccctaatgaa	1680
tgtaaatggg caaaaatttgc cctgaaatag gttttacatg aaaactccaa gaaaagttaa	1740
acatgtttca gtgaatagag atcctgctcc tttggcaagt tcctaaaaaaa cagtaataga	1800
tacgaggtga tgccctgtc agtggcaagg tttaaagatat ttctgtatctc gtgcc	1855

<210> 298
 <211> 1059
 <212> DNA
 <213> Homo sapien

<400> 298

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gcgcttgrgg agactmcgat gacagygcct tcatggagcc caggtaccac gtccgtggag 180
aagatctgga caagctccac agagctgccc tggggggta aagtccccag aaaggatctc 240
atcgctcatgc tcagggacac tgaygtAAC aagarggaca agaaaagag gactgtctta 300
catctggcct ctgccaatgg gaattcagaa gtagtaaaac tcstgctgga cagacgatgt 360
caacttaatg tccttgacaa caaaaagagg acagctctga yaaaggccgt acaatgccag 420
gaagatgaat gtgggttaat gttgctggaa catggcactg atccaaatat tccagatgag 480
tatggaaata ccactctrca ctaygctrtc tayaatgaag ataaattaat ggc当地
ctgcttttat aygggtctga tatcgaatca aaaaacaagg tatagatcta ctaattttat 600
cttcaaaata ctgaaatgca ttcatTTAA cattgacgtg tgtaaggcc agtcttccgt 660
atTTGGAAAGC tcaagcataa ctTGAATGAA aatatttga aatgaccta ttatctaaga 720
ctttattttA aatattgtta ttttcaaaaAGC agcattagag ggtacagttt tttttttta 780
aatgcacttc tggtaaatac ttttgtgaa aacactgaat ttgtAAAAGG taataacttac 840
tatttttcaa ttttccctc ctaggatttt tttcccctaa tgaatgtaa atggcaaaat 900
ttgcctgaa ataggTTTA catgaaaact ccaagaaaag ttaaacatgt ttcaagtgaat 960
agagatcctg ctccTTggc aagttcctaa aaaacagtaa tagatacggag gtgatgcgc 1020
tqtcactqgc aqgTTTaaq atatttctqa ttcqtcqcc 1059

<210> 299

<211> 329

<212> PRT

<213> Homo sapien

<400> 299

Met	Asp	Ile	Val	Val	Ser	Gly	Ser	His	Pro	Leu	Trp	Val	Asp	Ser	Phe
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Leu	His	Leu	Ala	Gly	Ser	Asp	Leu	Leu	Ser	Arg	Ser	Leu	Met	Ala	Glu
				20				25					30		
Glu	Tyr	Thr	Ile	Val	His	Ala	Ser	Phe	Ile	Ser	Cys	Ile	Ser	Ser	Ser
					35			40				45			
Leu	Asp	Gly	Gln	Gly	Glu	Arg	Gln	Glu	Gln	Arg	Gly	His	Phe	Trp	Arg
					50			55			60				
Pro	Gln	Arg	Leu	Leu	Cys	Glu	Asp	Ala	Trp	Glu	Gln	Glu	Val	Gln	Val
					65			70			75			80	
Val	Leu	Pro	Leu	Leu	Pro	Leu	Leu	Gln	Gly	Ser	Gly	Lys	Ser	Asn	Val
					85				90				95		
Val	Ala	Trp	Gly	Asp	Tyr	Asp	Asp	Ser	Ala	Phe	Met	Asp	Pro	Arg	Tyr
					100				105				110		
His	Val	His	Gly	Glu	Asp	Leu	Asp	Lys	Leu	His	Arg	Ala	Ala	Trp	Trp
					115			120			125				
Gly	Lys	Val	Pro	Arg	Lys	Asp	Leu	Ile	Val	Met	Leu	Arg	Asp	Thr	Asp
					130			135			140				
Val	Asn	Lys	Arg	Asp	Lys	Gln	Lys	Arg	Thr	Ala	Leu	His	Leu	Ala	Ser
					145			150			155			160	
Ala	Asn	Gly	Asn	Ser	Glu	Val	Val	Lys	Leu	Val	Leu	Asp	Arg	Arg	Cys
					165				170				175		
Gln	Leu	Asn	Val	Leu	Asp	Asn	Lys	Lys	Arg	Thr	Ala	Leu	Thr	Lys	Ala
					180			185				190			
Val	Gln	Cys	Gln	Glu	Asp	Glu	Cys	Ala	Leu	Met	Leu	Leu	Glu	His	Gly
					195			200			205				
Thr	Asp	Pro	Asn	Ile	Pro	Asp	Glu	Tyr	Gly	Asn	Thr	Thr	Leu	His	Tyr

210	215	220
Ala Val Tyr Asn Glu Asp Lys Leu Met Ala Lys Ala	Leu Leu Leu Tyr	
225	230	235
Gly Ala Asp Ile Glu Ser Lys Asn Lys His Gly	Leu Thr Pro Leu	240
245	250	255
Leu Gly Ile His Glu Gln Lys Gln Gln Val Val Lys Phe	Leu Ile Lys	
260	265	270
Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg Tyr Gly	Arg Thr Ala Leu	
275	280	285
Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile Val	Ser Pro Leu	Leu
290	295	300
Glu Gln Asn Val Asp Val Ser Ser Gln Asp Leu	Glu Arg Arg Pro	Glu
305	310	315
Ser Met Leu Phe Leu Val Ile Ile Met		320
		325

<210> 300

<211> 148

<212> PRT

<213> Homo sapien

<220>

<221> VARIANT

<222> (1)...(148)

<223> Xaa = Any Amino Acid

<400> 300

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Trp Thr Ser Ser Thr Glu Leu Pro Trp Trp Gly Lys Val	Pro Arg Lys		
20	25	30	
Asp Leu Ile Val Met Leu Arg Asp Thr Asp Val Asn Lys	Xaa Asp Lys		
35	40	45	
Gln Lys Arg Thr Ala Leu His Leu Ala Ser Ala Asn Gly	Asn Ser Glu		
50	55	60	
Val Val Lys Leu Xaa Leu Asp Arg Arg Cys Gln Leu Asn	Val Leu Asp		
65	70	75	80
Asn Lys Lys Arg Thr Ala Leu Xaa Lys Ala Val Gln Cys	Gln Glu Asp		
85	90	95	
Glu Cys Ala Leu Met Leu Leu Glu His Gly Thr Asp Pro	Asn Ile Pro		
100	105	110	
Asp Glu Tyr Gly Asn Thr Thr Leu His Tyr Ala Xaa Tyr	Asn Glu Asp		
115	120	125	
Lys Leu Met Ala Lys Ala Leu Leu Leu Tyr Gly Ala Asp	Ile Glu Ser		
130	135	140	
Lys Asn Lys Val			
145			

<210> 301

<211> 1155

<212> DNA

<213> Homo sapien

<400> 301

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agcaacgtgg gcacttctgg agaccacgac gactctgcta tgaagacact caggagcaag	180
atggcaagt ggtgcccca ctgcttccccct tgctgcagggg ggagtggcaa gagcaacgtg	240
ggcgcttcg gagaccacga cgactctgtt atgaagacac tcaggaacaa gatgggcaag	300
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ggagactacg atgacagtgc cttcatggag cccaggtacc acgtccgtgg agaagatctg	420
gacaagctcc acagagctgc ctgggtgggt aaagtccccca gaaaggatct catgtcatg	480
ctcaggggaca ctgacgtgaa caagaaggac aagcaaaaaga ggactgtct acatctggcc	540
tctgccaatg ggaattcaga agtagtaaaa ctccctgctgg acagacgtat tcaacttaat	600
gtccttgaca acaaaaagag gacagctctg ataaaggccg tacaatgcca ggaagatgaa	660
tgtgcgttaa tgggtctggaa acatggact gatccaaata ttccagatga gtatggaaat	720
accactctgc actacgttat ctataatgaa gataaattaa tggccaaagc actgtctta	780
tatgggtctg atatcgatc aaaaaacaag catggcctca caccactgtt acttgggtgt	840
catgagcaaa aacagcaagt cgtgaaattttttaatcaaga aaaaagcgaa tttaaatgca	900
ctggatagat atggaaggac tgctctcata cttgtgtat gttgtggatc agcaagtata	960
gtcagccttc tacttgagca aaatattgtt gtatcttctc aagatctatc tggacagacg	1020
gccagagagt atgctgtttc tagtcatcat catgtatcattt gccagttact ttctgactac	1080
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accagaaaaataataa	1155

<210> 302
<211> 2000
<212> DNA
<213> Homo sapien

<400> 302	
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agcaacgtgg gcacttctgg agaccacgac gactctgcta tgaagacact caggagcaag	180
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ggcgcttcg gagaccacga cgactctgtt atgaagacac tcaggaacaa gatgggcaag	300
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ggagactacg atgacagtgc cttcatggag cccaggtacc acgtccgtgg agaagatctg	420
gacaagctcc acagagctgc ctgggtgggt aaagtccccca gaaaggatct catgtcatg	480
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attctgattc	atgaagaaaa	gcagatagaa	gtggtgaaa	aatgaattc	tgagcttct	1860
cttagttgt	agaaagaaaa	agacatctt	catgaaaata	gtacgttgcg	ggaagaaatt	1920
gccatgctaa	gactggagct	agacacaatg	aaacatcaga	gccagctaaa	aaaaaaaaaa	1980
aaaaaaaaaa	aaaaaaaaaa					2000

<210> 303

<211> 2040

<212> DNA

<213> Homo sapien

<400> 303

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ggcgcttctg	gagaccacga	cgactctgt	atgaagacac	tcaggaacaa	gatgggcaag	300
tggtgctgcc	actgctccc	ctgctgcagg	gggagccgca	agagcaaggt	ggcgcttgg	360
ggagactacg	atgacagtgc	cttcatggag	cccaggtacc	acgtccgtgg	agaagatctg	420
gacaagctcc	acagagctgc	ctgggtgggt	aaagtcccc	gaaaggatct	catcgctatg	480
ctcaggggaca	ctgacgtgaa	caagaaggac	aagcaaaaga	ggactgctc	acatctggcc	540
tctgccaatg	ggaattcaga	agtagtaaaa	ctcctgctgg	acagacgatg	tcaacttaat	600
gtccttgaca	acaaaaagag	gacagctctg	ataaaggccg	tacaatgca	ggaagatgaa	660
tgtgcgttaa	tgttgctgga	acatggact	gatccaaata	ttccagatga	gtatggaaat	720
accactctgc	actacgctat	ctataatgaa	gataaattaa	tggccaaagc	actgcttta	780
tatggtgctg	atatcgaaat	aaaaaacaag	catggcctca	caccactgtt	acttggtgta	840
catgagcaaa	aacagcaagt	cgtgaaattt	ttaatcaaga	aaaaagcgaa	tttaaatgca	900
ctggatagat	atggaggac	tgctctcata	cttgcgtat	gttgtggatc	agcaagtata	960
gtcagccttc	tacttgagca	aaatattgtat	gtatcttc	aagatctatc	tggacagacg	1020
gccagagagt	atgcgtttc	tagtcatcat	catgtaattt	gccagttact	ttctgactac	1080
aaagaaaaac	agatgctaaa	aatctttct	gaaaacagca	atccagaaca	agacttaaag	1140
ctgacatcatg	aggaagagtc	acaaagggtc	aaaggcagt	aaaatagcca	gccagagaaa	1200
atgtctcaag	aaccagaaat	aaataaggat	ggtgatagag	aggttgaaga	agaaatgaag	1260
aagcatgaaa	gtaataatgt	gggattacta	gaaaacctga	ctaattgggt	cactgctggc	1320
aatggtgata	atggattaat	tcctcaaagg	aagagcagaa	cacctgaaaa	tcagcaattt	1380
cctgacaacg	aaagtgaaga	gtatcacaga	atttgcgaat	tagttctga	ctacaaagaa	1440
aaacagatgc	caaataactc	ttctgaaaac	agcaacccag	aacaagactt	aaagctgaca	1500
tcagaggaag	agtccaaaag	gcttgagg	agtggaaatg	gccagccaga	gaaaagatct	1560
caagaaccag	aaataaataa	ggatggtgat	agagagctag	aaaattttat	ggctatcgaa	1620
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cagcaatttc	ctgacactga	gaatgaagag	tatcacagt	acgaacaaaa	tgataactcg	1800
aagcaatttt	gtgaagaaca	gaacactgga	atattacacg	atgagattct	gattcatgaa	1860
gaaaagcaga	tagaagtgg	tgaaaaatg	aattctgagc	tttctttag	ttgtaaagaaa	1920
gaaaagaca	tcttgcata	aaatagtacg	ttgcggaaag	aaattgccat	gctaagactg	1980
gagctagaca	caatgaaaca	tcagagccag	ctaaaaaaa	aaaaaaaaaa	aaaaaaaaaa	2040

<210> 304

<211> 384

<212> PRT

<213> Homo sapien

<400> 304

Met	Val	Val	Glu	Val	Asp	Ser	Met	Pro	Ala	Ala	Ser	Ser	Val	Lys	
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Pro	Phe	Gly	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp	Cys	Cys	Arg	Cys	Phe
			20				25						30		
Pro	Cys	Cys	Arg	Glu	Ser	Gly	Lys	Ser	Asn	Val	Gly	Thr	Ser	Gly	Asp
			35				40					45			
His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp
			50			55					60				
Cys	Arg	His	Cys	Phe	Pro	Cys	Arg	Gly	Ser	Gly	Lys	Ser	Asn	Val	
			65			70			75			80			
Gly	Ala	Ser	Gly	Asp	His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Asn
			85					90				95			
Lys	Met	Gly	Lys	Trp	Cys	Cys	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser
			100				105					110			
Gly	Lys	Ser	Lys	Val	Gly	Ala	Trp	Gly	Asp	Tyr	Asp	Asp	Ser	Ala	Phe
			115				120			125					
Met	Glu	Pro	Arg	Tyr	His	Val	Arg	Gly	Glu	Asp	Leu	Asp	Lys	Leu	His
			130				135				140				
Arg	Ala	Ala	Trp	Trp	Gly	Lys	Val	Pro	Arg	Lys	Asp	Leu	Ile	Val	Met
			145				150			155			160		
Leu	Arg	Asp	Thr	Asp	Val	Asn	Lys	Lys	Asp	Lys	Gln	Lys	Arg	Thr	Ala
					165				170			175			
Leu	His	Leu	Ala	Ser	Ala	Asn	Gly	Asn	Ser	Glu	Val	Val	Lys	Leu	Leu
			180				185				190				
Leu	Asp	Arg	Arg	Cys	Gln	Leu	Asn	Val	Leu	Asp	Asn	Lys	Lys	Arg	Thr
			195				200			205					
Ala	Leu	Ile	Lys	Ala	Val	Gln	Cys	Gln	Glu	Asp	Glu	Cys	Ala	Leu	Met
			210				215			220					
Leu	Leu	Glu	His	Gly	Thr	Asp	Pro	Asn	Ile	Pro	Asp	Glu	Tyr	Gly	Asn
			225				230			235			240		
Thr	Thr	Leu	His	Tyr	Ala	Ile	Tyr	Asn	Glu	Asp	Lys	Leu	Met	Ala	Lys
					245			250			255				
Ala	Leu	Leu	Leu	Tyr	Gly	Ala	Asp	Ile	Glu	Ser	Lys	Asn	Lys	His	Gly
			260				265			270					
Leu	Thr	Pro	Leu	Leu	Gly	Val	His	Glu	Gln	Lys	Gln	Gln	Val	Val	
			275				280			285					
Lys	Phe	Leu	Ile	Lys	Lys	Ala	Asn	Leu	Asn	Ala	Leu	Asp	Arg	Tyr	
			290				295			300					
Gly	Arg	Thr	Ala	Leu	Ile	Leu	Ala	Val	Cys	Cys	Gly	Ser	Ala	Ser	Ile
			305				310			315			320		
Val	Ser	Leu	Leu	Leu	Glu	Gln	Asn	Ile	Asp	Val	Ser	Ser	Gln	Asp	Leu
					325				330			335			
Ser	Gly	Gln	Thr	Ala	Arg	Glu	Tyr	Ala	Val	Ser	Ser	His	His	His	Val
			340				345				350				
Ile	Cys	Gln	Leu	Leu	Ser	Asp	Tyr	Lys	Glu	Lys	Gln	Met	Leu	Lys	Ile
			355				360			365					
Ser	Ser	Glu	Asn	Ser	Asn	Pro	Glu	Asn	Val	Ser	Arg	Thr	Arg	Asn	Lys
			370				375			380					

<210> 305

<211> 656

<212> PRT

<213> Homo sapien

<400> 305

Met	Val	Val	Glu	Val	Asp	Ser	Met	Pro	Ala	Ala	Ser	Ser	Val	Lys	Lys
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Pro	Phe	Gly	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp	Cys	Cys	Arg	Cys	Phe
	20					25							30		
Pro	Cys	Cys	Arg	Glu	Ser	Gly	Lys	Ser	Asn	Val	Gly	Thr	Ser	Gly	Asp
	35					40						45			
His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Ser	Lys	Met	Gly	Lys	Trp
	50					55						60			
Cys	Arg	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser	Gly	Lys	Ser	Asn	Val
	65					70					75			80	
Gly	Ala	Ser	Gly	Asp	His	Asp	Asp	Ser	Ala	Met	Lys	Thr	Leu	Arg	Asn
						85					90			95	
Lys	Met	Gly	Lys	Trp	Cys	Cys	His	Cys	Phe	Pro	Cys	Cys	Arg	Gly	Ser
						100				105			110		
Gly	Lys	Ser	Lys	Val	Gly	Ala	Trp	Gly	Asp	Tyr	Asp	Asp	Ser	Ala	Phe
						115				120			125		
Met	Glu	Pro	Arg	Tyr	His	Val	Arg	Gly	Glu	Asp	Leu	Asp	Lys	Leu	His
						130			135			140			
Arg	Ala	Ala	Trp	Trp	Gly	Lys	Val	Pro	Arg	Lys	Asp	Leu	Ile	Val	Met
	145					150				155			160		
Leu	Arg	Asp	Thr	Asp	Val	Asn	Lys	Lys	Asp	Lys	Gln	Lys	Arg	Thr	Ala
						165				170			175		
Leu	His	Leu	Ala	Ser	Ala	Asn	Gly	Asn	Ser	Glu	Val	Val	Lys	Leu	Leu
						180				185			190		
Leu	Asp	Arg	Arg	Cys	Gln	Leu	Asn	Val	Leu	Asp	Asn	Lys	Lys	Arg	Thr
						195			200			205			
Ala	Leu	Ile	Lys	Ala	Val	Gln	Cys	Gln	Glu	Asp	Glu	Cys	Ala	Leu	Met
						210			215			220			
Leu	Leu	Glu	His	Gly	Thr	Asp	Pro	Asn	Ile	Pro	Asp	Glu	Tyr	Gly	Asn
	225					230				235			240		
Thr	Thr	Leu	His	Tyr	Ala	Ile	Tyr	Asn	Glu	Asp	Lys	Leu	Met	Ala	Lys
						245				250			255		
Ala	Leu	Leu	Leu	Tyr	Gly	Ala	Asp	Ile	Glu	Ser	Lys	Asn	Lys	His	Gly
						260			265			270			
Leu	Thr	Pro	Leu	Leu	Leu	Gly	Val	His	Glu	Gln	Lys	Gln	Gln	Val	Val
						275			280			285			
Lys	Phe	Leu	Ile	Lys	Lys	Ala	Asn	Leu	Asn	Ala	Leu	Asp	Arg	Tyr	
						290			295			300			
Gly	Arg	Thr	Ala	Leu	Ile	Leu	Ala	Val	Cys	Cys	Gly	Ser	Ala	Ser	Ile
	305					310				315			320		
Val	Ser	Leu	Leu	Leu	Glu	Gln	Asn	Ile	Asp	Val	Ser	Ser	Gln	Asp	Leu
						325				330			335		
Ser	Gly	Gln	Thr	Ala	Arg	Glu	Tyr	Ala	Val	Ser	Ser	His	His	His	Val
						340			345			350			
Ile	Cys	Gln	Leu	Leu	Ser	Asp	Tyr	Lys	Glu	Lys	Gln	Met	Leu	Lys	Ile
						355			360			365			
Ser	Ser	Glu	Asn	Ser	Asn	Pro	Glu	Gln	Asp	Leu	Lys	Leu	Thr	Ser	Glu
						370			375			380			
Glu	Glu	Ser	Gln	Arg	Phe	Lys	Gly	Ser	Glu	Asn	Ser	Gln	Pro	Glu	Lys
	385					390				395			400		
Met	Ser	Gln	Glu	Pro	Glu	Ile	Asn	Lys	Asp	Gly	Asp	Arg	Glu	Vlu	Glu
						405				410			415		

Glu Glu Met Lys Lys His Glu Ser Asn Asn Val Gly Leu Leu Glu Asn
 420 425 430
 Leu Thr Asn Gly Val Thr Ala Gly Asn Gly Asp Asn Gly Leu Ile Pro
 435 440 445
 Gln Arg Lys Ser Arg Thr Pro Glu Asn Gln Gln Phe Pro Asp Asn Glu
 450 455 460
 Ser Glu Glu Tyr His Arg Ile Cys Glu Leu Val Ser Asp Tyr Lys Glu
 465 470 475 480
 Lys Gln Met Pro Lys Tyr Ser Ser Glu Asn Ser Asn Pro Glu Gln Asp
 485 490 495
 Leu Lys Leu Thr Ser Glu Glu Ser Gln Arg Leu Glu Gly Ser Glu
 500 505 510
 Asn Gly Gln Pro Glu Leu Glu Asn Phe Met Ala Ile Glu Glu Met Lys
 515 520 525
 Lys His Gly Ser Thr His Val Gly Phe Pro Glu Asn Leu Thr Asn Gly
 530 535 540
 Ala Thr Ala Gly Asn Gly Asp Asp Gly Leu Ile Pro Pro Arg Lys Ser
 545 550 555 560
 Arg Thr Pro Glu Ser Gln Gln Phe Pro Asp Thr Glu Asn Glu Glu Tyr
 565 570 575
 His Ser Asp Glu Gln Asn Asp Thr Gln Lys Gln Phe Cys Glu Glu Gln
 580 585 590
 Asn Thr Gly Ile Leu His Asp Glu Ile Leu Ile His Glu Glu Lys Gln
 595 600 605
 Ile Glu Val Val Glu Lys Met Asn Ser Glu Leu Ser Leu Ser Cys Lys
 610 615 620
 Lys Glu Lys Asp Ile Leu His Glu Asn Ser Thr Leu Arg Glu Glu Ile
 625 630 635 640
 Ala Met Leu Arg Leu Glu Leu Asp Thr Met Lys His Gln Ser Gln Leu
 645 650 655

<210> 306

<211> 671

<212> PRT

<213> Homo sapien

<400> 306

Met Val Val Glu Val Asp Ser Met Pro Ala Ala Ser Ser Val Lys Lys
 1 5 10 15
 Pro Phe Gly Leu Arg Ser Lys Met Gly Lys Trp Cys Cys Arg Cys Phe
 20 25 30
 Pro Cys Cys Arg Glu Ser Gly Lys Ser Asn Val Gly Thr Ser Gly Asp
 35 40 45
 His Asp Asp Ser Ala Met Lys Thr Leu Arg Ser Lys Met Gly Lys Trp
 50 55 60
 Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Val
 65 70 75 80
 Gly Ala Ser Gly Asp His Asp Asp Ser Ala Met Lys Thr Leu Arg Asn
 85 90 95
 Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly Ser
 100 105 110
 Gly Lys Ser Lys Val Gly Ala Trp Gly Asp Tyr Asp Asp Ser Ala Phe
 115 120 125
 Met Glu Pro Arg Tyr His Val Arg Gly Glu Asp Leu Asp Lys Leu His

130	135	140
Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp	Leu Ile Val Met	
145	150	155
Leu Arg Asp Thr Asp Val Asn Lys Lys Asp	Lys Gln Lys Arg Thr Ala	160
165	170	175
Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu	Val Val Lys Leu Leu	
180	185	190
Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp	Asn Lys Lys Arg Thr	
195	200	205
Ala Leu Ile Lys Ala Val Gin Cys Gln Glu Asp	Glu Cys Ala Leu Met	
210	215	220
Leu Leu Glu His Gly Thr Asp Pro Asn Ile Pro	Asp Glu Tyr Gly Asn	
225	230	235
Thr Thr Leu His Tyr Ala Ile Tyr Asn Glu Asp	Lys Leu Met Ala Lys	240
245	250	255
Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser	Lys Asn Lys His Gly	
260	265	270
Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln	Lys Gln Gln Val Val	
275	280	285
Lys Phe Leu Ile Lys Lys Ala Asn Leu Asn Ala	Leu Asp Arg Tyr	
290	295	300
Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys	Gly Ser Ala Ser Ile	
305	310	315
Val Ser Leu Leu Leu Glu Gln Asn Ile Asp Val	Ser Ser Gln Asp Leu	320
325	330	335
Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser	Ser His His His Val	
340	345	350
Ile Cys Gln Leu Leu Ser Asp Tyr Lys Glu Lys	Gln Met Leu Lys Ile	
355	360	365
Ser Ser Glu Asn Ser Asn Pro Glu Gln Asp Leu	Lys Leu Thr Ser Glu	
370	375	380
Glu Glu Ser Gln Arg Phe Lys Gly Ser Glu Asn	Ser Gln Pro Glu Lys	
385	390	395
Met Ser Gln Glu Pro Glu Ile Asn Lys Asp Gly	Asp Arg Glu Val Glu	400
405	410	415
Glu Glu Met Lys Lys His Glu Ser Asn Asn Val	Gly Leu Leu Glu Asn	
420	425	430
Leu Thr Asn Gly Val Thr Ala Gly Asn Gly Asp	Asn Gly Leu Ile Pro	
435	440	445
Gln Arg Lys Ser Arg Thr Pro Glu Asn Gln Gln	Phe Pro Asp Asn Glu	
450	455	460
Ser Glu Glu Tyr His Arg Ile Cys Glu Leu Val	Ser Asp Tyr Lys Glu	
465	470	475
Lys Gln Met Pro Lys Tyr Ser Ser Glu Asn Ser	Asn Pro Glu Gln Asp	
485	490	495
Leu Lys Leu Thr Ser Glu Glu Glu Ser Gln Arg	Leu Glu Gly Ser Glu	
500	505	510
Asn Gly Gln Pro Glu Lys Arg Ser Gln Glu Pro	Glu Ile Asn Lys Asp	
515	520	525
Gly Asp Arg Glu Leu Glu Asn Phe Met Ala Ile	Glu Glu Met Lys Lys	
530	535	540
His Gly Ser Thr His Val Gly Phe Pro Glu Asn	Leu Thr Asn Gly Ala	
545	550	555
Thr Ala Gly Asn Gly Asp Asp Gly Leu Ile Pro	Pro Pro Arg Lys Ser Arg	560

	565	570	575
Thr Pro Glu Ser Gln Gln Phe Pro Asp Thr Glu Asn Glu Glu Tyr His			
580	585	590	
Ser Asp Glu Gln Asn Asp Thr Gln Lys Gln Phe Cys Glu Glu Gln Asn			
595	600	605	
Thr Gly Ile Leu His Asp Glu Ile Leu Ile His Glu Glu Lys Gln Ile			
610	615	620	
Glu Val Val Glu Lys Met Asn Ser Glu Leu Ser Leu Ser Cys Lys Lys			
625	630	635	640
Glu Lys Asp Ile Leu His Glu Asn Ser Thr Leu Arg Glu Glu Ile Ala			
645	650	655	
Met Leu Arg Leu Glu Leu Asp Thr Met Lys His Gln Ser Gln Leu			
660	665	670	

<210> 307

<211> 800

<212> DNA

<213> Homo sapien

<400> 307

atkagcttcc gcttctgaca acactagaga tccctccccct ccctcagggt atggccctcc	60
acttcatttt tggtacataa catctttata ggacagggggt aaaatcccaa tactaacagg	120
agaatgctta ggactctaac aggttttga gaatgtgttg gtaagggcca ctcaatccaa	180
ttttcttgg tcctccttgt ggtcttaggag gacaggcaag ggtgcagatt ttcaagaatg	240
catcagtaag ggccactaaa tccgaccctc ctcgttcctc cttgtggct gggagggaaaa	300
ctagtgttcc tgggtgttg tcaagtgc caactattcc gatcagcagg gtccaggac	360
cactgcagg tctgggcag ggggagaaac aaaacaaacc aaaaccatgg gcrgtttgt	420
ctttcagatg gaaacactc aggcatcaac aggctcacct ttgaaatgca tcctaagcca	480
atgggacaaaa tttgacccac aaaccctgga aaaagaggtg gctcattttt tttgcactat	540
ggcttggccc caacattctc tctctgatgg ggaaaaatgg ccacctgagg gaagtacaga	600
ttacaatact atcctgcagc ttgaccttt ctgtaagagg gaaggcaaat ggagtgaaat	660
accttatgtc caagtttct tttcattgaa ggagaataca ctatgcaaag cttgaaattt	720
acatcccaca ggaggacctc tcagcttacc cccatatctc agcctcccta tagctccc	780
tcctatttagt gataagctc	800

<210> 308

<211> 102

<212> PRT

<213> Homo sapien

<220>

<221> VARIANT

<222> (1)...(102)

<223> Xaa = Any Amino Acid

<400> 308

Met Gly Xaa Phe Val Phe Gln Met Gly Asn Thr Gln Ala Ser Thr Gly			
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Ser Pro Leu Lys Cys Ile Leu Ser Gln Trp Asp Lys Phe Asp Pro Gln			
20	25	30	
Thr Leu Glu Lys Glu Val Ala His Phe Phe Cys Thr Met Ala Trp Pro			
35	40	45	
Gln His Ser Leu Ser Asp Gly Glu Lys Trp Pro Pro Glu Gly Ser Thr			
50	55	60	

Asp Tyr Asn Thr Ile Leu Gln Leu Asp Leu Phe Cys Lys Arg Glu Gly
 65 70 75 80
 Lys Trp Ser Glu Ile Pro Tyr Val Gln Ala Phe Phe Ser Leu Lys Glu
 85 90 95
 Asn Thr Leu Cys Lys Ala
 100

<210> 309
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 309
 Leu Met Ala Glu Glu Tyr Thr Ile Val
 1 5

<210> 310
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 310
 Lys Leu Met Ala Lys Ala Leu Leu Leu
 1 5

<210> 311
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 311
 Gly Leu Thr Pro Leu Leu Leu Gly Ile
 1 5

<210> 312
 <211> 10
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Made in the lab

<400> 312
 Lys Leu Val Leu Asp Arg Arg Cys Gln Leu
 1 5 10

<210> 313
<211> 1852
<212> DNA
<213> Homo sapiens

<400> 313
ggcacgagaa ttaaaaaccct cagcaaaaca ggcatagaag ggacatacct taaagtaata 60
aaaaccaccc atgacaagcc cacagccaac ataataactaa atggggaaaaa gttagaagca 120
tttcctctga gaactgcaac aataaataca aggatgctgg attttgtcaa atgcctttc 180
tgtgtctgtt gagatgctta tgtactttt cttaatttc tgtttatgtt attatcacat 240
ttattgactt gcctgtgtt gaccggaaga gctgggtgt ttctcaggag ccaccgtgt 300
ctgcggcagc ttccggataa cttgaggctg catcactggg gaagaaaacac aytccgttcc 360
gtggcgctga tggctgagga cagagcttca gtgtgcttc tctgcactg gcttcgttcc 420
ggagttttc cttcatagtt catccatatg gctccagagg aaaattatat tatTTTgtta 480
tggatgaaga gtattacgtt gtgcagatat actgcagtgtt cttcatctc ttagtgttca 540
ttgggttaggt tccaccatgt tgccgcagat gacatgattt ctttgcactgt gtctggctga 600
aaagtgtttt tttgtgaatg gatattgtgg tttctggatc tcatttcctgt tgggtggaca 660
gctttctcca ctttgcgttca agtgacactgc tttccatgtt gttatggctt gaggagtata 720
ccatcggtca tgcattttc atttccgtca tttcttcctc ctttgcgttca cagggggagc 780
ggcaagagca acgtggcac ttctggagac cacaacgact ctttgcgttca gacgcttggg 840
agcaagaggt gcaagtgggtt ctgcactgc ttccatgtt gcaaggggggagc cggcaagagc 900
aacgtggtcg ctttggggaga ctacgtatc agcgccttca tggatcccac gtaccacgtc 960
catggagaag atctggacaa gttccacaga gtttgcgttca ggggtaaagt cccccagaaag 1020
gatctcatcg tcatgttca ggtttttttt gttttttttt gttttttttt gttttttttt 1080
gctctacatc tggctctgc caatggaaat tcagaatgtt taaaactcgtt gctggacaga 1140
cgatgttcaac ttaatgttca ttgtttttttt tttttttttt tttttttttt 1200
tgccaggaag atgaatgttca ttgtttttttt tttttttttt tttttttttt 1260
gatgtttttt gaaataccac tttttttttt tttttttttt tttttttttt 1320
aaagcactgc tttttttttt tttttttttt tttttttttt tttttttttt 1380
ctgtttttttt gtatcatgtt gttttttttt tttttttttt tttttttttt 1440
gcaatttttt atgcgttca gttttttttt tttttttttt tttttttttt 1500
ggatcagcaa gtatgttca gttttttttt tttttttttt tttttttttt 1560
ctggaaagac gttttttttt tttttttttt tttttttttt tttttttttt 1620
tttctgttca caaagaaaaaa cttttttttt tttttttttt tttttttttt 1680
aagactttttt gttttttttt tttttttttt tttttttttt tttttttttt 1740
agccagagct agaagatttt tttttttttt tttttttttt tttttttttt 1800
tggattttttt agaaaacccctt actaaccgttcc ctttgcgttca tttttttttt 1852

<210> 314
<211> 879
<212> DNA
<213> Homo sapiens

<400> 314
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aacgtggca ctttgcgttca ctttgcgttca tttttttttt tttttttttt 120
tgcaagtgggtt gttttttttt tttttttttt tttttttttt tttttttttt 180
gcttggggag actacatgtt gttttttttt tttttttttt tttttttttt 240
gatctggaca agtccacacg agtccgttca tttttttttt tttttttttt 300
gtcatgttca gggacacggc tttttttttt tttttttttt tttttttttt 360
ctggccctgtt cttttttttt tttttttttt tttttttttt tttttttttt 420
cttaatgttca ttgtttttttt tttttttttt tttttttttt tttttttttt 480
gatgtttttt gttttttttt tttttttttt tttttttttt tttttttttt 540

ggaaataccta ctctacacta tgctgtctac aatgaagata aattaatggc caaaggactg 600
 ctcttatacg gtgctgatat cgaatcaaaa aacaagcatg gcctcacacc actgctactt 660
 ggtatacatg agcaaaaaca gcaagtggtg aaatttttaa tcaagaaaaa agcgaattta 720
 aatgcgctgg atagatatgg aagaactgct ctcatacttg ctgtatgttg tggatcagca 780
 agtatagtca gccctctact tgagcaaaat gttgatgtat cttctcaaga tctggaaaga 840
 cggccagaga gtatgctgtt tctagtcatc atcatgtaa 879

<210> 315

<211> 292

<212> PRT

<213> Homo sapiens

<400> 315

Met His Leu Ser Phe Pro Ala Phe Leu Pro Pro Trp Met Asp Arg Gly			
5	10	15	

Ser Gly Lys Ser Asn Val Gly Thr Ser Gly Asp His Asn Asp Ser Ser			
20	25	30	

Val Lys Thr Leu Gly Ser Lys Arg Cys Lys Trp Cys Cys His Cys Phe			
35	40	45	

Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn Val Val Ala Trp Gly Asp			
50	55	60	

Tyr Asp Asp Ser Ala Phe Met Asp Pro Arg Tyr His Val His Gly Glu			
65	70	75	80

Asp Leu Asp Lys Leu His Arg Ala Ala Trp Trp Gly Lys Val Pro Arg			
85	90	95	

Lys Asp Leu Ile Val Met Leu Arg Asp Thr Asp Val Asn Lys Arg Asp			
100	105	110	

Lys Gln Lys Arg Thr Ala Leu His Leu Ala Ser Ala Asn Gly Asn Ser			
115	120	125	

Glu Val Val Lys Leu Val Leu Asp Arg Arg Cys Gln Leu Asn Val Leu			
130	135	140	

Asp Asn Lys Lys Arg Thr Ala Leu Thr Lys Ala Val Gln Cys Gln Glu			
145	150	155	160

Asp Glu Cys Ala Leu Met Leu Leu Glu His Gly Thr Asp Pro Asn Ile			
165	170	175	

Pro Asp Glu Tyr Gly Asn Thr Thr Leu His Tyr Ala Val Tyr Asn Glu			
180	185	190	

Asp Lys Leu Met Ala Lys Ala Leu Leu Tyr Gly Ala Asp Ile Glu			
195	200	205	

Ser Lys Asn Lys His Gly Leu Thr Pro Leu Leu Gly Ile His Glu			
210	215	220	

Gln Lys Gln Gln Val Val Lys Phe Leu Ile Lys Lys Lys Ala Asn Leu
 225 230 235 240

Asn Ala Leu Asp Arg Tyr Gly Arg Thr Ala Leu Ile Leu Ala Val Cys
 245 250 255

Cys Gly Ser Ala Ser Ile Val Ser Pro Leu Leu Glu Gln Asn Val Asp
 260 265 270

Val Ser Ser Gln Asp Leu Glu Arg Arg Pro Glu Ser Met Leu Phe Leu
 275 280 285

Val Ile Ile Met
 290

<210> 316

<211> 584

<212> DNA

<213> Homo sapiens

<400> 316

atgtggccca aattccccctc cccctacagc ttgaagggga cataaccaat agcctggggt 60
 ttttttgtgg tccttggag atttctttgc ttattttctt ctgggtgggg gtgatttagag 120
 gaggcttatac actaatagga aggggagcta tagggaggct aggatatggg ggtaagctga 180
 gaggtcctcc tgtggatgt aaatttcaag ctttgcatacg tgtattctcc ttcaatgaaa 240
 agaaagcttg gacataaggt atttcactcc atttgccttc cctcttacag aaaaggtcaa 300
 gctgcaggat agtattgtaa tctgtacttc cctcaggtgg ccattttcc ccatcagaga 360
 gagaatgttg gggcaagcc atagtgcaga aaaaaaaatg agccacctct ttttcaggg 420
 tttgtgggtc aaatttgcattt cattggctta ggatgcattt caaaggtgag cctgttgatg 480
 cctgagtggtt tcccattctga aagacaaaac tgcccatggg tttggtttgt tttgtttctc 540
 cccctgccccca agaactatca aactcctgag ccaacaacta aaaa 584

<210> 317

<211> 829

<212> DNA

<213> Homo sapiens

<400> 317

attagcttcc gcttctgaca acactagaga tccctccccct ccctcaggggt atggccctcc 60
 acttcatttt tggcacataa catctttata ggacaggggt aaaaatccaa tactaacagg 120
 agaatgctta ggactctaacc aggttttga gaatgtgtt gtaaggggcca ctcaatccaa 180
 tttttcttgg tcctcattgt ggtcttaggag gacaggcaag ggtgcagatt ttcaagaatg 240
 catcagtaag ggccactaaa tccgacccctc ctcgttccctt cttgtgttctt gggaggaaaa 300
 ctatgtttc tggatgtgtt tcacttcc gatcagcagg gtccaggag 360
 cactgcaggat tcttggccag ggggagaaac aaaacaaacc aaaaccatgg gcagtttgc 420
 ctttcagatg ggaaacactc aggcatcaac aggctcacct ttgaaatgca tcctaagcca 480
 atgggacaaa ttgacccac aaaccctggaa aaaagaggtg gtcattttt tttgcactat 540
 ggcttggccc caacattctc tctctgttgg ggaaaaatgg ccacctgagg gaagtacaga 600
 ttacaataact atcctgcagg ttgacccat tctgttgggg gaaggcaaat ggagtgaaat 660
 accttatgttca aagctttctt tttcattgaa ggagaataca ctatgcaaag cttgaaattt 720
 acatcccaca ggaggacccctc tcagcttacc cccatatctt agcctccctt tagctccctt 780
 tccttattgtt gataaggccctc ctctaatcac cccccccccccag aagaaaataa 829

<210> 318
<211> 30
<212> PRT
<213> *Homo sapien*

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<400> 318
Thr Ala Ala Ser Asp Asn Phe Gln Leu Ser Gln Gly Gly Gln Gly Phe
1           5           10          15

Ala Ile Pro Ile Gly Gln Ala Met Ala Ile Ala Gly Gln Ile
                    20          25          30

```

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<210> 319  
<211> 41  
<212> DNA  
<213> Artificial Sequence
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<220>
<223> PCR primer

<400> 319

ggcctctgccc aatgggaact cagaagtagt aaaactccctgc 41

```
<210> 320
<211> 41
<212> DNA
<213> Artificial Sequence
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<220>
<223> PCR primer

<400> 320

gcaggagtt tactacttct gagttccat tggcagaggc c 41

```
<210> 321
<211> 60
<212> DNA
<213> Artificial Sequence
```

<220>
<223> PCR primer

321

gggaaattcc cgctggtgcc gcgcggcagc cctatggtgg ttgagggtga
ttccatgccc 500
600

<210> 322
<211> 42
<212> DNA
<213> Artificial Sequence

<220>
<223> PCR primer

<400> 322

cccgaaattct tatttatttc tggttcttga gacattttct gg 42

<210> 323
<211> 1590
<212> DNA
<213> Homo sapiens

<400> 323

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cagggattcg ccattccgat cgggcaggcg atggcgatcg cgggcccagat caagcttccc 120
accgttcata tcgggcctac cgcccttcctc ggcttggtg ttgtcgacaa caacggcaac 180
ggccgcacgag tccaacgcgt ggtcgggagc gctccggcgg caagtctcg catctccacc 240
ggcgcacgtga tcaccgcggt cgacggcgct ccgatcaact cgccaccgc gatggcggac 300
gcgcctaacg ggcacatcc cggtgcacgtc atctcggtga cctggcaaac caagtccggc 360
ggcacgcgta cagggAACGT gacattggcc gagggACCCC cggccgaatt cccgctggtg 420
ccgcgcggca gcccctatggt ggttggggtt gattccatgc cgctgcgttc ttctgtgaag 480
aaggcatttg gtctcaggag caagatggc aagtgggtct gccgttgctt cccctgctgc 540
aggagagagcg gcaagagcaa cgtgggact tctggagacc acgacgactc tgctatgaag 600
acactcagga gcaagatggg caagtgggtgc cgccactgct tcccctgctg cagggggagt 660
ggcaagagca acgtgggcgc ttctggagac cacgacgact ctgctatgaa gacactcagg 720
aacaagatgg gcaagtgggtg ctgccactgc ttcccctgct gcagggggag cggcaagagc 780
aagggtggcgc ctggggaga ctacgtgac agygcctca tgagggccag gtaccacgtc 840
cgtggagaag atctggacaa gctccacaga gctgcctggt ggggtaaagt ccccagaaaag 900
gatctcatcg tcatgctcag ggacactgac gtgaacaaga agacaagca aaagaggact 960
gctctacatc tggcctctgc caatggaat tcagaagtag taaaactcct gctggacaga 1020
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tgccaggaag atgaatgtgc gttaatgttg ctggAACATG gcactgatcc aaatattcca 1140
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aaagcactgc tcttatatgg tgctgatatac gaataaaaaa acaagcatgg cctcacacca 1260
ctgttacttg gtgtacatga gcaaaaacag caagtctgtca aatttttaat caagaaaaaa 1320
gcgaatttaa atgcactgga tagatatgga aggactgctc tcataacttgc tgtatgttgt 1380
ggatcagcaa gtatagtcag cttctactt gagcaaaaata ttgatgtatc ttctcaagat 1440
ctatctggac agacggccag agagtatgct gtttctagtc atcatcatgt aatttggccag 1500
ttacttctg actacaaga aaaacagatg ctaaaaatct cttctgaaaa cagcaatcca 1560
gaaaatgtct caagaaccag aaataaataaa 1590

<210> 324
<211> 529
<212> PRT
<213> Homo sapiens

<400> 324

Met His His His His His His Thr Ala Ala Ser Asp Asn Phe Gln Leu

5	10	15
Ser Gln Gly Gly Gln Gly Phe Ala Ile Pro Ile Gly Gln Ala Met Ala		
20	25	30
Ile Ala Gly Gln Ile Lys Leu Pro Thr Val His Ile Gly Pro Thr Ala		
35	40	45
Phe Leu Gly Leu Gly Val Val Asp Asn Asn Gly Asn Gly Ala Arg Val		
50	55	60
Gln Arg Val Val Gly Ser Ala Pro Ala Ala Ser Leu Gly Ile Ser Thr		
65	70	75
Gly Asp Val Ile Thr Ala Val Asp Gly Ala Pro Ile Asn Ser Ala Thr		
85	90	95
Ala Met Ala Asp Ala Leu Asn Gly His His Pro Gly Asp Val Ile Ser		
100	105	110
Val Thr Trp Gln Thr Lys Ser Gly Gly Thr Arg Thr Gly Asn Val Thr		
115	120	125
Leu Ala Glu Gly Pro Pro Ala Glu Phe Pro Leu Val Pro Arg Gly Ser		
130	135	140
Pro Met Val Val Glu Val Asp Ser Met Pro Ala Ala Ser Ser Val Lys		
145	150	155
160		
Lys Pro Phe Gly Leu Arg Ser Lys Met Gly Lys Trp Cys Cys Arg Cys		
165	170	175
Phe Pro Cys Cys Arg Glu Ser Gly Lys Ser Asn Val Gly Thr Ser Gly		
180	185	190
Asp His Asp Asp Ser Ala Met Lys Thr Leu Arg Ser Lys Met Gly Lys		
195	200	205
Trp Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Lys Ser Asn		
210	215	220
Val Gly Ala Ser Gly Asp His Asp Asp Ser Ala Met Lys Thr Leu Arg		
225	230	235
240		
Asn Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly		
245	250	255
Ser Gly Lys Ser Lys Val Gly Ala Trp Gly Asp Tyr Asp Ser Ala		
260	265	270
Phe Met Glu Pro Arg Tyr His Val Arg Gly Glu Asp Leu Asp Lys Leu		
275	280	285
His Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp Leu Ile Val		

290	295	300
Met Leu Arg Asp Thr Asp Val Asn Lys Lys Asp Lys Gln Lys Arg Thr		
305	310	315
320		
Ala Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu Val Val Lys Leu		
325	330	335
Leu Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg		
340	345	350
Thr Ala Leu Ile Lys Ala Val Gln Cys Gln Glu Asp Glu Cys Ala Leu		
355	360	365
Met Leu Leu Glu His Gly Thr Asp Pro Asn Ile Pro Asp Glu Tyr Gly		
370	375	380
Asn Thr Thr Leu His Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala		
385	390	395
400		
Lys Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys His		
405	410	415
Gly Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln Lys Gln Gln Val		
420	425	430
Val Lys Phe Leu Ile Lys Lys Ala Asn Leu Asn Ala Leu Asp Arg		
435	440	445
Tyr Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser		
450	455	460
Ile Val Ser Leu Leu Leu Glu Gln Asn Ile Asp Val Ser Ser Gln Asp		
465	470	475
480		
Leu Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser Ser His His His		
485	490	495
Val Ile Cys Gln Leu Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys		
500	505	510
Ile Ser Ser Glu Asn Ser Asn Pro Glu Asn Val Ser Arg Thr Arg Asn		
515	520	525
Lys		

<210> 325
<211> 1155
<212> DNA
<213> Homo sapiens

<400> 325
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aggagcaaga	tggccaagtg	gtgccaccac	cgcttcccc	gctgcagggg	gagcggcaag	120
agacaacatgg	gcacttctgg	agaccacgac	gactccctta	tgaagatgct	caggagcaag	180
atgggcaga	gttgcgc	ctgcttccc	tgctgcaggg	ggagcgcac	gagcaacgtg	240
ggcacttctg	gagaccatga	aaactcctt	atgaagatgc	tcaggagcaa	gatgggcaag	300
tggtgtctgc	actgcttccc	ctgctgcagg	gggagcggca	agagcaacgt	gggcgc	360
ggagactacg	accacagcgc	cttcatggag	ccgaggtacc	acatccgtc	agaagatctg	420
gacaagctcc	acagagctgc	ctgggtgggt	aaagtcccc	gaaaggatct	catcgtcatg	480
ctcagggaca	ctgacatgaa	caagagggac	aaggaaaaa	ggactgctct	acatttgcc	540
tctgccaatg	gaaattcaga	agtagtacaa	ctctgtctgg	acagacatg	tcaacttaat	600
gtccttgaca	acaaaaaaaag	gacagctctg	ataaaaggcc	tacaatgcca	ggaagatgaa	660
tgtgtgtta	tgttgctgga	acatggcgct	gatcgaaaata	ttccagatga	gtatggaaat	720
accgctctac	actatgctat	ctacaatgaa	gataaaattaa	tggccaaagc	actgctctta	780
tatggtgctg	atattgaatc	aaaaaacaag	gttggcctca	caccacttt	gcttggcgta	840
catgaacaaa	aacagcaagt	ggtgaaattt	ttaatcaaga	aaaaagctaa	tttaaatgta	900
cttgatagat	atgaaaggac	tgcctcata	cttgcgttat	gttgcgtatc	agcaagtata	960
gtcaatcttc	tacttgagca	aaatgttgat	gtatcttc	aagatctatc	tggacagacg	1020
gccagagagt	atgctgttcc	tagtcatcat	catgtaattt	gtgaattact	ttctgactat	1080
aaagaaaaaac	agatgctaaa	aatctttct	gaaaacagca	atccagaaaa	tgtctcaaga	1140
accagaaata	aataa					1155

<210> 326

<211> 384

<212> PRT

<213> Homo sapiens

<400> 326

Met Val Ala Glu Val Cys Ser Met Pro Thr Ala Ser Thr Val Lys Lys
5 10 15

Pro Phe Asp Leu Arg Ser Lys Met Gly Lys Trp Cys His His Arg Phe
20 25 30

His Asp Asp Ser Phe Met Lys Met Leu Arg Ser Lys Met Gly Lys Cys
50 55 60

Cys Arg His Cys Phe Pro Cys Cys Arg Gly Ser Gly Thr Ser Asn Val
65 70 75 80

Gly Thr Ser Gly Asp His Glu Asn Ser Phe Met Lys Met Leu Arg Ser
85 90 95

Lys Met Gly Lys Trp Cys Cys His Cys Phe Pro Cys Cys Arg Gly Ser
 100 105 110

Gly Lys Ser Asn Val Gly Ala Trp Gly Asp Tyr Asp His Ser Ala Phe
115 120 125

Met Glu Pro Arg Tyr His Ile Arg Arg Glu Asp Leu Asp Lys Leu His
 130 135 140

Arg Ala Ala Trp Trp Gly Lys Val Pro Arg Lys Asp Leu Ile Val Met

145	150	155	160
Leu Arg Asp Thr Asp Met Asn Lys Arg Asp Lys Glu Lys Arg Thr Ala			
165	170	175	
Leu His Leu Ala Ser Ala Asn Gly Asn Ser Glu Val Val Gln Leu Leu			
180	185	190	
Leu Asp Arg Arg Cys Gln Leu Asn Val Leu Asp Asn Lys Lys Arg Thr			
195	200	205	
Ala Leu Ile Lys Ala Ile Gln Cys Gln Glu Asp Glu Cys Val Leu Met			
210	215	220	
Leu Leu Glu His Gly Ala Asp Arg Asn Ile Pro Asp Glu Tyr Gly Asn			
225	230	235	240
Thr Ala Leu His Tyr Ala Ile Tyr Asn Glu Asp Lys Leu Met Ala Lys			
245	250	255	
Ala Leu Leu Leu Tyr Gly Ala Asp Ile Glu Ser Lys Asn Lys Val Gly			
260	265	270	
Leu Thr Pro Leu Leu Leu Gly Val His Glu Gln Lys Gln Gln Val Val			
275	280	285	
Lys Phe Leu Ile Lys Lys Ala Asn Leu Asn Val Leu Asp Arg Tyr			
290	295	300	
Gly Arg Thr Ala Leu Ile Leu Ala Val Cys Cys Gly Ser Ala Ser Ile			
305	310	315	320
Val Asn Leu Leu Leu Glu Gln Asn Val Asp Val Ser Ser Gln Asp Leu			
325	330	335	
Ser Gly Gln Thr Ala Arg Glu Tyr Ala Val Ser Ser His His His Val			
340	345	350	
Ile Cys Glu Leu Leu Ser Asp Tyr Lys Glu Lys Gln Met Leu Lys Ile			
355	360	365	
Ser Ser Glu Asn Ser Asn Pro Glu Asn Val Ser Arg Thr Arg Asn Lys			
370	375	380	